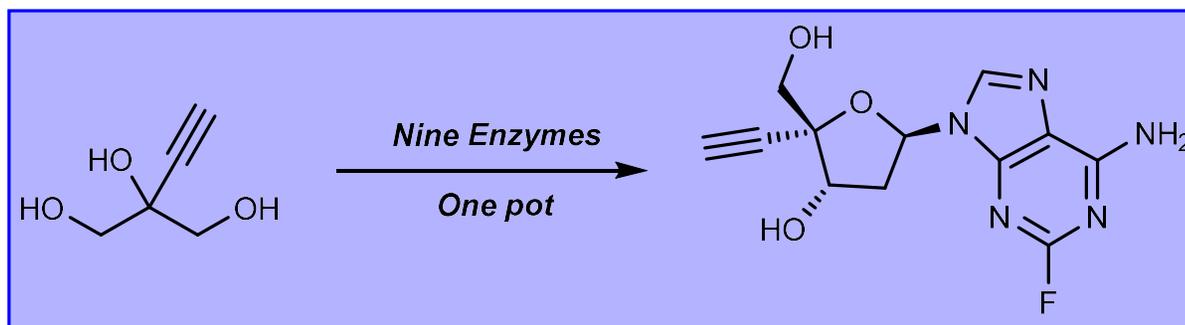


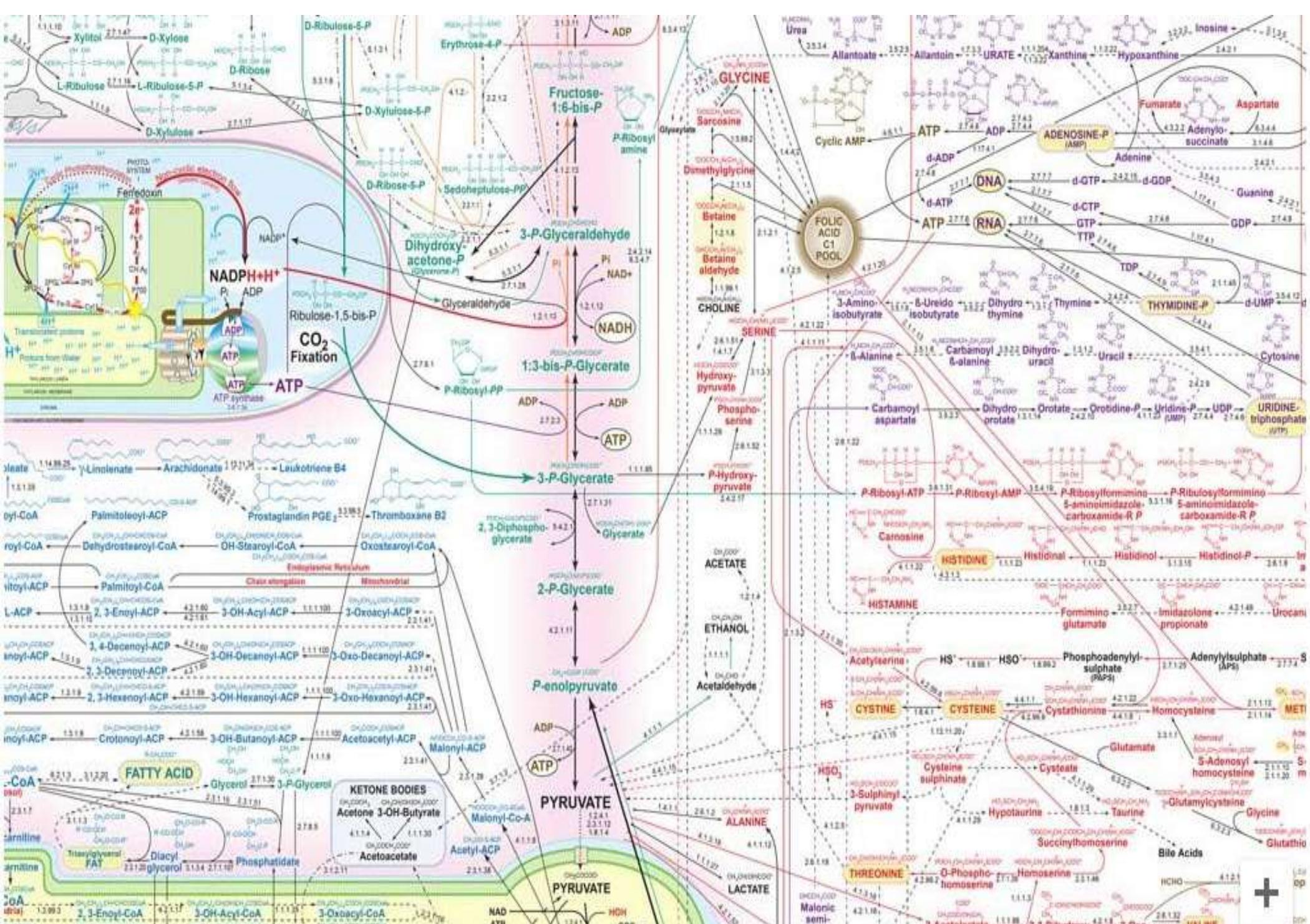
Recent Advances in Biocatalytic Cascades:

Matt Bock

Denmark Group Meeting

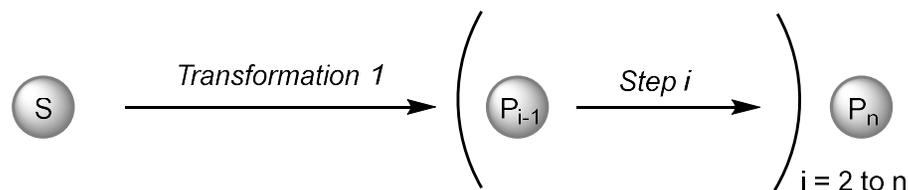
23 November 2021



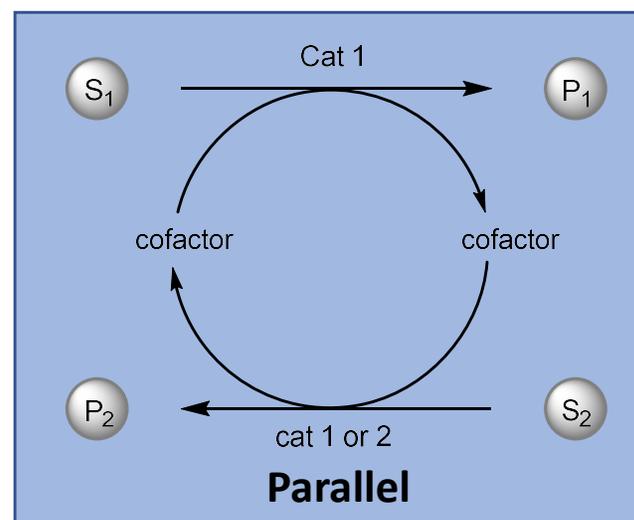
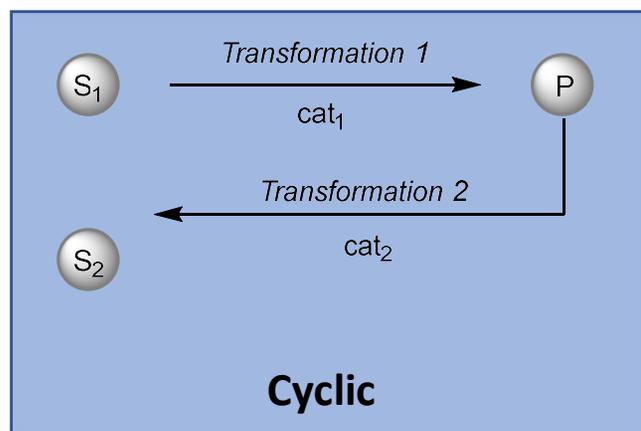
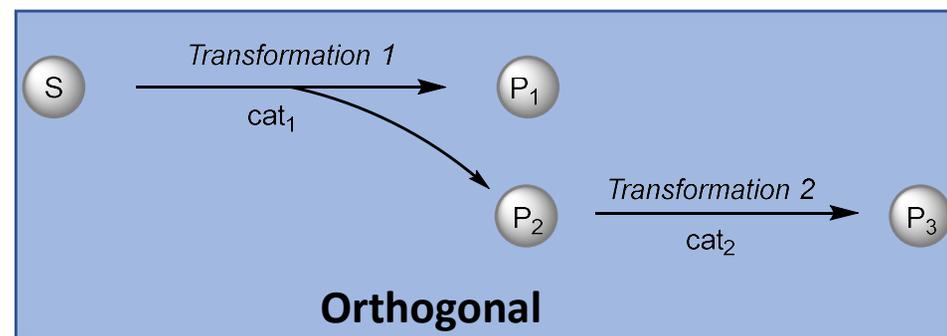
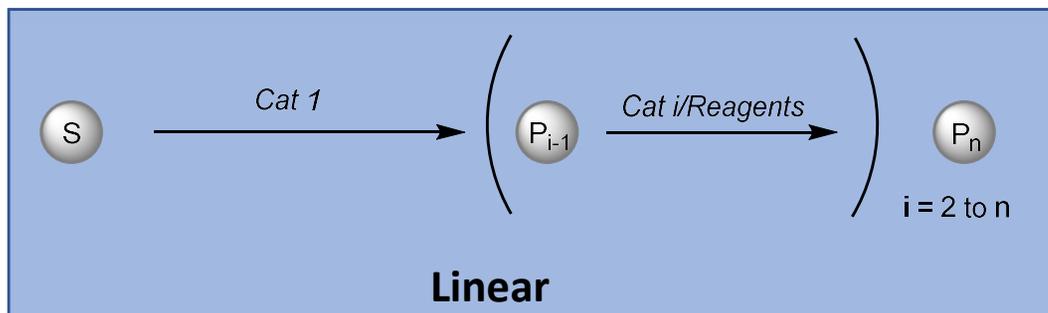


What is a biocatalytic cascade?

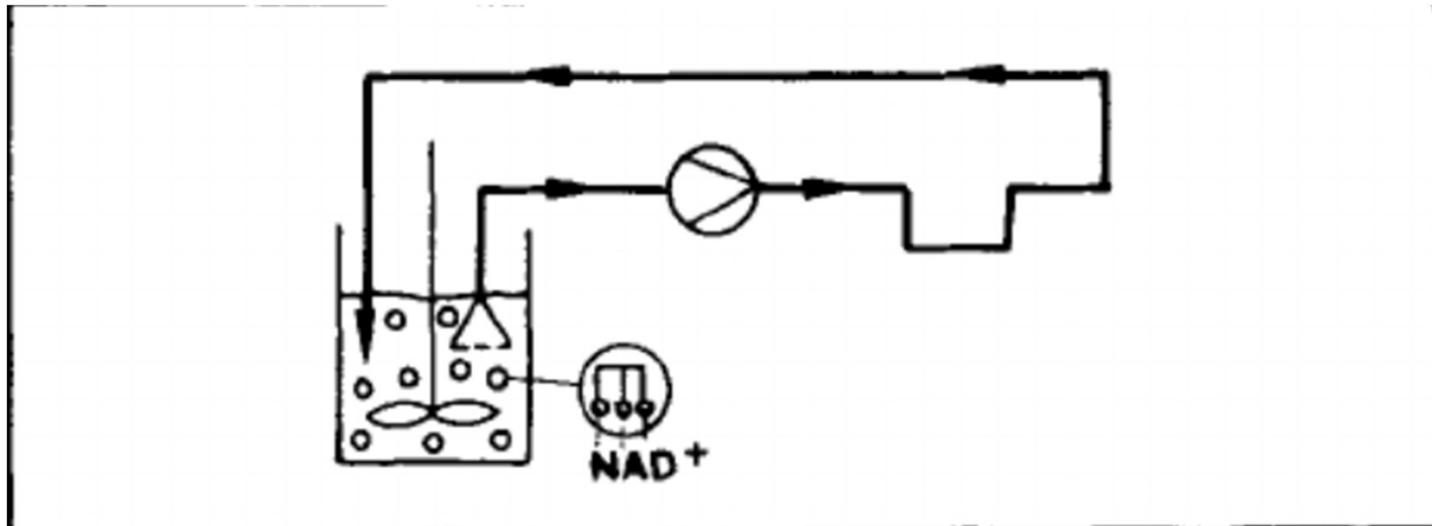
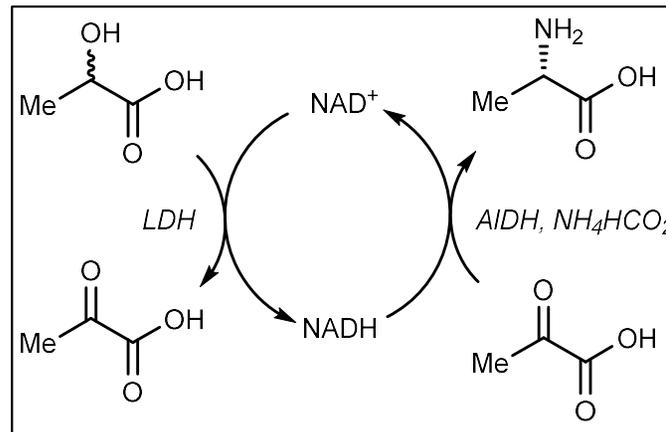
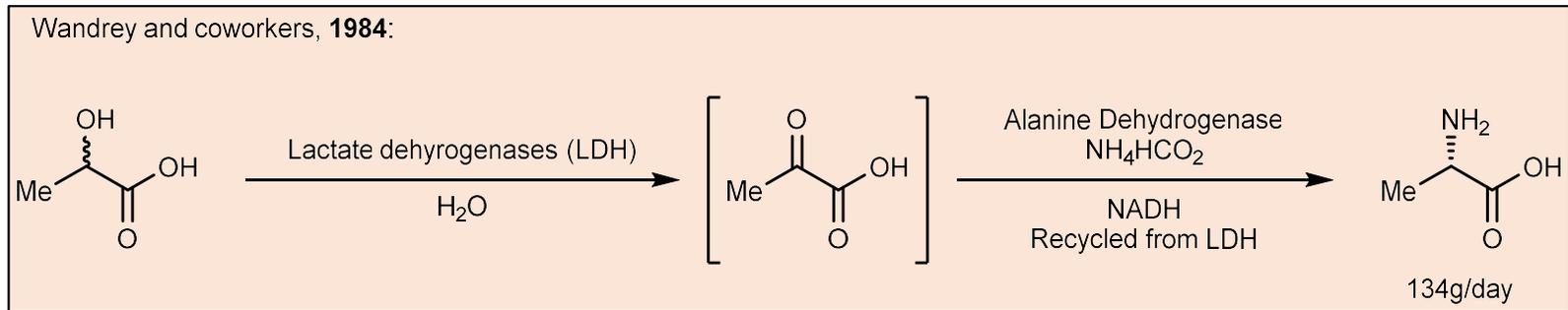
A reaction comprising of two or more transformations in the same pot where one step includes a biocatalyst



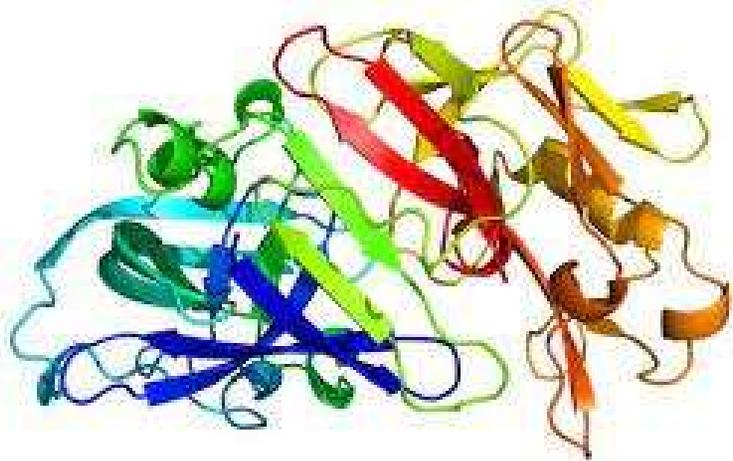
Classifications:



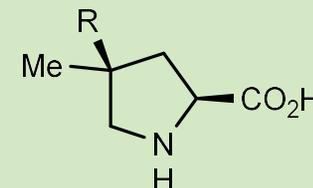
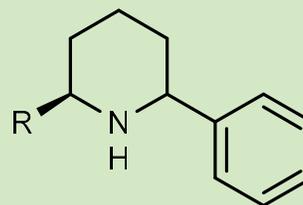
The First *in vitro*, Artificial Biocatalytic Cascade



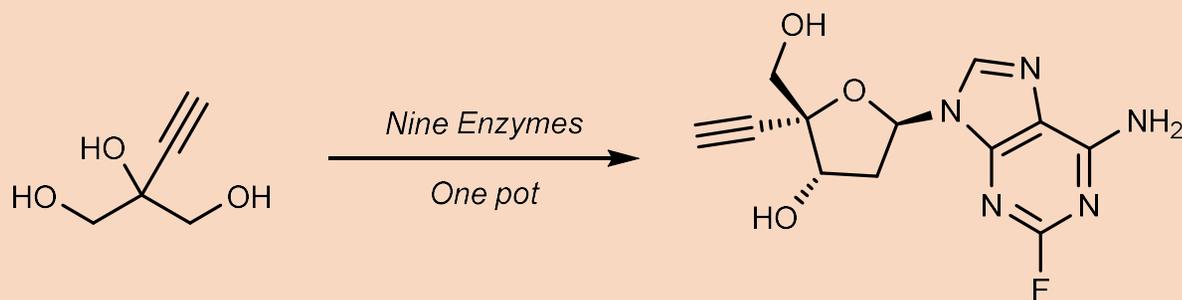
Outline



Enzyme classifications



Biocatalytic cascades for the synthesis of nitrogen heterocycles



CODEXIS®

Pharmaceuticals prepared by biocatalytic cascades

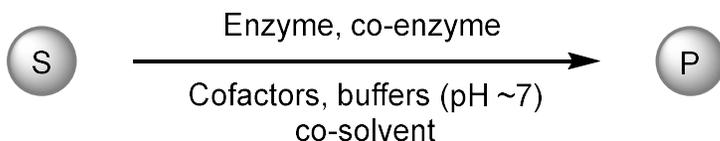
Enzyme Classification

IUPAC's Classification of Enzymes

Class	Reaction Type	Enzyme Examples
Oxidoreductase	Redox Chemistry	Ketoreductases, Iminoreductases, Oxidases
Transferase	FG Addition or transfer	Aminotrasferases, Kinases
Hydrolase	Hydrolytic reactions	Nitrilase, Lipase, Amidase
Lyase	Eliminations	Aldolases, Decarboxylases
Isomerase	Isomerizations	Epimerases
Ligase	Coupling reactions often macromolecules	C-C, C-N, C-O, C-S Ligases

What's in a Biocatalytic reaction?

Generalized reaction scheme



Cofactors: “reagents”

Co-enzyme: Regenerate cofactors or remove byproducts

Buffers: Aqueous. Mixture of salts, system specific

Co-solvent: DMSO, IPA, THF, etc.

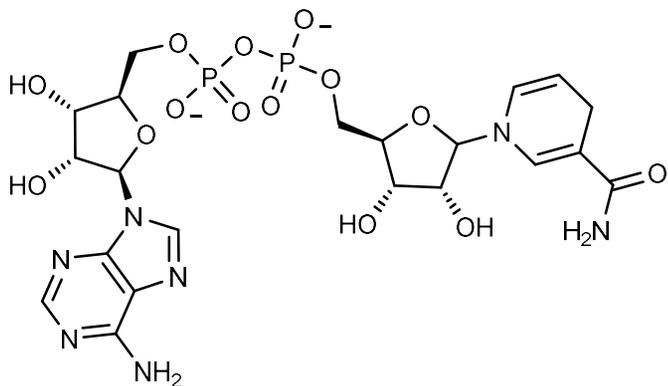
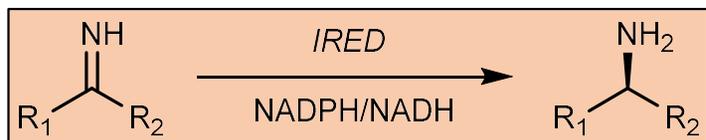
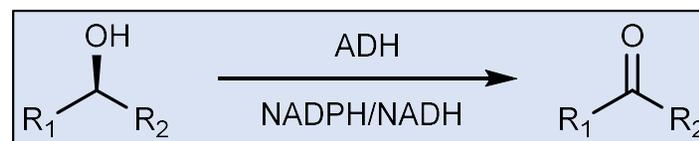
Temperatures: room temp or slightly elevated (35 °C)

A significant number of these reactions are reversible!

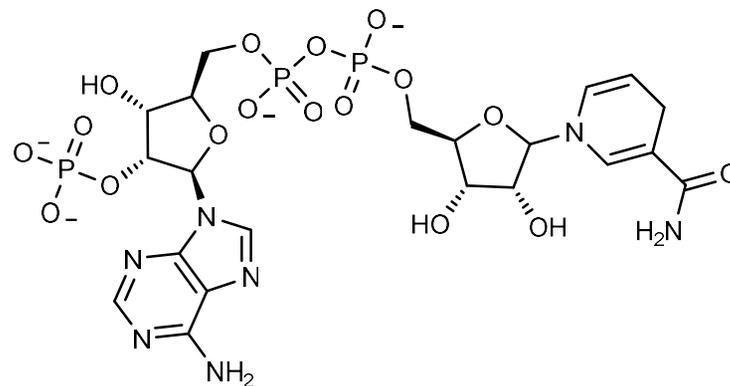
For the purpose of this talk, specific enzymes are not named, just listed as a general transform

Oxidoreductases

Generalized schemes:

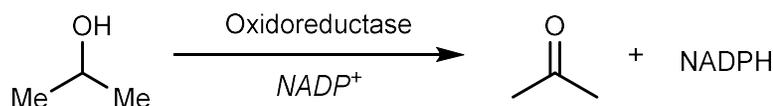
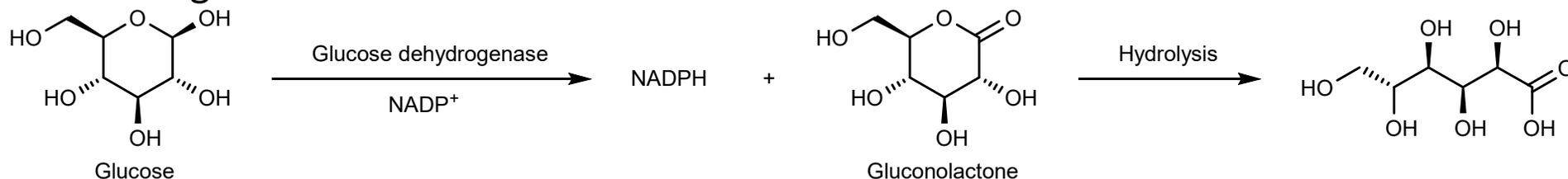


NADH



NADPH

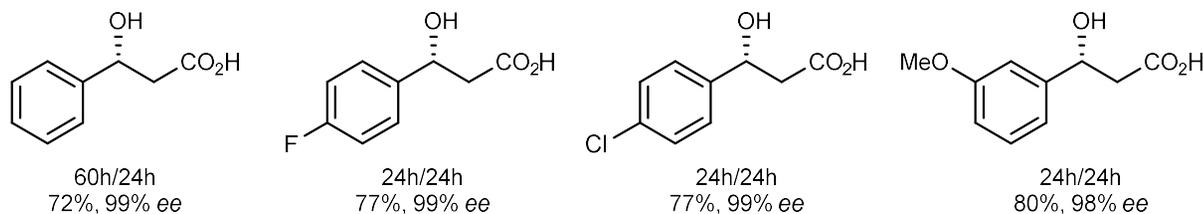
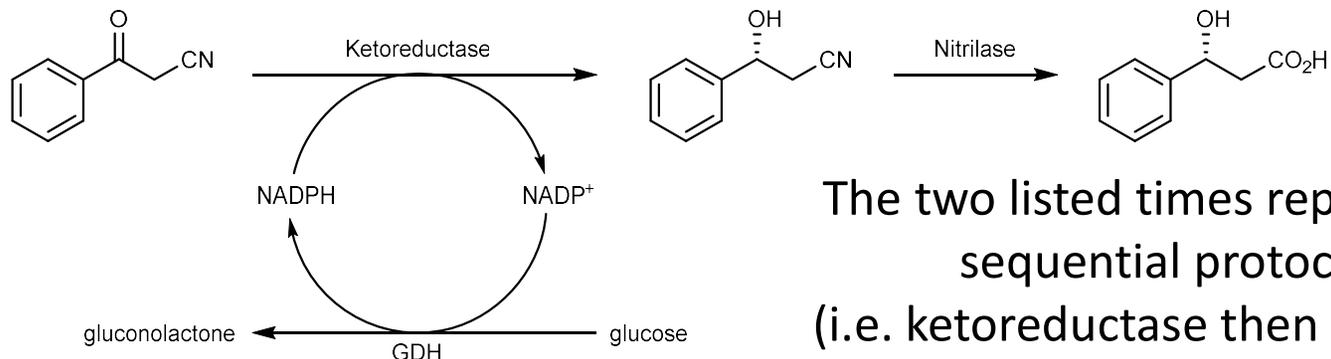
Cofactor regeneration:



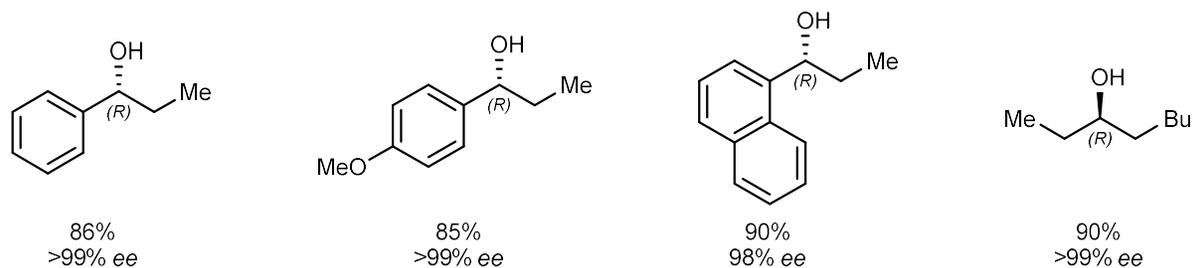
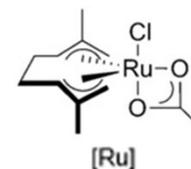
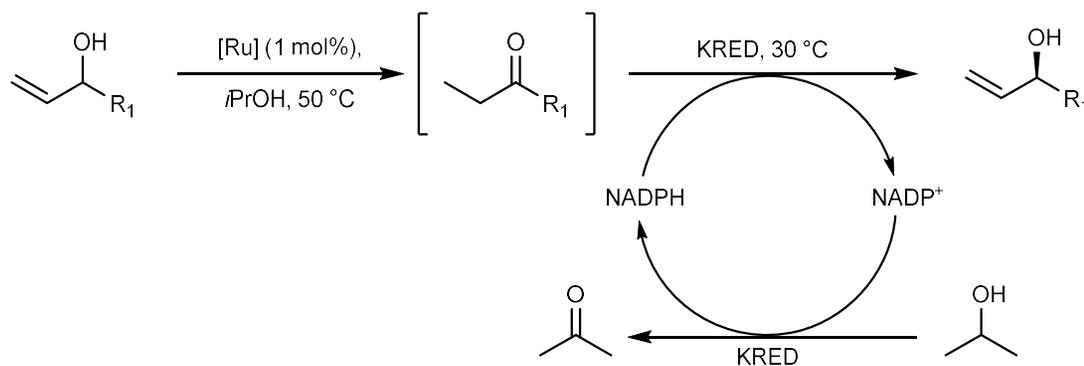
While not as mature, ketoreductases and iminoreductases are starting to see use 7

Examples of KREDs

Synthesis of β -hydroxy acids

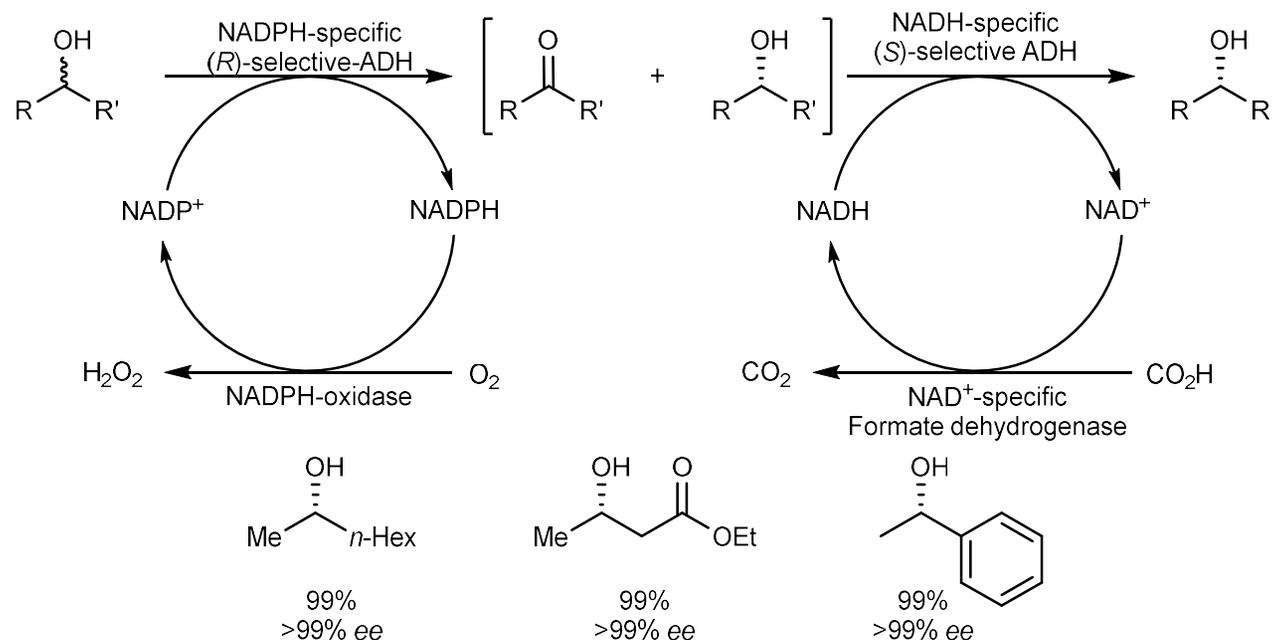


Synthesis of benzylic alcohols

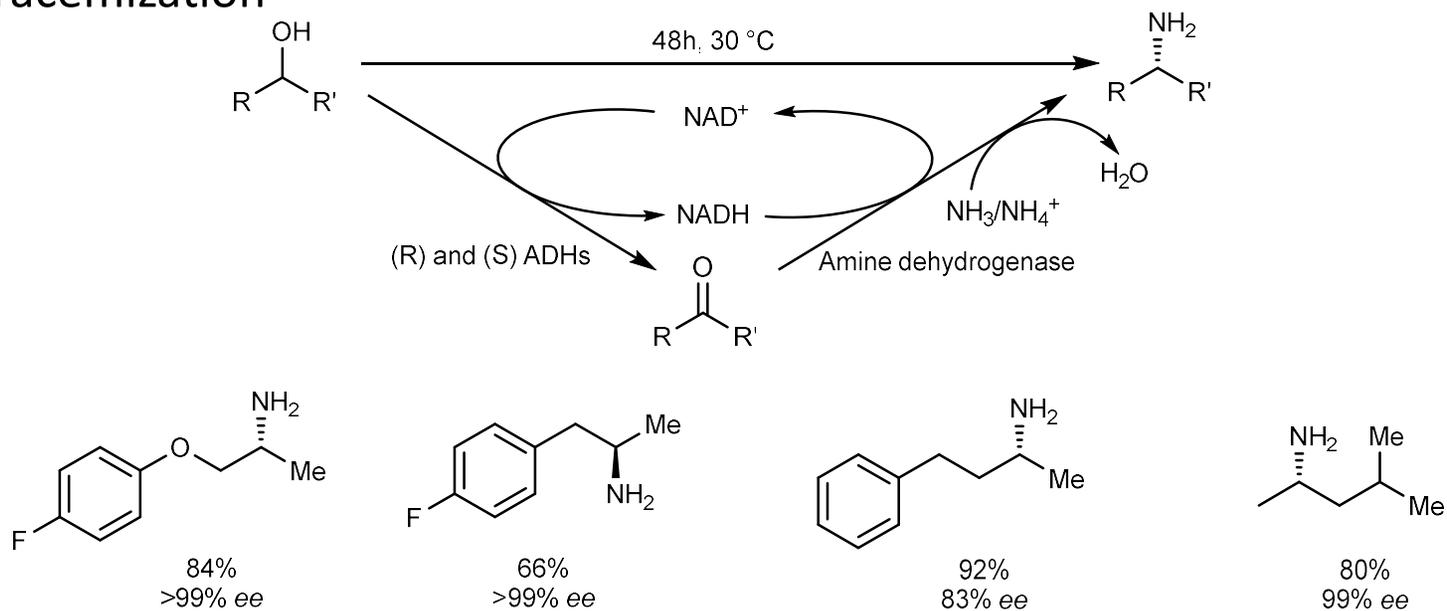


Examples of ADHs

Deracemization with two ADHs

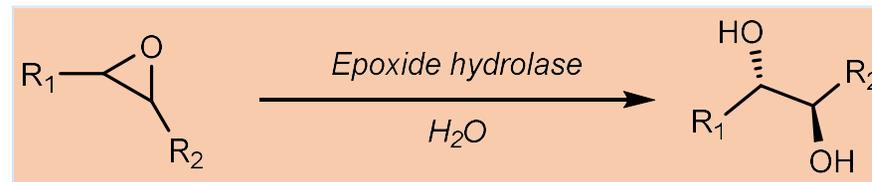
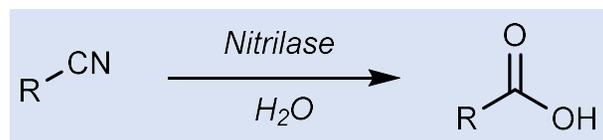
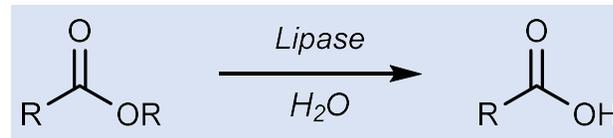
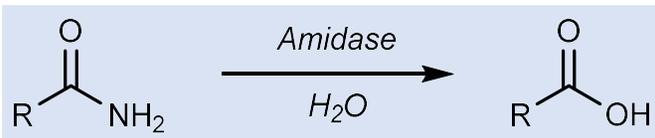


Another deracemization

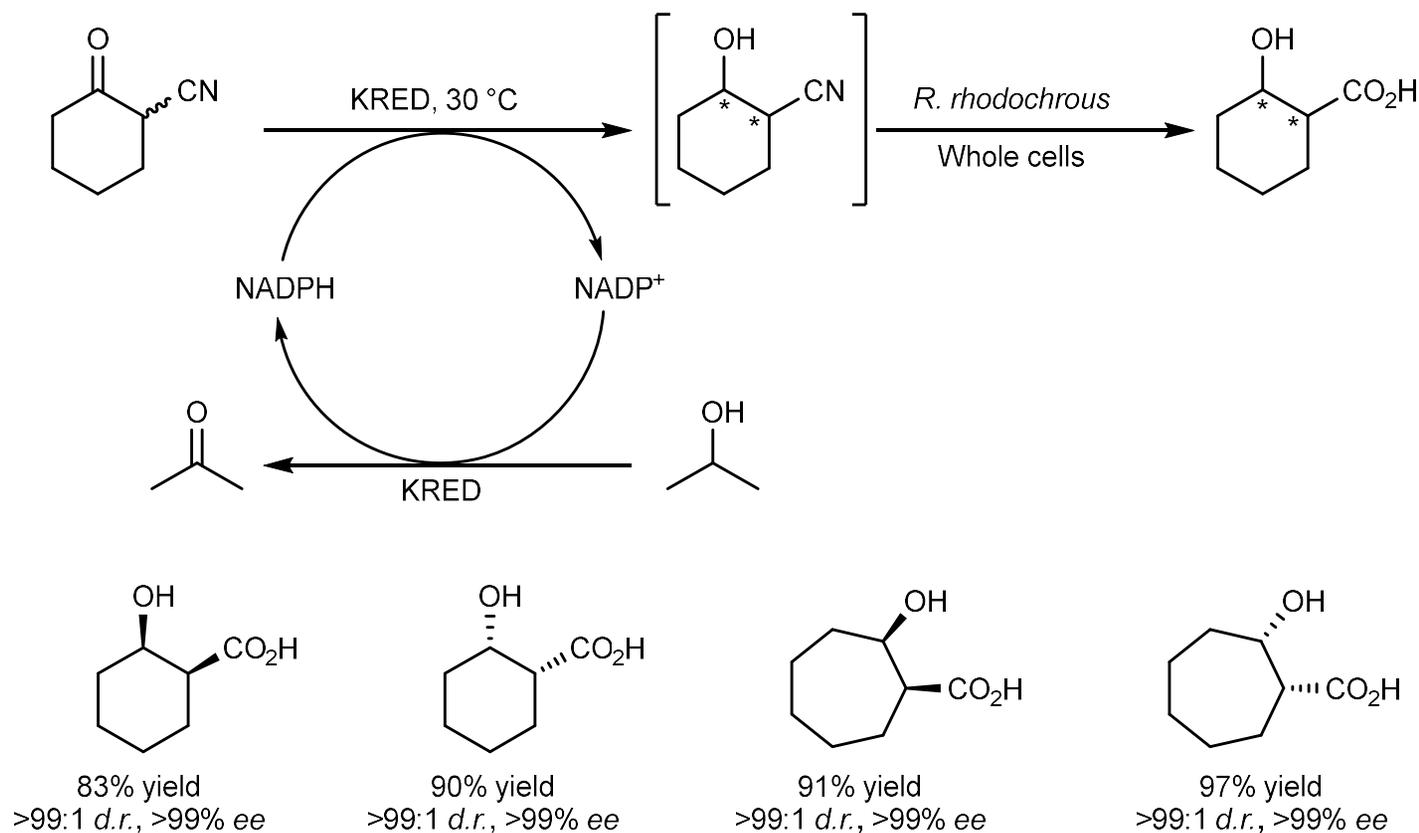


Hydrolases

Generalized schemes:

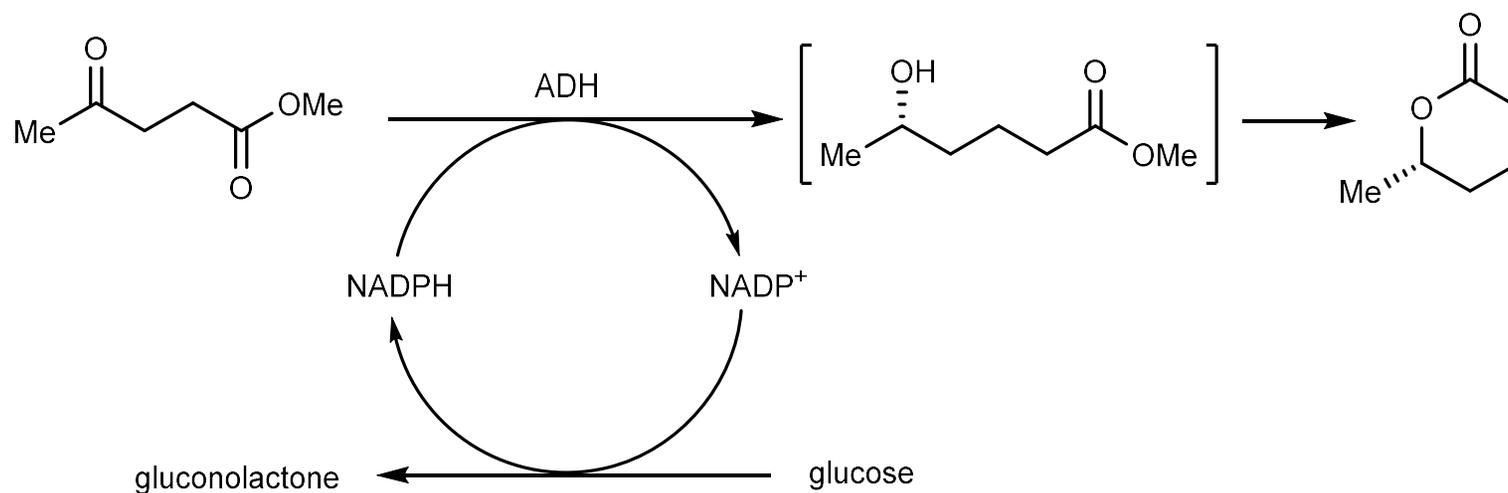
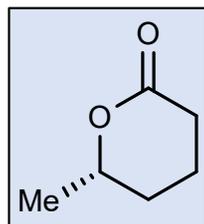


Synthesis of cyclic acids



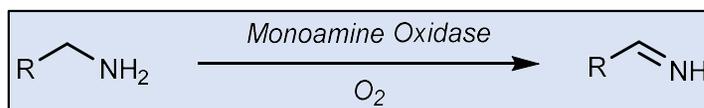
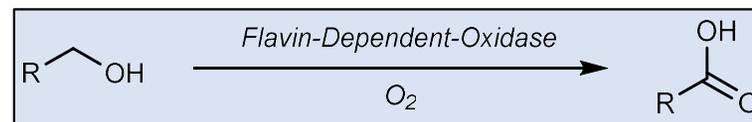
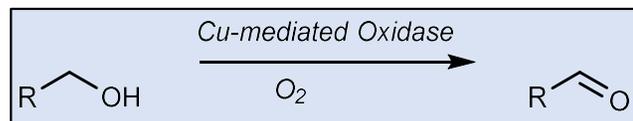
Group Problem #1

Using a biocatalytic cascade, prepare the following compound

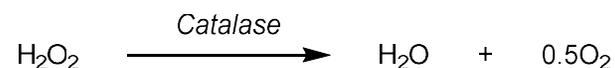


Oxidases

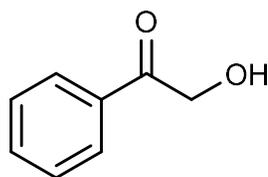
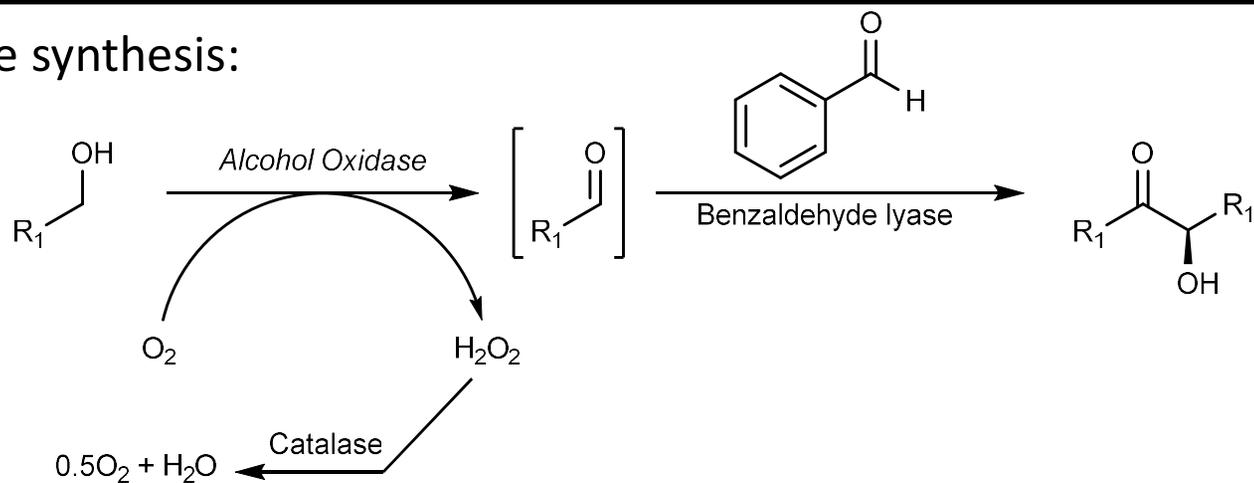
Generalized schemes:



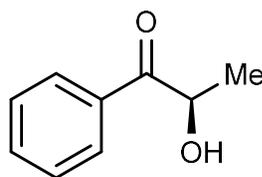
- Often very substrate specific.
- Normally paired with one or two co-enzymes
 - Catalase for breakdown of H_2O_2
 - A peroxidase to maintain Cu oxidation state



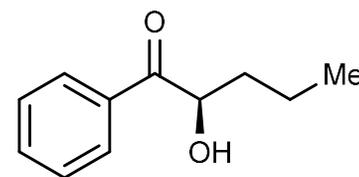
α -hydroxy ketone synthesis:



> 99% Conv.



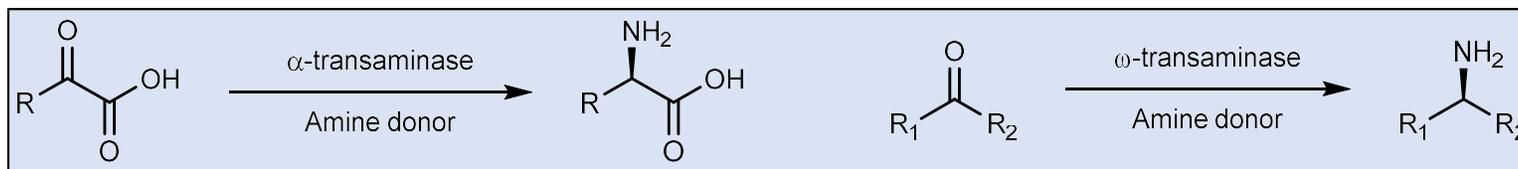
> 99% Conv.
99% ee



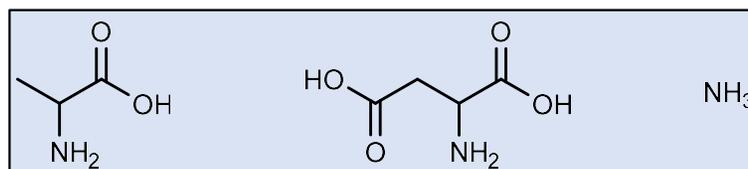
> 91% Conv.
98% ee

Transaminases

Generalized scheme:



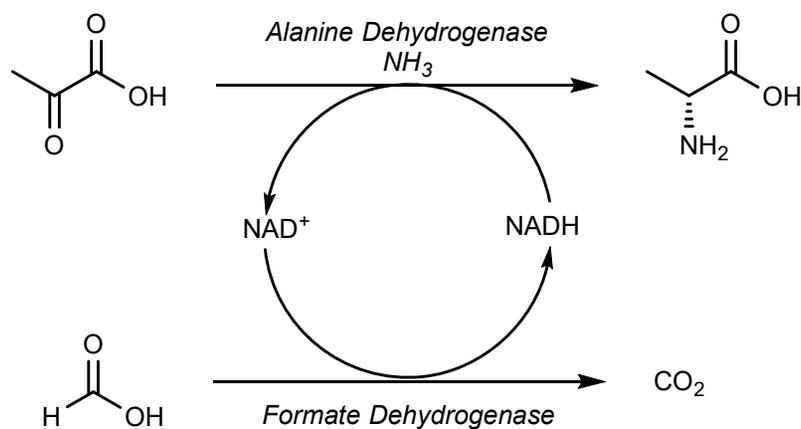
Common Amine donors



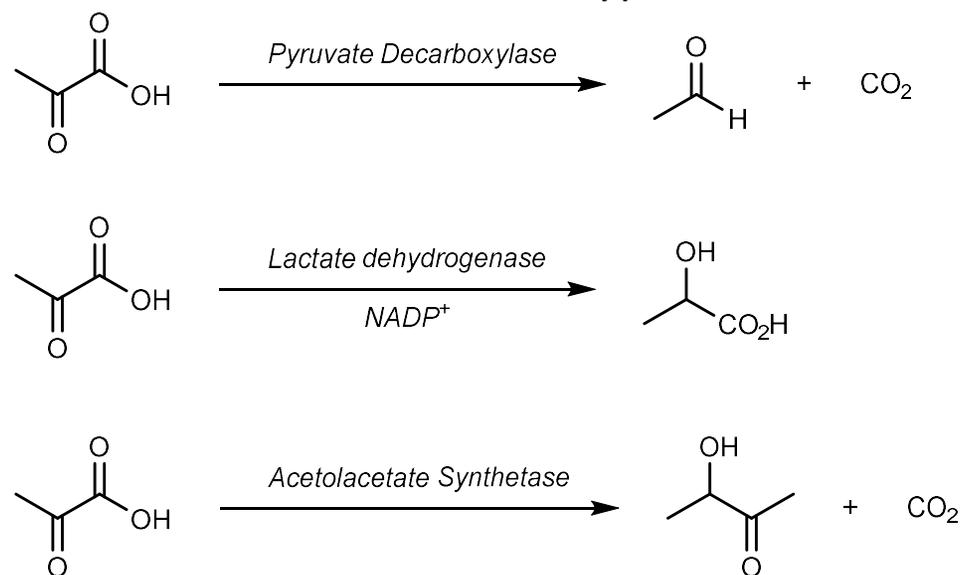
While a powerful reaction, equilibrium is favored for starting material.

Strategies to Shift Equilibrium:

Regeneration of amine donor

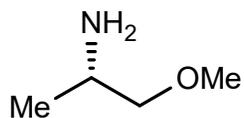
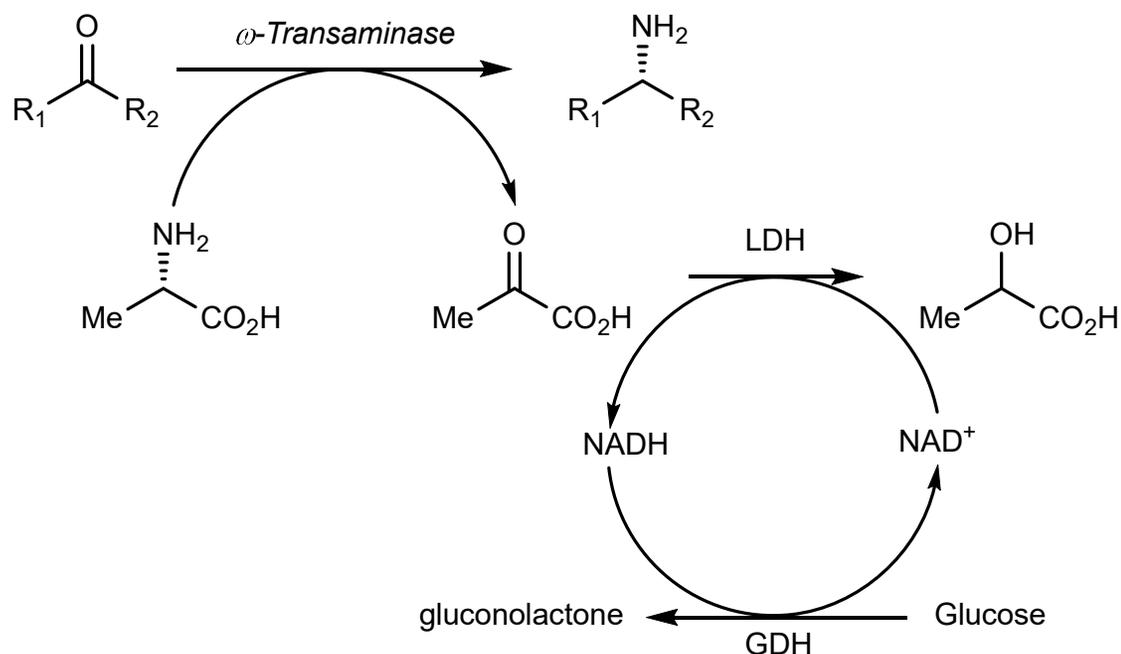


Removal of keto-acid byproduct

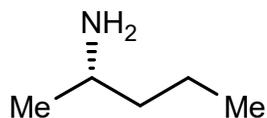


Transaminases

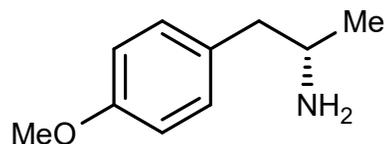
Asymmetric reductive amination



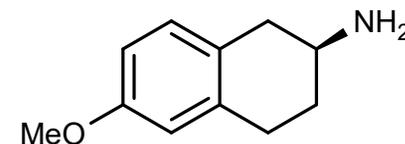
97% Conv.
>99% ee



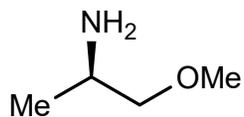
97% Conv.
>99% ee



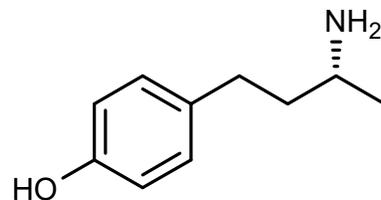
94% Conv.
>99% ee



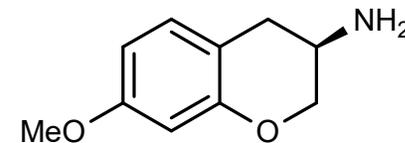
99% Conv.
>99% ee



99% Conv.
>99% ee



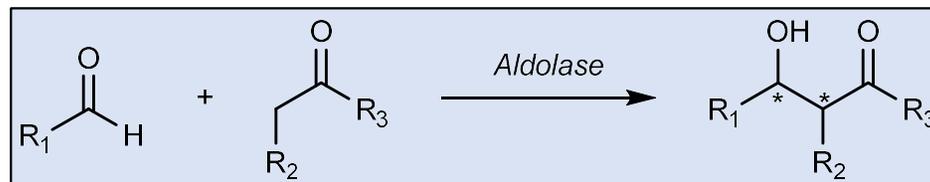
99% Conv.
>99% ee



99% Conv.
>99% ee

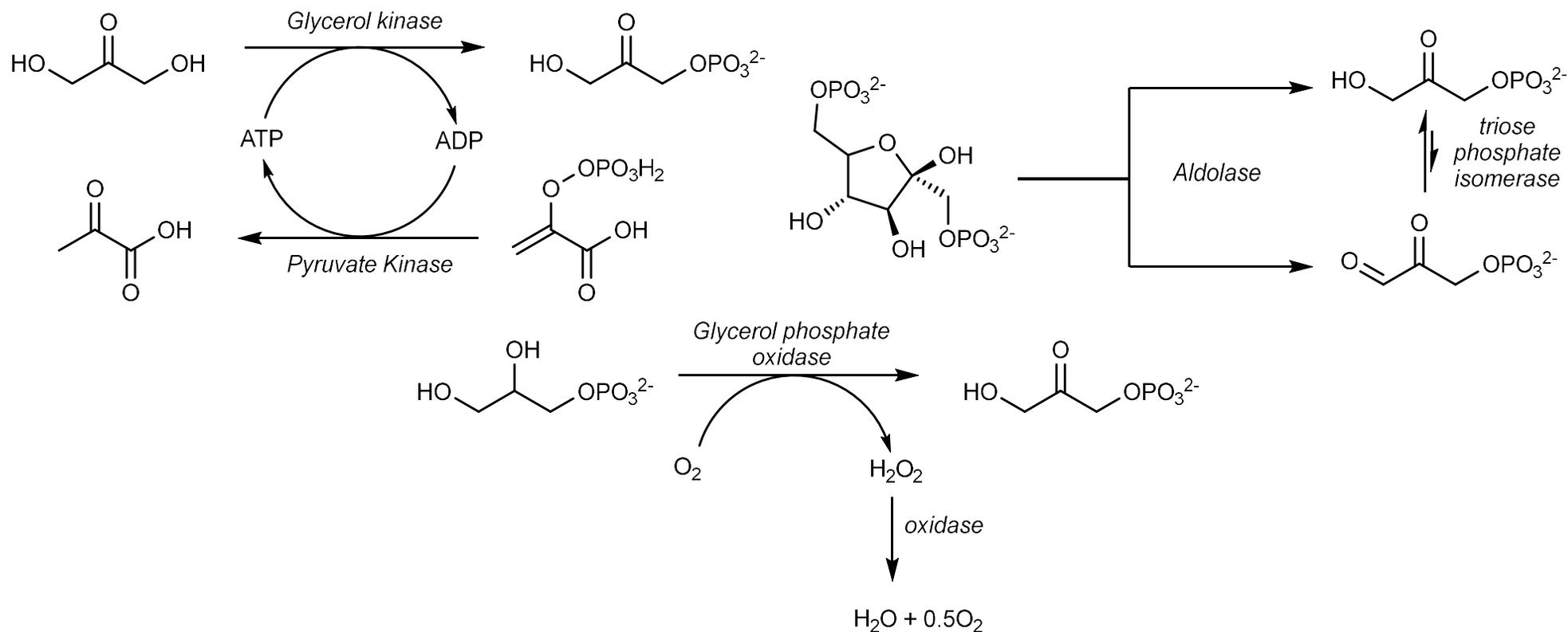
Aldolases

Generalized schemes:

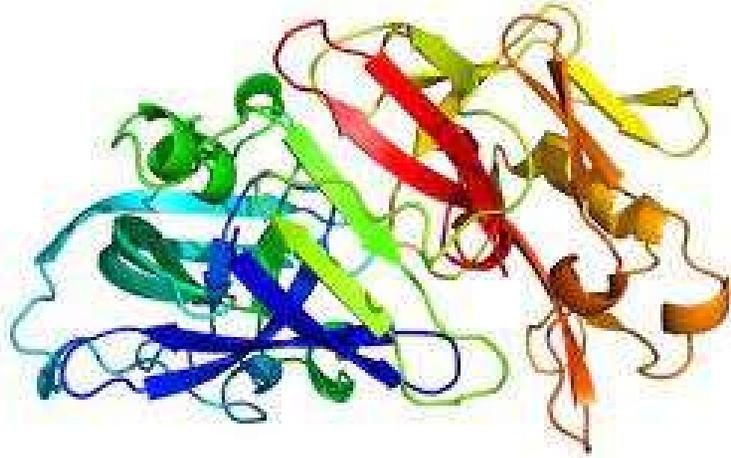


- Range of acceptors is typically broad
- Donors are typically limited to dihydroxyacetone phosphate (DHAP) derivatives

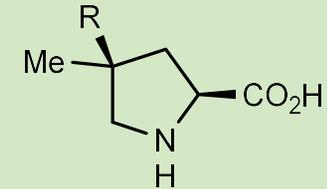
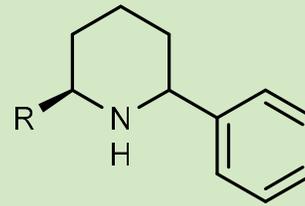
In Situ Preparation of DHAP:



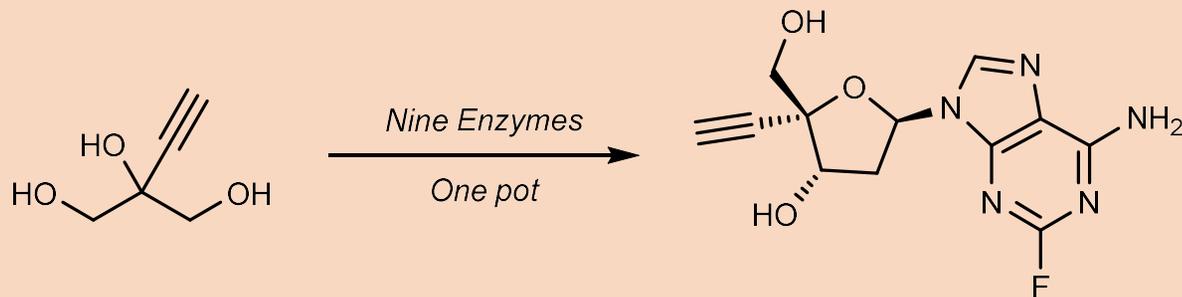
Outline



Enzyme classifications



Biocatalytic cascades for the synthesis of nitrogen heterocycles



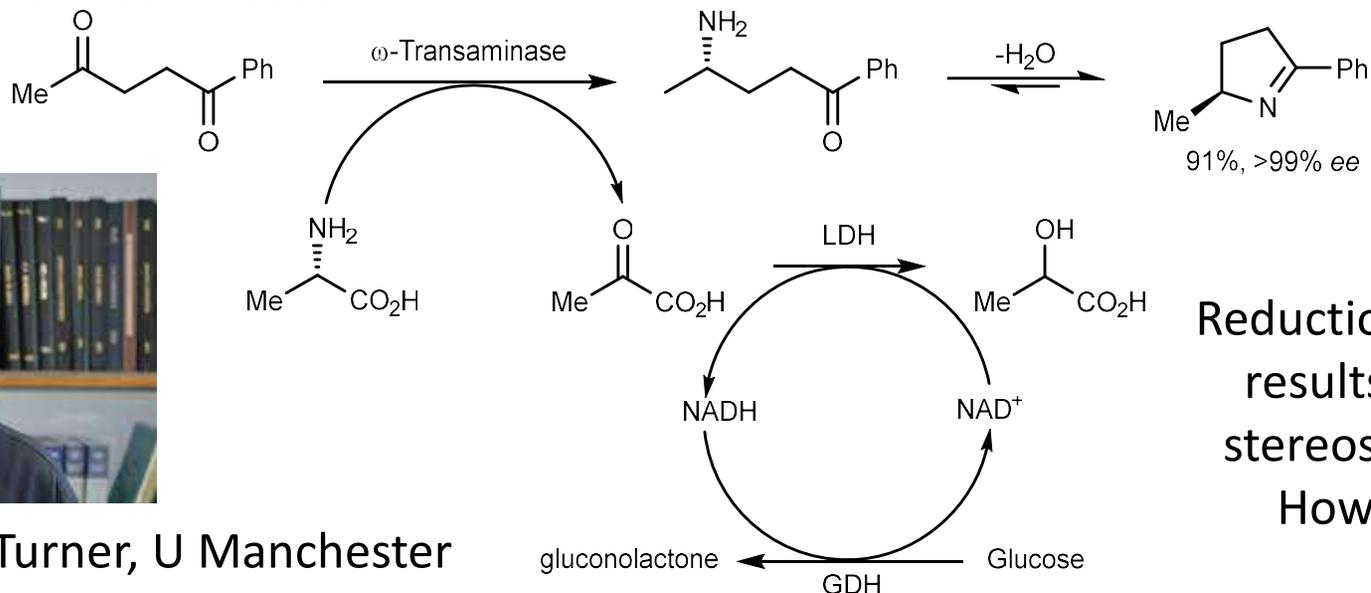
MERCK
INVENTING FOR LIFE

CODEXIS®

Pharmaceuticals prepared by biocatalytic cascades

Preparation of Disubstituted Pyrrolidines

Cyclization with transaminases:

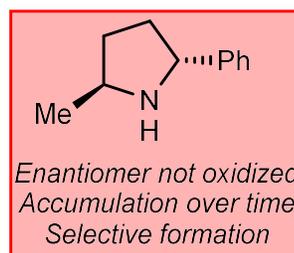
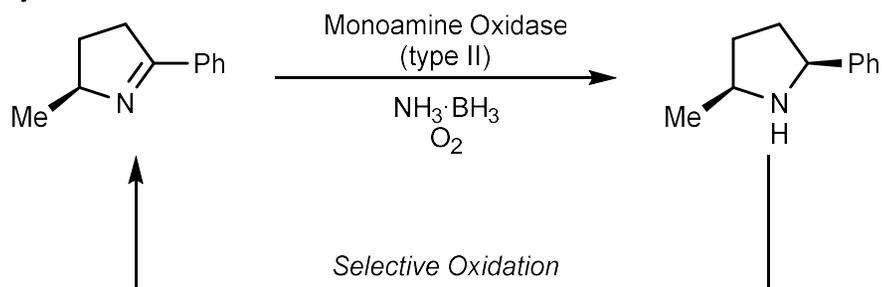


Reduction of imine
results in poor
stereoselectivity
How to fix?



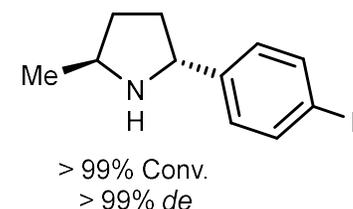
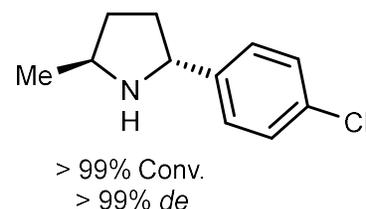
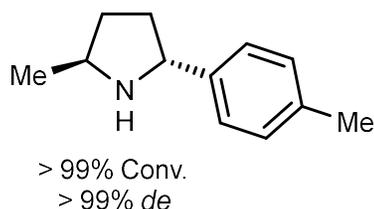
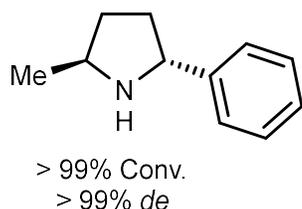
Prof. Nicholas Turner, U Manchester

Cyclization with transaminases:



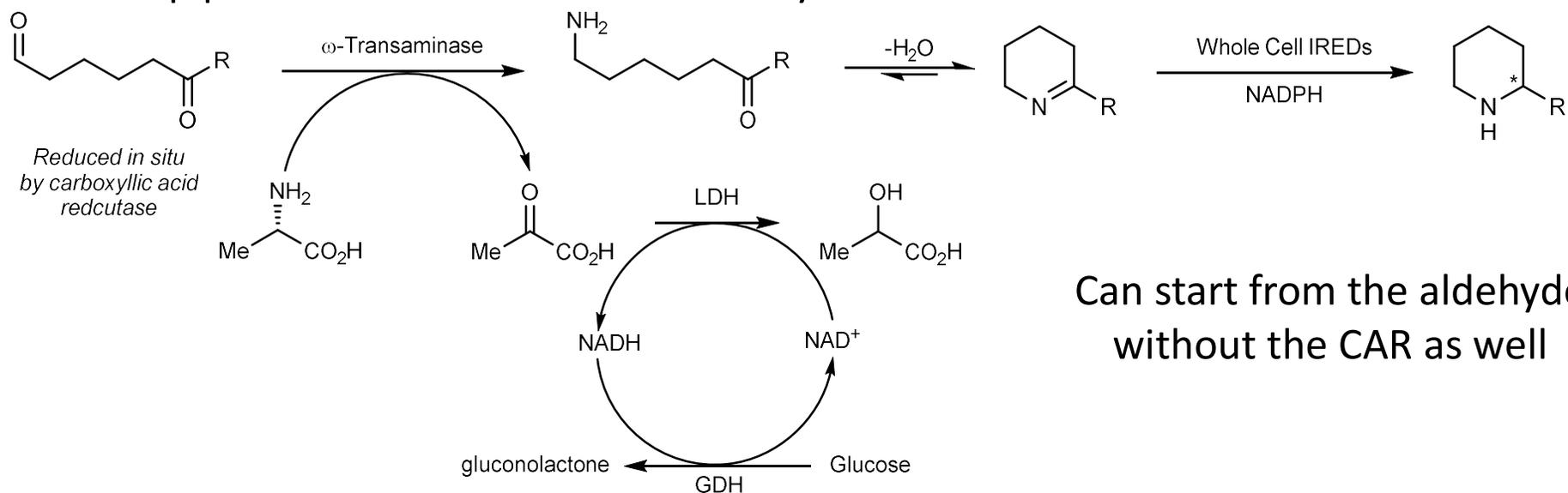
Strategy used previously
to prepare monosubstituted
pyrrolidines

Substrates prepared in one-pot:

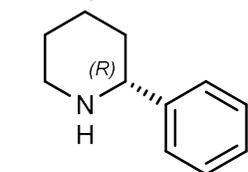


Synthesis of Substituted Piperidines

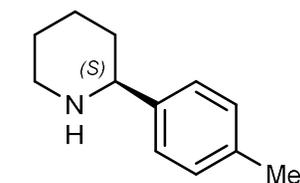
Access to piperidines *via* IREDs and a carboxylic acid reductase:



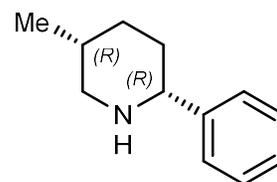
Selected Examples:



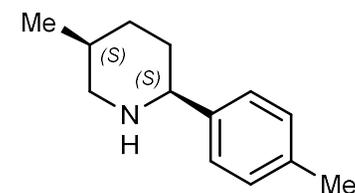
> 98% Conv.
93% e.e.
CAR, ATA, (S)-IRED



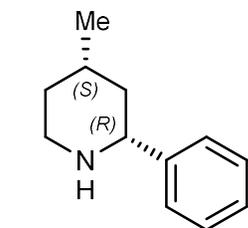
> 98% Conv.
98% e.e.
CAR, ATA, (R)-IRED



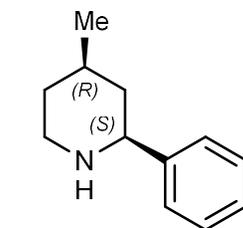
95% Conv.
>98% e.e., >98% d.e.
ATA, (S)-IRED



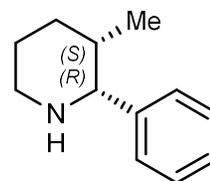
96% Conv.
>98% e.e., 29% d.e.
ATA, (R)-IRED



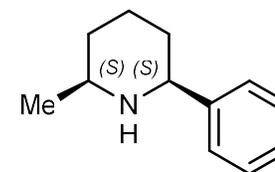
98% Conv.
>98% e.e., 95% d.e.
CAR, ATA, (S)-IRED



74% Conv.
>98% e.e., 92% d.e.
CAR, ATA, (R)-IRED



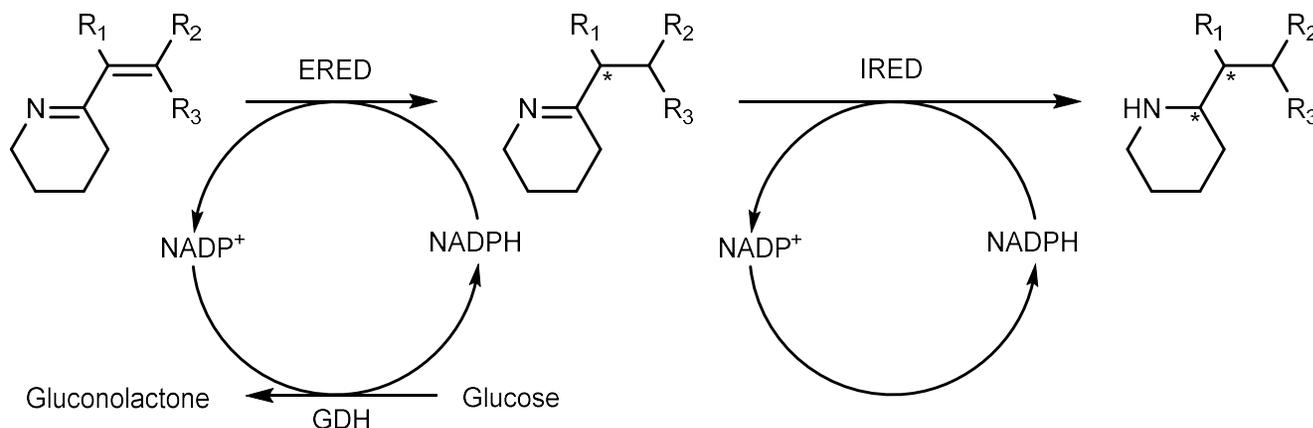
89% Conv.
81% e.e., 81% d.e.
CAR, ATA, (S)-IRED



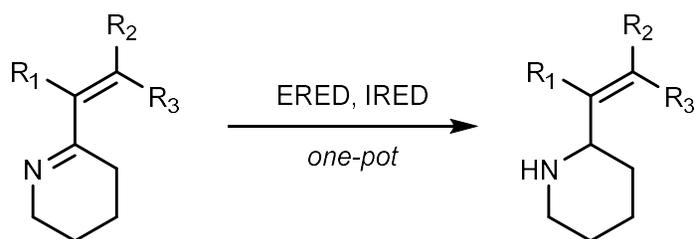
81% Conv.
98% e.e., 98% d.e.
ATA, (R)-IRED
Sequential Protocol

Further Substitution

Contiguous stereocenters:

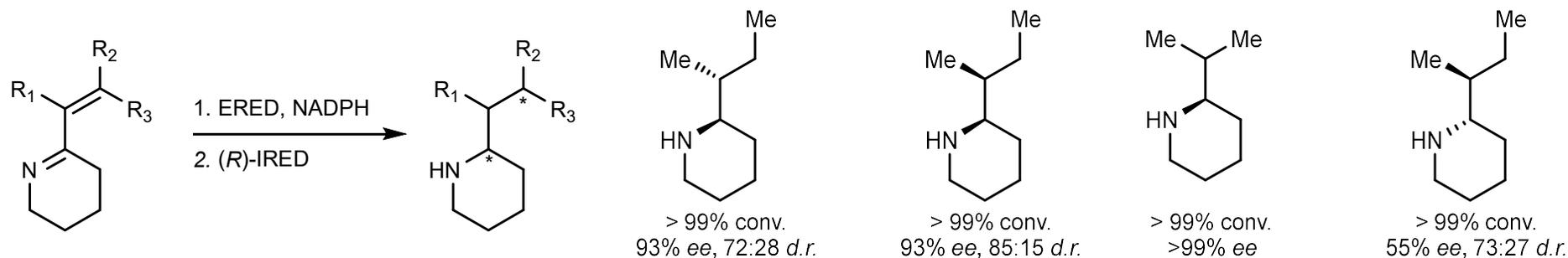


Challenge:
rate of competing
reductions



Allylic amines are not
acceptors for EREDS

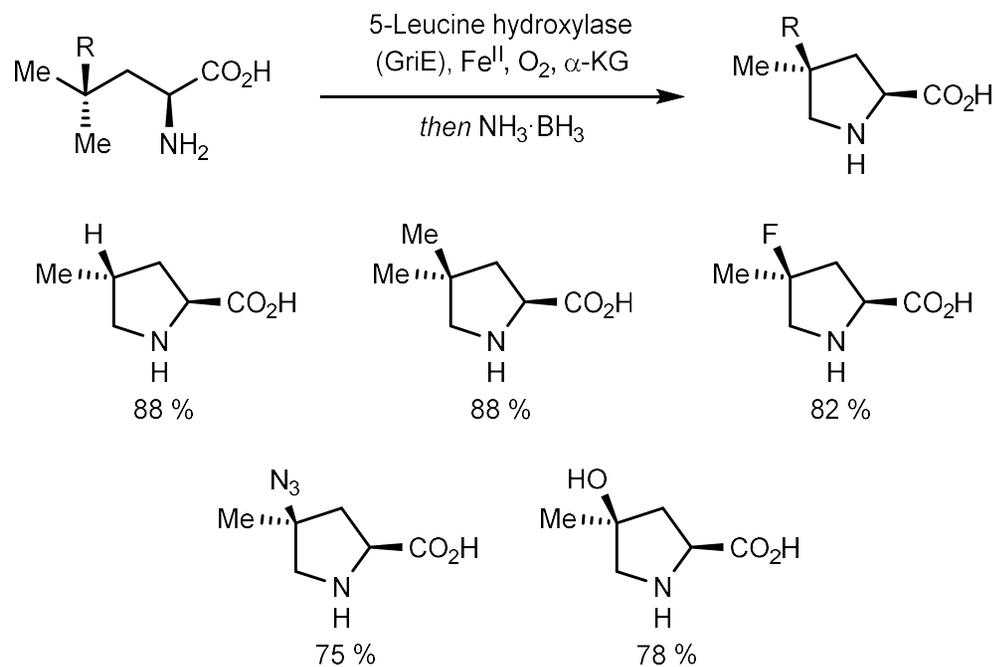
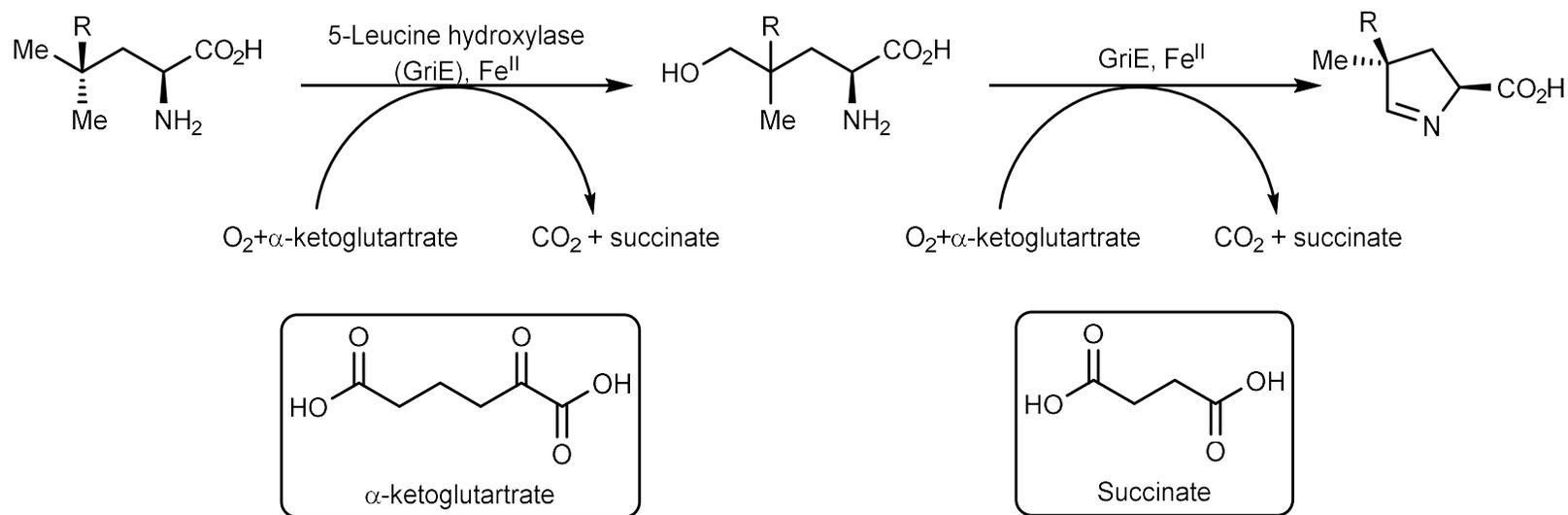
Sequential Protocol:



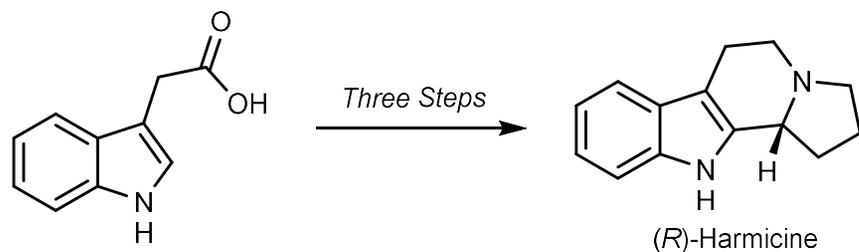
5 and 7 membered rings also reported,

Synthesis of Substituted Prolines

Leucine-hydroxylases:

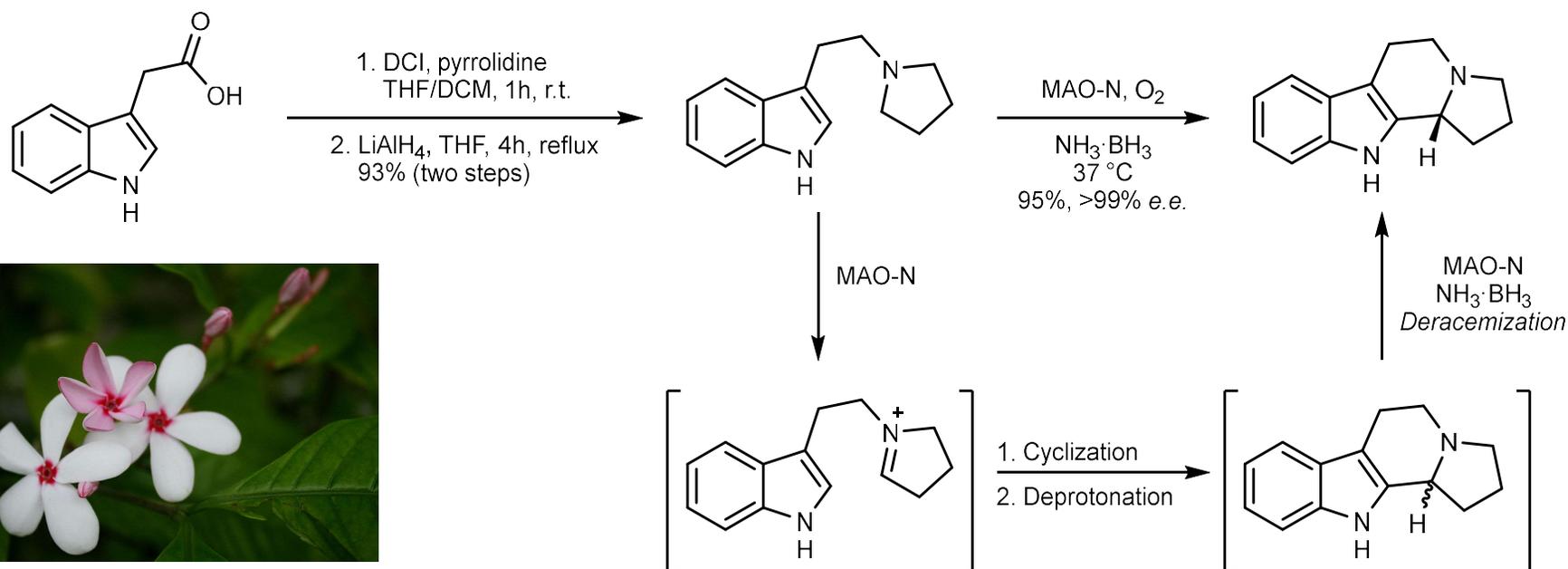


Group Problem #3



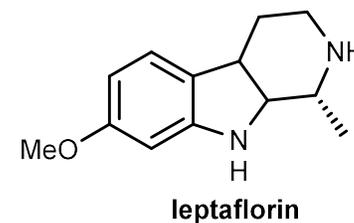
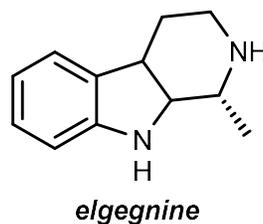
How would you prepare this natural product using biocatalysis?

Turner's Synthesis of Harmicine:



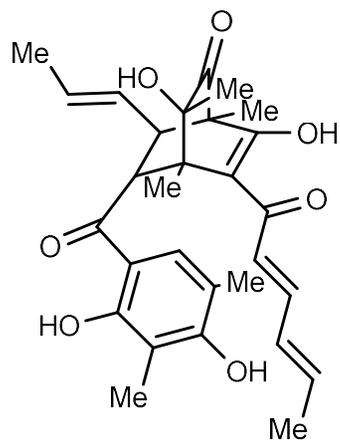
Kopsia griffithii

Similarly prepared by deracemization:

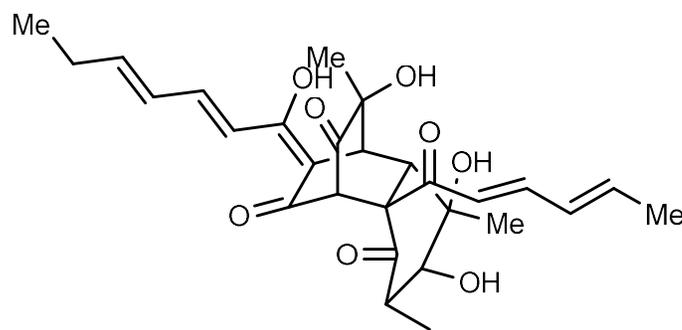


Interlude: NP Synthesis

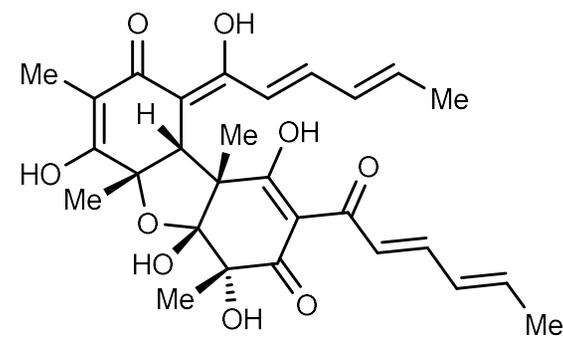
Natural products formed by biocatalytic cascades (Oxidative dearomatization, OQ-methide):



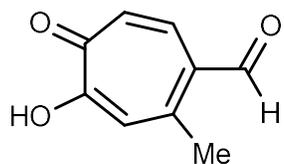
sorbiquinol
Gulder, 2017



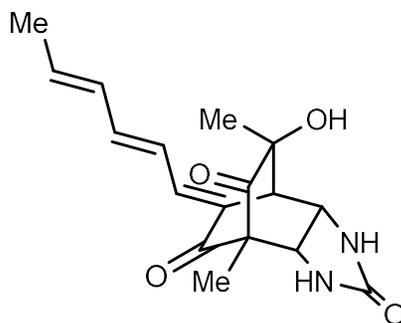
trichodimerol
Gulder, 2017



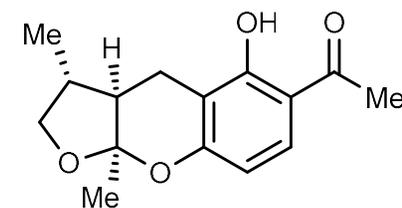
bisvertinolone
Gulder, 2018



stipitatic aldehyde
Narayan, 2018

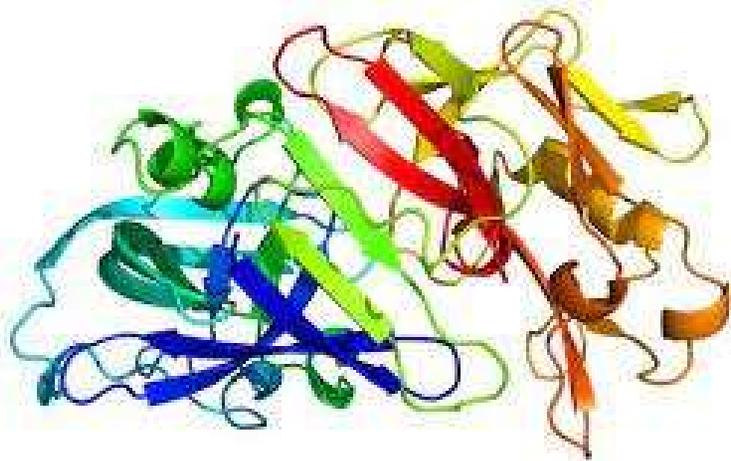


urea sorbicillinoid
Narayan, 2018

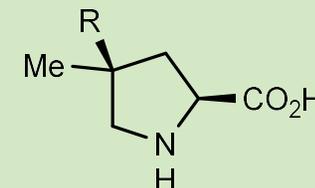
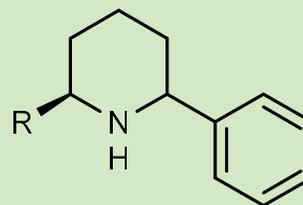


(-)-xyloketal D
Narayan, 2019

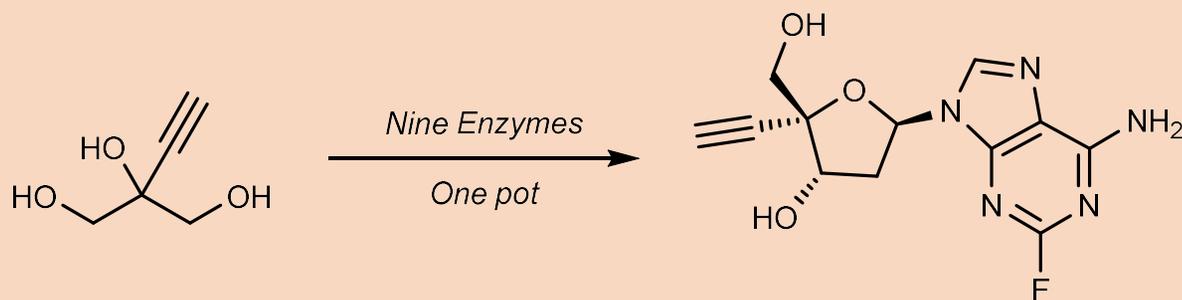
Outline



Enzyme classifications



Biocatalytic cascades for the synthesis of nitrogen heterocycles



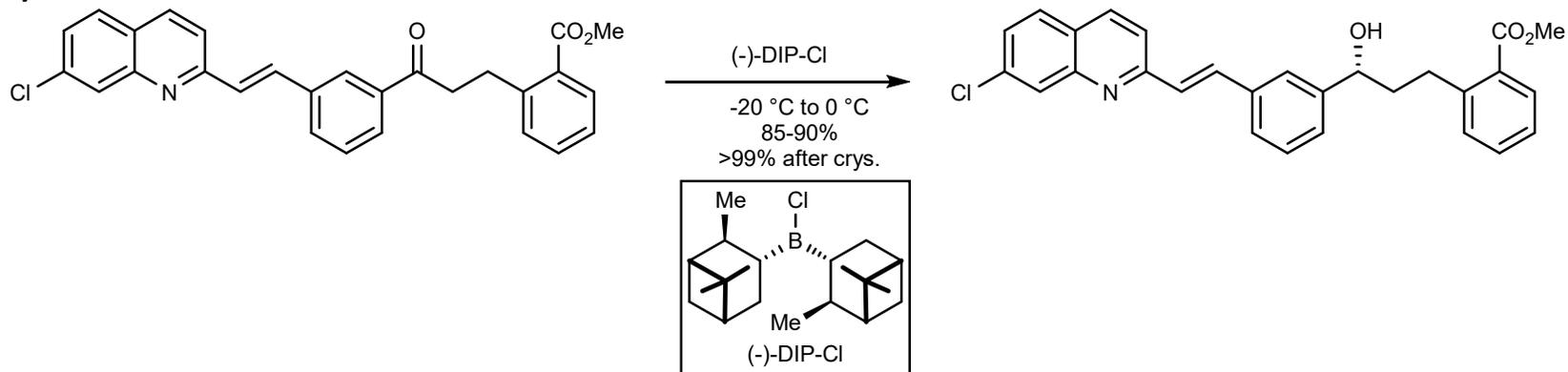
MERCK
INVENTING FOR LIFE

CODEXIS®

Pharmaceuticals prepared by biocatalytic cascades

Montelukast

Original synthesis:



Biocatalysis approach:

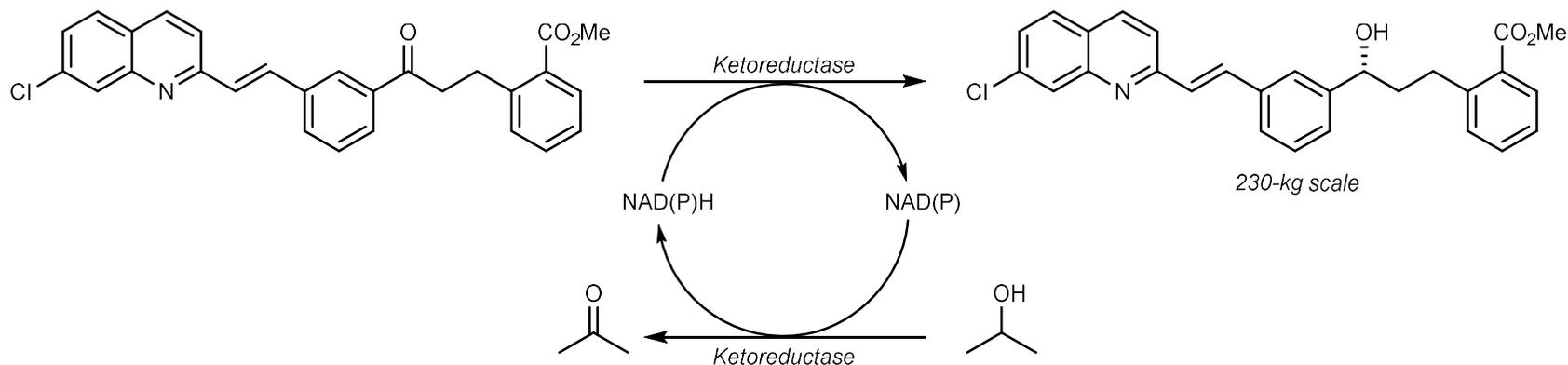
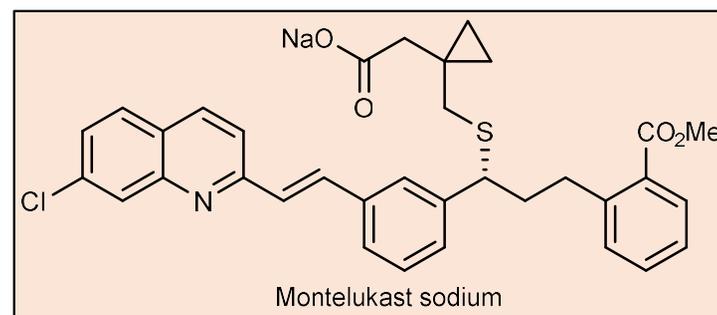


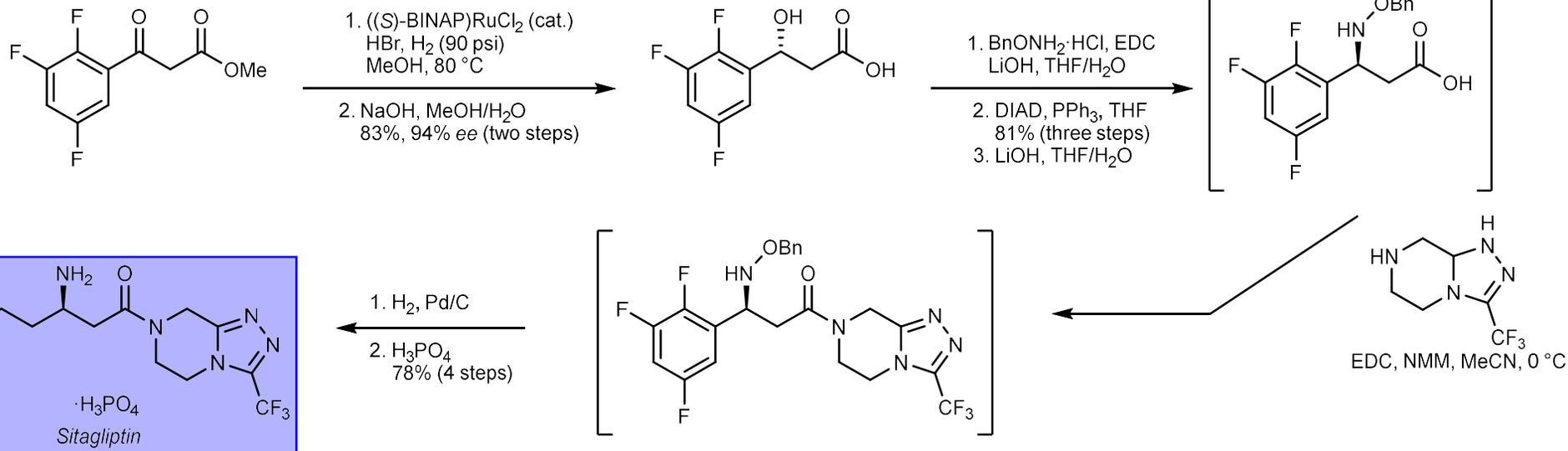
Table 5. Process conditions and performances of the DIP-Cl process vs the KRED process

	(-)-DIP-Cl	KRED
substrate loading	~100 g/L	100 g/L
reaction conditions	moisture sensitive	aqueous
reagent or catalyst loading	at least 1.8 equiv (at least 150 wt %)	3–5 wt %
optical purity	crystallization needed to upgrade >99% ee	crude product is >99.9% ee (upgrade not needed)
yield	85–90%	90–98%

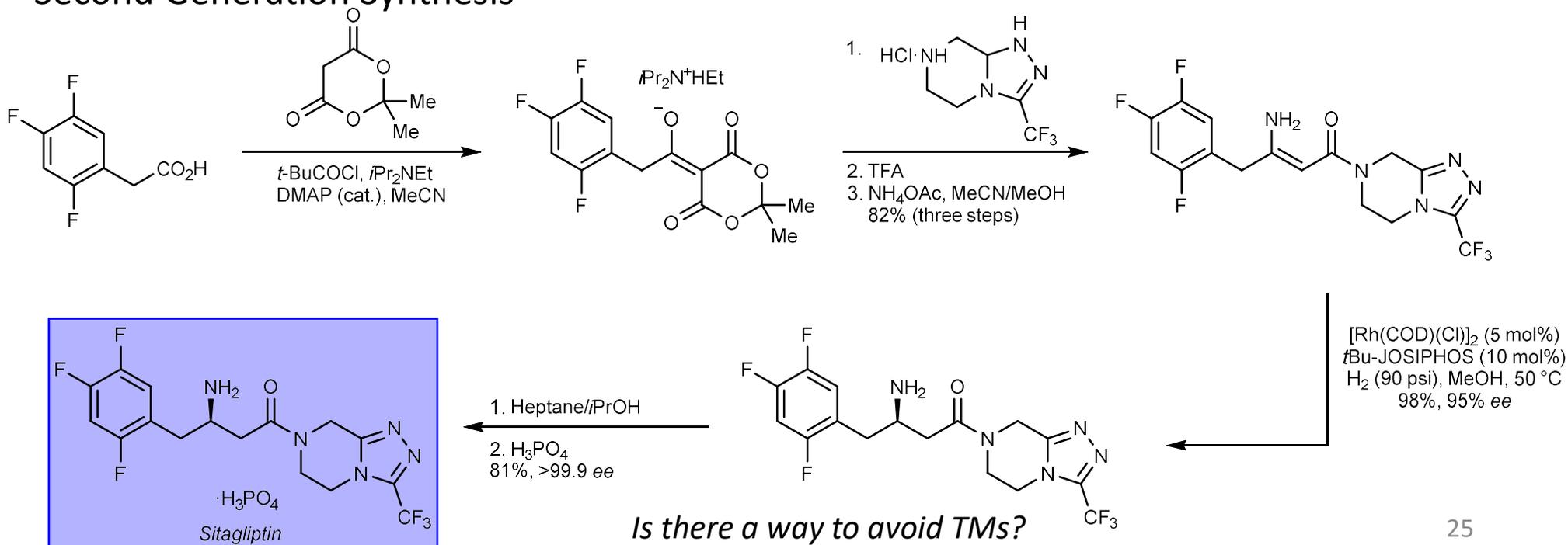


Sitagliptin

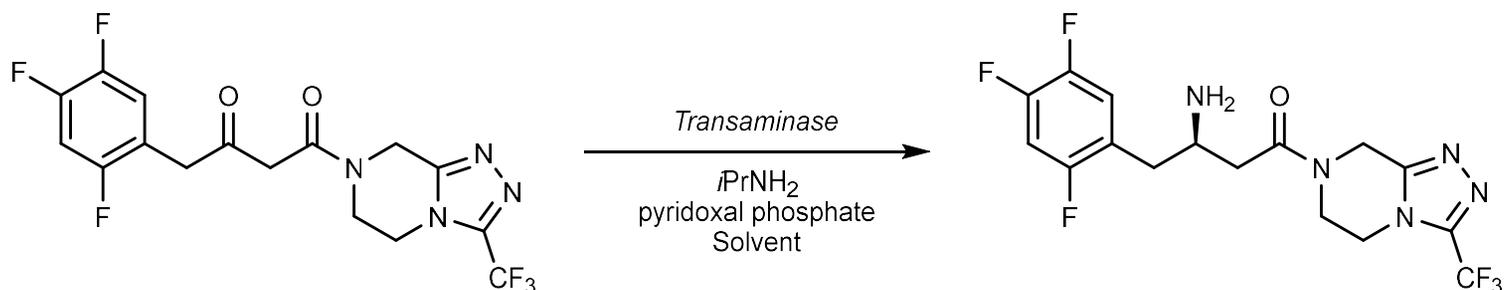
First Generation Synthesis



Second Generation Synthesis



Sitagliptin-Optimization

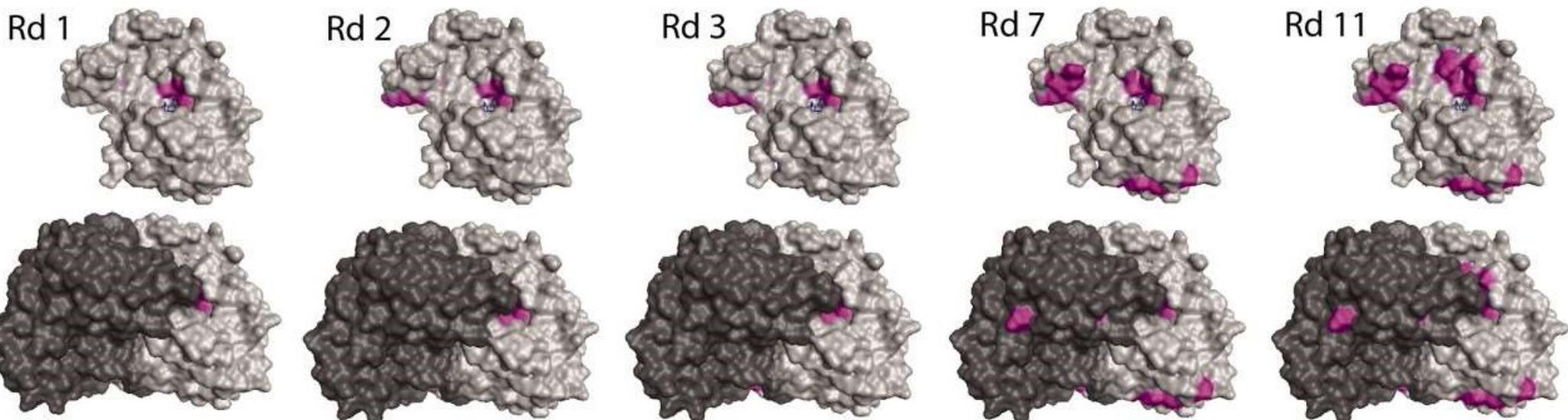


Added Mutations*	[Substrate] in g/l	Assay changes	Round identified	Improvement over parent†
S223P	2		1a	Not active
Y26H;† V65A;† V69G; F122I; A284G	2		1b	first active
H62T; G136Y; E137I; V199I; A209L; T282S	2		2	75
S8P; H26Y; G69C; M94I; I137T; G215C	5	5% DMSO to 5% MeOH; RT to 30°C	3	9
L61Y; C69T; Y136F; T137E	10	0.5 to 1 M <i>i</i> PrNH ₂ ; pH 7.5 to pH 8.5	4	4
D81G; I94L; I96L; T178S; L269P; P297S; S321P	40	5 to 10% MeOH; 30 to 45°C	5	1.4
Y60F; L94I; A169L; S178T; G217N; L273Y	100	10 to 20% MeOH	6	1.6
S124H	100	20% MeOH to 25% DMSO	7	1.1
I122M; H124N	100		8	1.1
Q329H	100		9	1.9
N124T; Y150S; V152C; H329Q	50	25 to 50% DMSO; 0.5% acetone	10	2
S126T	50		11	1.4

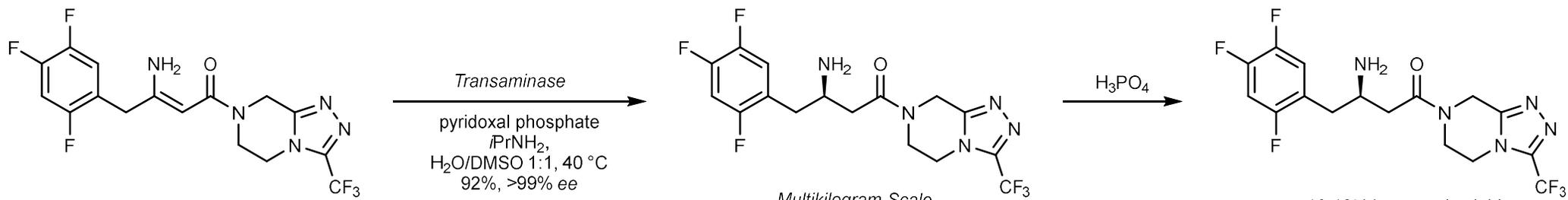
0.7% conversion/24h

Sitagliptin — a “Formal” Cascade

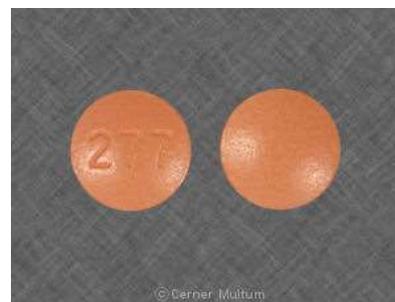
Effect of directed evolution on the binding site



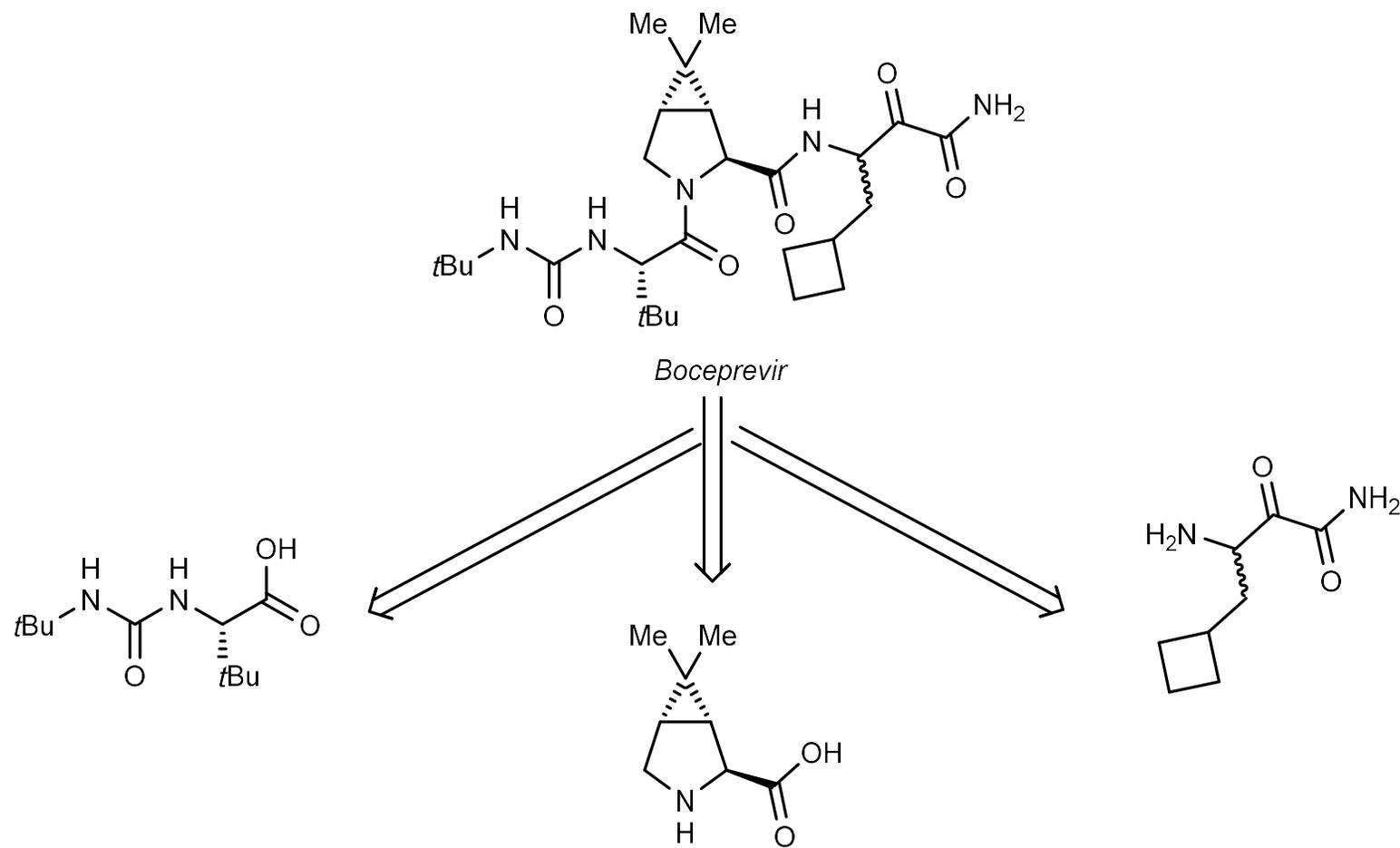
Final Route:



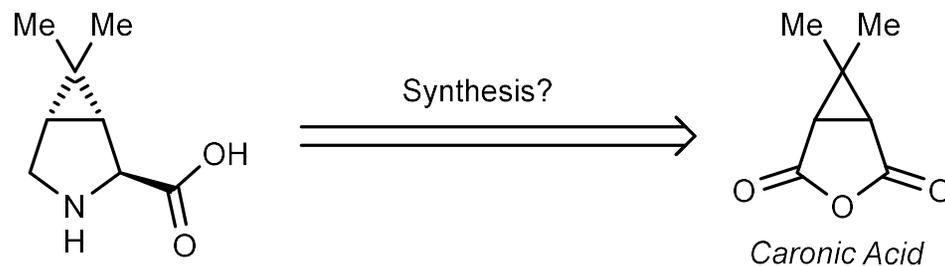
10-13% increase in yield
53% increase in productivity
19% reduction in waste



Boceprevir



Previous synthesis:
Classical resolution
Max yield of 50%



Boceprevir

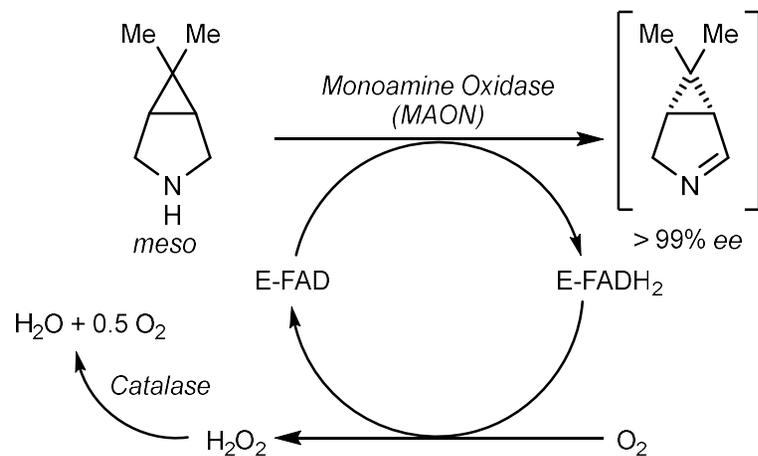
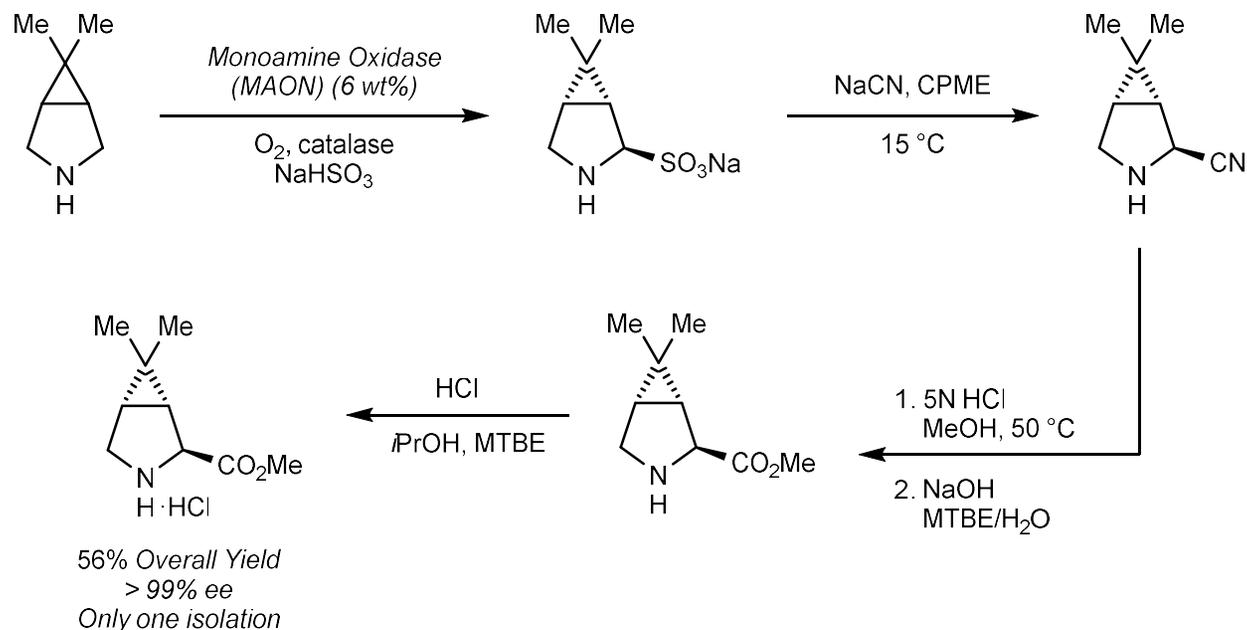


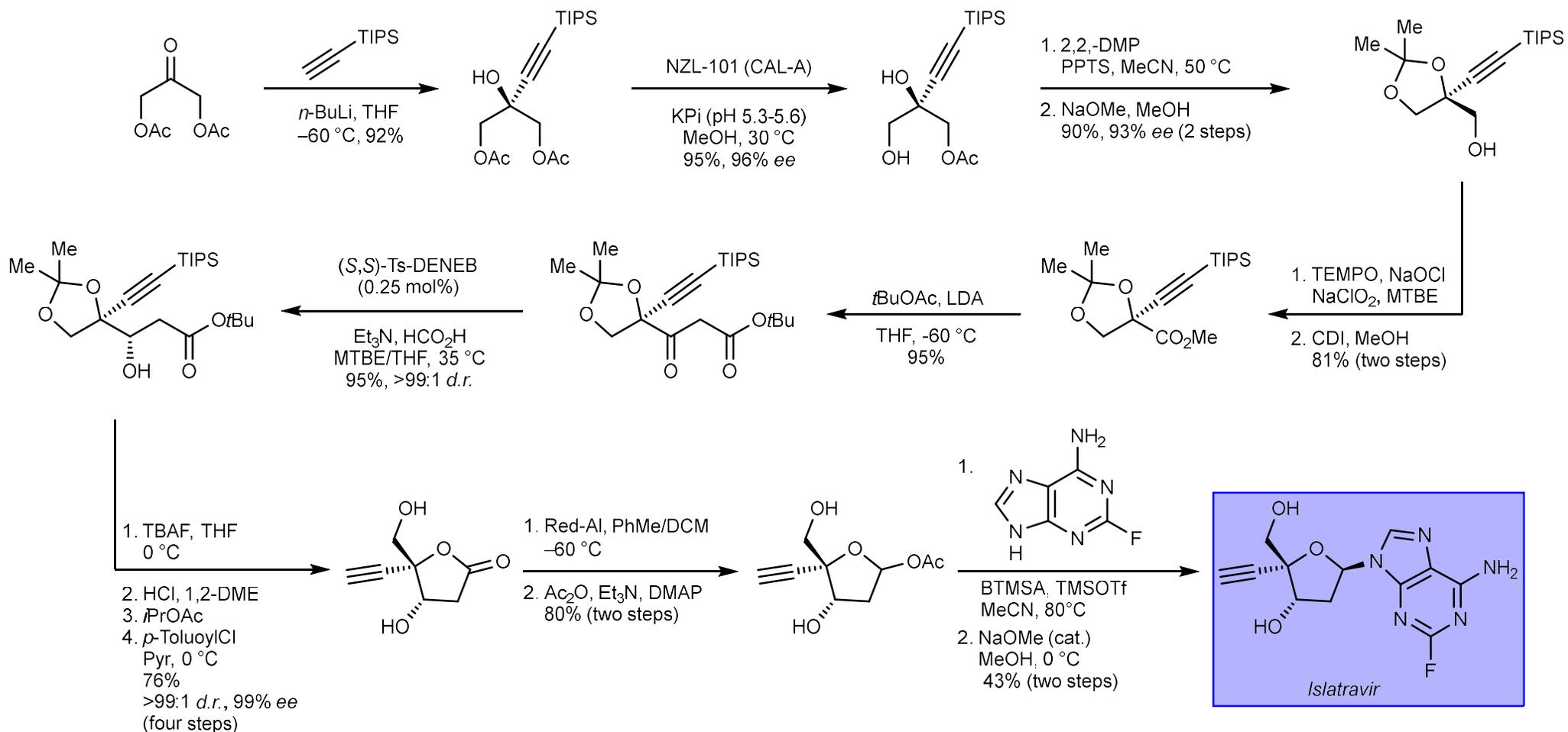
Table 1. Summary of MAON Improvement over the Four Rounds of Evolution

round	MAON variant	mutation	diversity source	improvement
	wild type	<i>A. niger</i> wild type		
1	156	<i>A. niger</i> + A289V, K348Q	active site targeted library	activity
2	274	many (see the Supporting Information)	<i>A. niger</i> and <i>A. oryzae</i>	thermal stability at 40 °C
2	291	150 + S465G	diversity from round 1	activity and solubility
3	301/304/308	many (see the Supporting Information)	MAON274 and MAON291	thermal stability at 50 °C
4	401	304 + F382L	active site library	8-fold increase in activity



Islatravir

First Generation:

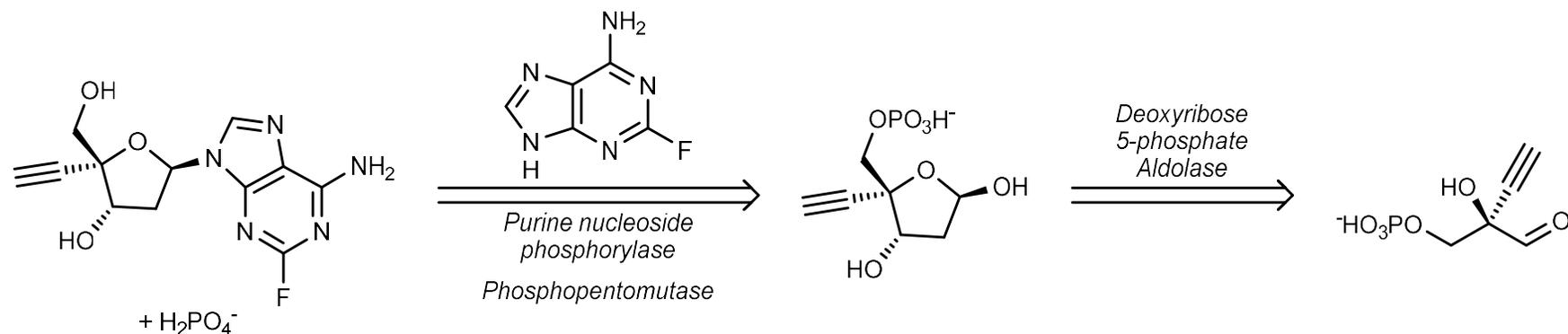


Disadvantages:

1. Multiple protecting group manipulations
2. Anomeric bond-forming step has poor yield
3. Long route—17 steps!

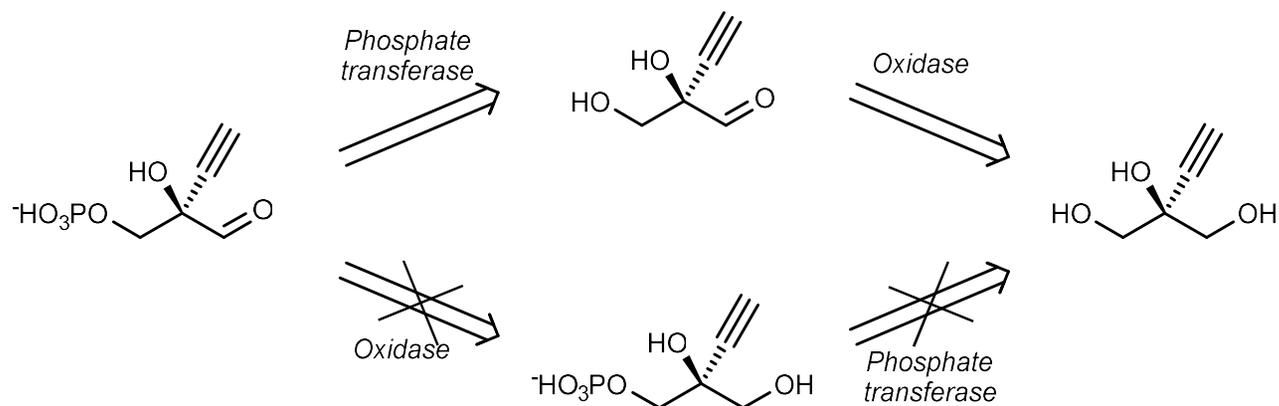
Islatravir

The Bacterial Nucleoside Salvage Pathway—Applied to Islatravir:



Could this cascade work in reverse to give the product? Accept unnatural substrates?

Retrosynthesis of the triol:



- Enzymes had been shown to work individually with unnatural substrates, not in sequence
- Major equilibrium challenges were anticipated
- How to setup a process like this?

Islatravir

The Cascade:

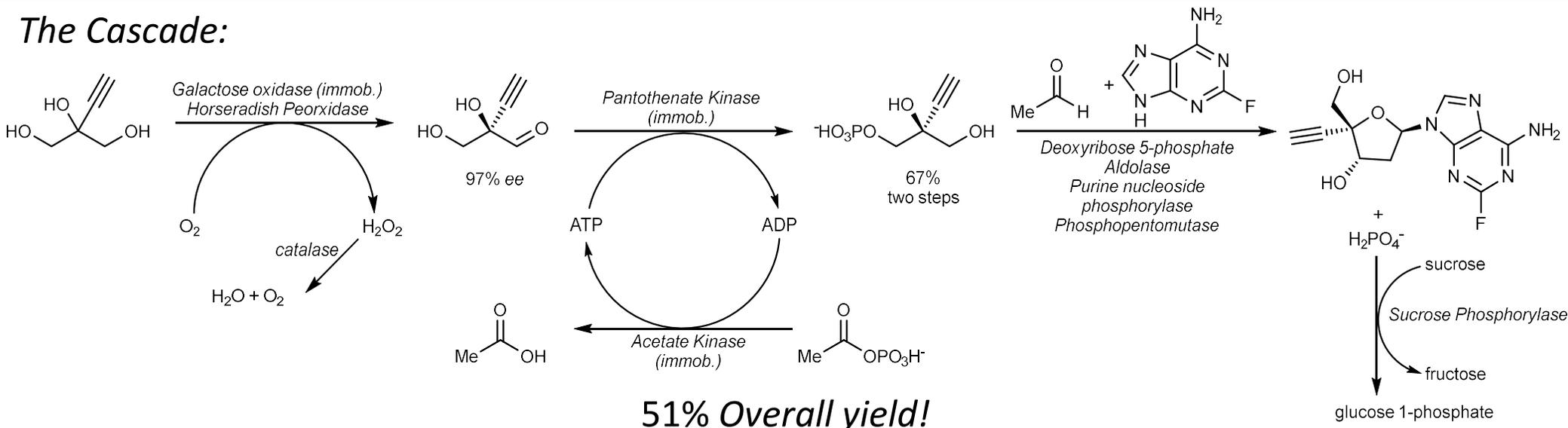


Table 1. Properties and performance of evolved enzymes used in the biocatalytic pathway.

Enzyme	Source organism	Evolution focus	Rounds of evolution	Global amino acids changed (no.)	Starting enzyme		Evolved variant	
					Conversion (selectivity)	Loading (%)*	Conversion (selectivity)	Loading (%)*
Oxidase (GOase)	<i>Fusarium graminearum</i>	Stereoselectivity	12	34	Variant M1: 33%††	100	80%†§	20
		Activity			(8:92 R:S)		(90:10 R:S)	
PanK	<i>E. coli</i>	Activity	3	10	<1% (5:1 R:S)†¶	10	>95% (10:1 R:S)†#	10
DERA	<i>S. halifaxensis</i>	Acetaldehyde tolerance	2	11	97%**[>98:1:1 (3S4R):(3R4R):(3S4S)]	5	97%**[>98:1:1 (3S4R):(3R4R):(3S4S)]	0.2
PPM	<i>E. coli</i>	Activity	2	5	0.5%††	0.5	34%††	0.5
PNP	<i>E. coli</i>	Activity	4	7	0.18%††	0.125	62%††	0.125
					(>99.5:0.5 dr)		(>99.5:0.5 dr)	

*Enzyme loading refers to the mass of lyophilized clarified cell lysate relative to the mass of the reaction substrate. Results may reflect improvements in enzyme expression as well as activity. †Reaction with nonimmobilized enzymes. ‡GOase-M1-Strep and F2-Strep: 172 mM **6**, pH 7.5, 0.2 mM CuSO₄, 25°C, 4 hours. §GOase-13BB-His: 258 mM **6**, pH 7.5, 0.2 mM CuSO₄, 25°C, 4 hours. ¶44 mM **7**, pH 7.5, 20°C, 18 hours. #235 mM **7**, pH 6.4, 20°C, 18 hours. **142 mM **5**, 420 mM acetaldehyde, pH 7.2, 30°C, 24 hours. ††15 mM **4**, 5 mM MnCl₂, pH 7.5, 40°C, 18 hours. †††13 mM **1b**, pH 7.5, 40°C, 16 hours.

Islatravir

The Cascade:

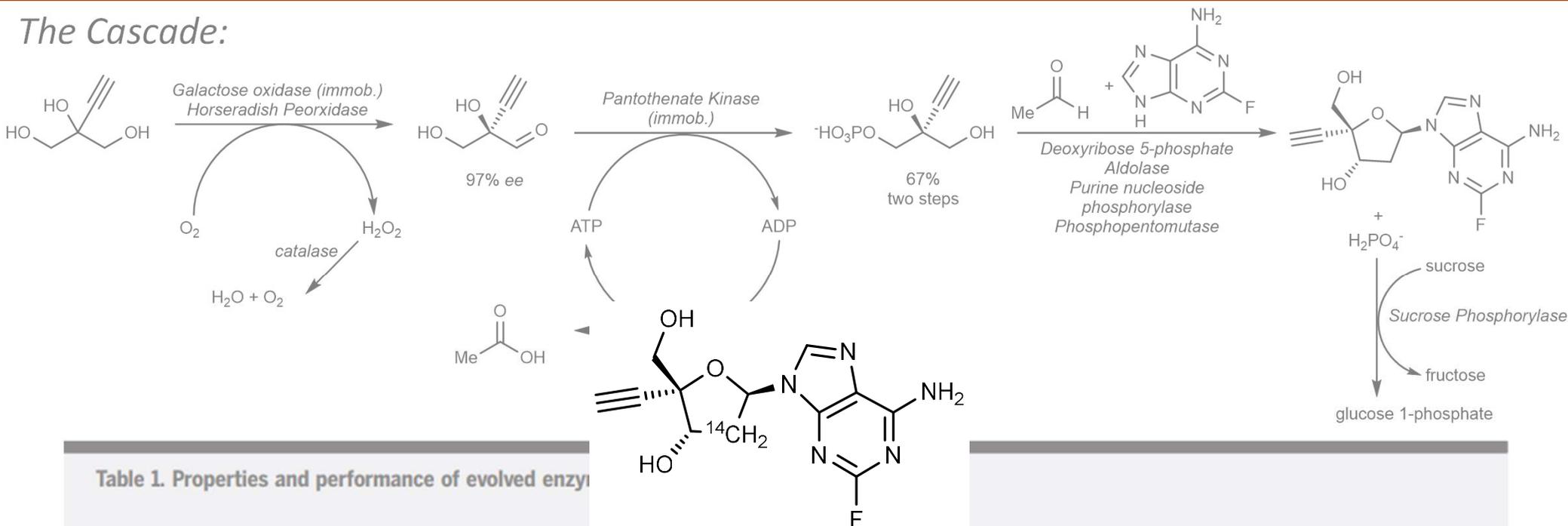


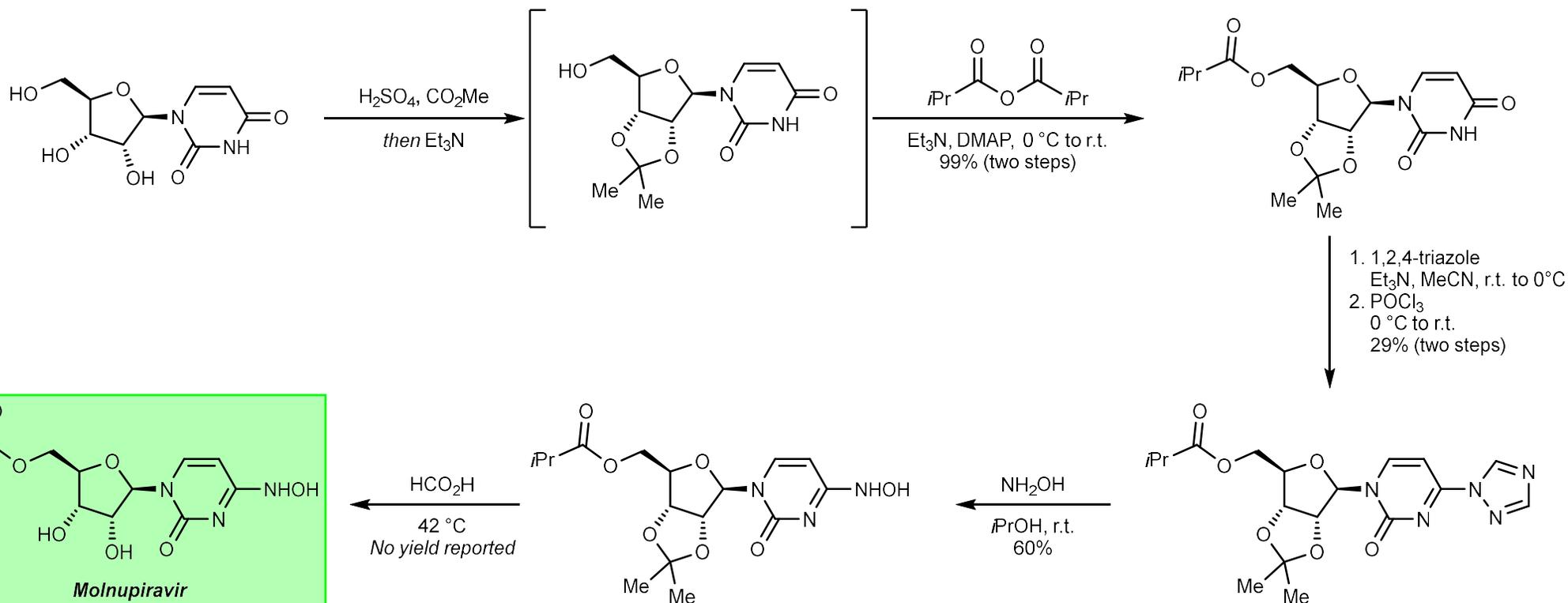
Table 1. Properties and performance of evolved enzymes

Enzyme	Source organism	Evolution focus	Rounds of evolution	Residues changed (no.)	Enzyme loading (selectivity)	Enzyme loading (%)*	Conversion (selectivity)	Enzyme loading (%)*
Oxidase (GOase)	<i>Fusarium graminearum</i>	Stereoselectivity	12	34	Variant M1: 33%†‡	100	80%†§	20
		Activity			(8:92 R:S)		(90:10 R:S)	
PanK	<i>E. coli</i>	Activity	3	10	<1% (5:1 R:S)†¶	10	>95% (10:1 R:S)†#	10
DERA	<i>S. halifaxensis</i>	Acetaldehyde tolerance	2	11	97%**[>98:1:1 (3S4R):(3R4R):(3S4S)]	5	97%**[>98:1:1 (3S4R):(3R4R):(3S4S)]	0.2
PPM	<i>E. coli</i>	Activity	2	5	0.5%††	0.5	34%††	0.5
PNP	<i>E. coli</i>	Activity	4	7	0.18%††	0.125	62%††	0.125
					(>99.5:0.5 dr)		(>99.5:0.5 dr)	

*Enzyme loading refers to the mass of lyophilized clarified cell lysate relative to the mass of the reaction substrate. Results may reflect improvements in enzyme expression as well as activity. †Reaction with nonimmobilized enzymes. ‡GOase-M1-Strep and F2-Strep: 172 mM **6**, pH 7.5, 0.2 mM CuSO₄, 25°C, 4 hours. §GOase-13BB-His: 258 mM **6**, pH 7.5, 0.2 mM CuSO₄, 25°C, 4 hours. ¶144 mM **7**, pH 7.5, 20°C, 18 hours. #235 mM **7**, pH 6.4, 20°C, 18 hours. **142 mM **5**, 420 mM acetaldehyde, pH 7.2, 30°C, 24 hours. ††15 mM **4**, 5 mM MnCl₂, pH 7.5, 40°C, 18 hours. †††13 mM **1b**, pH 7.5, 40°C, 16 hours.

Molnupiravir

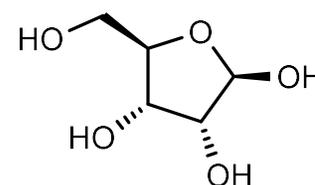
Patent route:



Disadvantages of route:

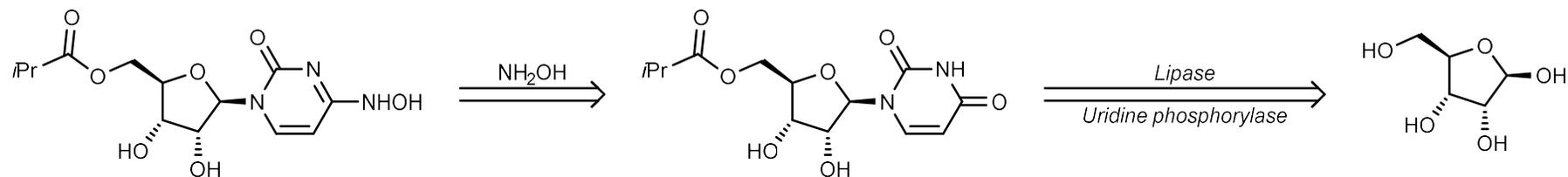
- Relies on supply-chain-dependent uridine
- Maximum of 17% yield
- Multiple isolation steps
- Protecting groups
- Not supply chain friendly

Could a route be prepared from ribose?

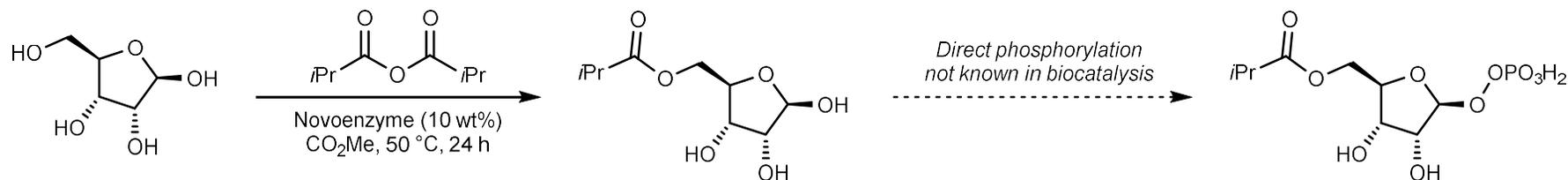


Molnupiravir

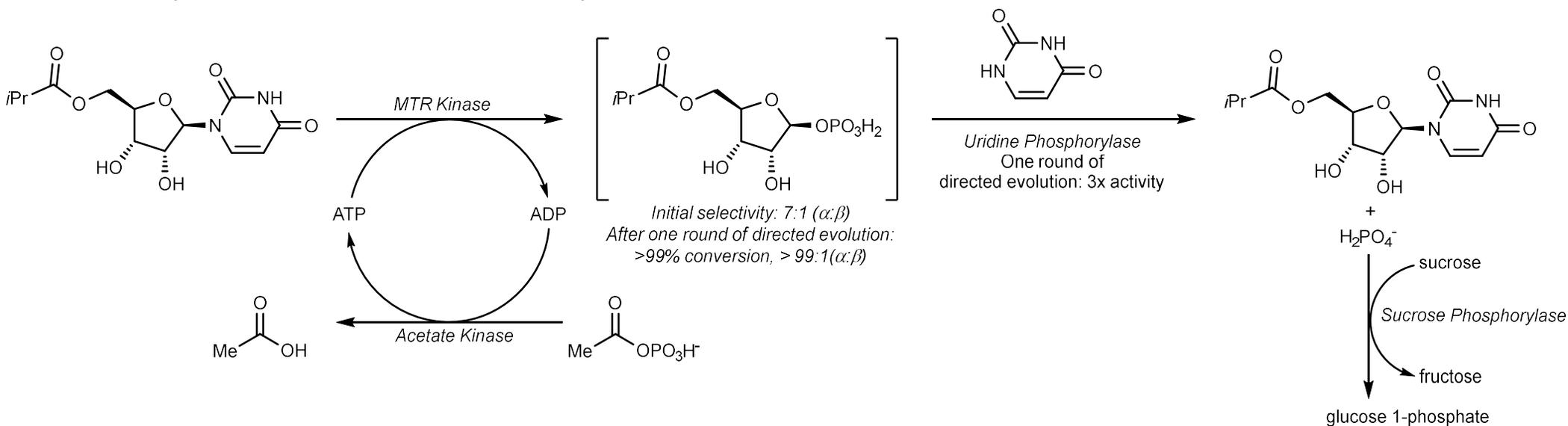
Retrosynthetic Analysis:



Initial Route:



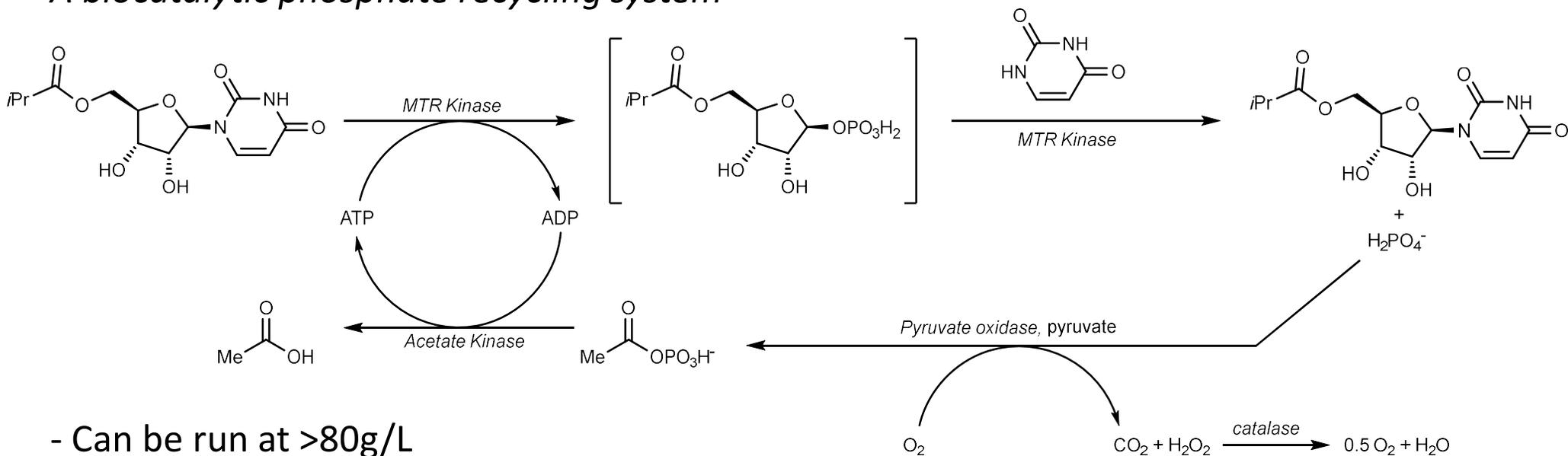
Development of a novel biocatalytic cascade:



Could further improvements be made by recycling the inorganic phosphate?

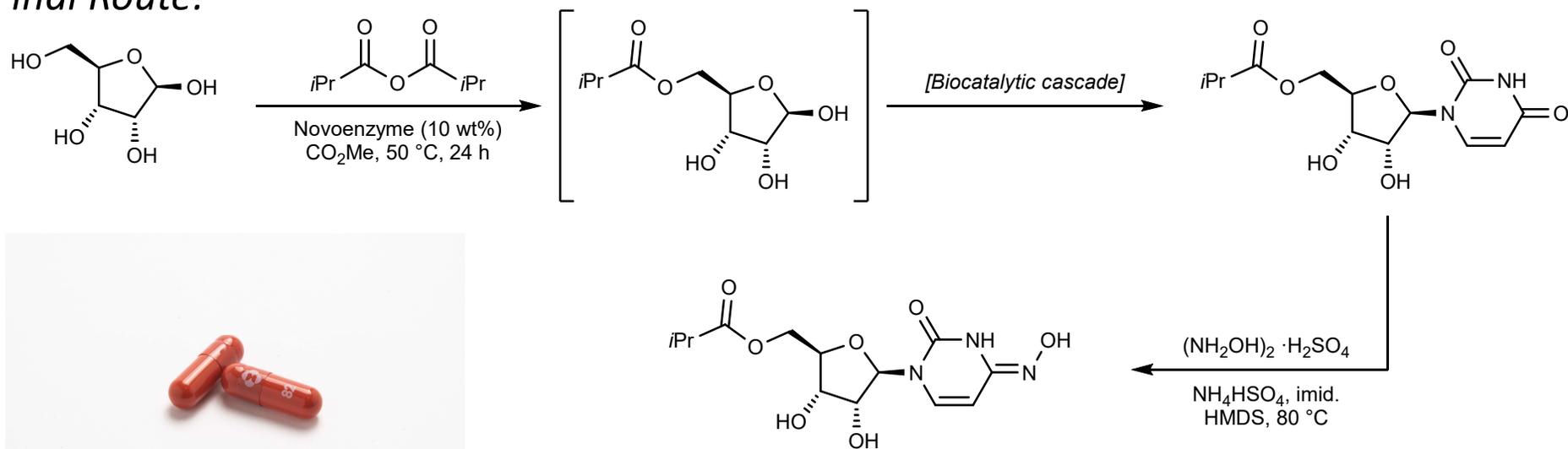
Molnupiravir

A biocatalytic phosphate recycling system

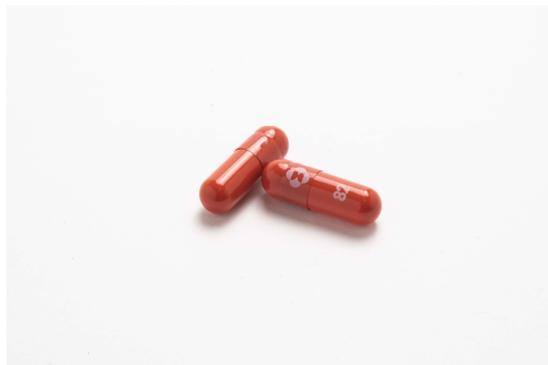


- Can be run at >80g/L

Final Route:



3 Steps
Decagram Scale
69% overall yield
70% shorter; ~7x yield
improvement



Conclusions

Conclusions:

- Biocatalytic cascades are fantastic at:
 - Performing protecting group free synthesis
 - Performing exceptionally chemoselective and stereoselective transformations
 - Obtaining high-value chemicals from cheap starting materials
 - Limitations
 - Evolution of enzymes is often necessary for scaling-up
 - Finding the “right” enzyme can be very tricky
 - Biocatalysis is not non-specialist friendly
-

Future Directions:

- Cascades with P450s
- Evolving or discovering aldolases to expand possible donors
- Preparation of artificial metallenzymes in cascades (few examples)
- Implementation of a directed evolution system for non-specialists
- Combining chemocatalysis and biocatalysis (beyond reductions)
- Designing systems that can be done on meaningful scale (beyond industry)

Further Reading

ChemSusChem
Chemistry–Sustainability–Energy–Materials



Chemistry Europe
European Chemical Societies Publishing

RETURN TO ISSUE | < PREV REVIEW NEXT >

Artificial Biocatalytic Linear Cascades for Preparation of Organic Molecules

Joerg H. Schrittwieser[†], Stefan Velikogne[‡], Mélanie Hall[†], and Wolfgang Kroutil^{††}

View Author Information

Cite this: *Chem. Rev.* 2018, 118, 1, 270–348
Publication Date: May 8, 2017
<https://doi.org/10.1021/acs.chemrev.7b00033>
Copyright © 2017 American Chemical Society
[RIGHTS & PERMISSIONS](#)

Article Views	Altmetric	Citations
13328	-	301

[LEARN ABOUT THESE METRICS](#)

Review | Full Access

Broadening the Scope of Biocatalysis in Sustainable Organic Synthesis

Prof. Roger A. Sheldon Prof. Dean Brady

First published: 01 April 2019 | <https://doi.org/10.1002/cssc.201900351> | Citations: 102

Review | Free Access

Biocatalysis: A Pharma Perspective

Joseph P. Adams, Murray J. B. Brown Alba Diaz-Rodriguez, Richard C. Lloyd, Gheorghe-Doru Roiban

First published: 05 May 2019 | <https://doi.org/10.1002/adsc.201900424> | Citations: 43

SECTIONS

PDF TOOLS SHARE

Constructing Biocatalytic Cascades: In Vitro and in Vivo Approaches to de Novo Multi-Enzyme Pathways

Scott P. France[†], Lorna J. Hepworth[†], Nicholas J. Turner^{*}, and Sabine L. Flitsch^{*†}

View Author Information

Cite this: *ACS Catal.* 2017, 7, 1, 710–724
Publication Date: December 12, 2016
<https://doi.org/10.1021/acscatal.6b02979>
Copyright © 2016 American Chemical Society
[RIGHTS & PERMISSIONS](#) Subscribed

Article Views	Altmetric	Citations
5063	4	220

[LEARN ABOUT THESE METRICS](#)

Share Add to Export

PDF (3 MB)

SUBJECTS: Chemical reactions, Alcohols, Peptides and proteins, Amines, Genetics