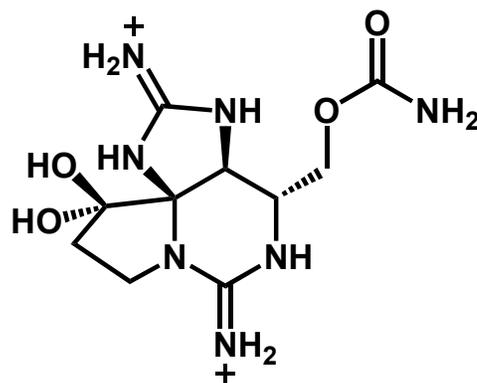


Total Synthesis of Saxitoxin



John Baird

April 11, 2006

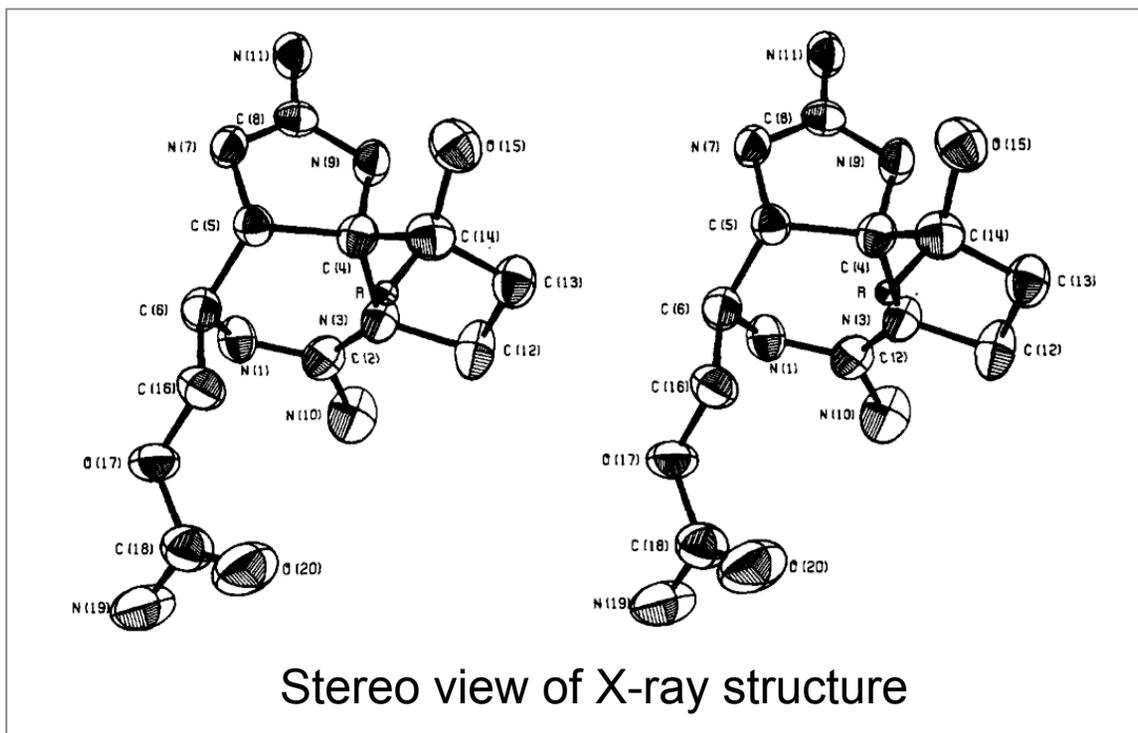
Isolation and Properties

First isolated in 1957
by Schantz et al

X-Ray Structure
obtained in 1975

Freely soluble in water
and methanol

Optical rotation $+84.8^\circ$



Amorphous solid with formula $C_{10}H_{17}N_7O_4$

Reported to be one of the most toxic non-protein compounds known

mw 299.3 g/mol

Biological Properties of Saxitoxin

Potent neurotoxin isolated from toxic Alaska butter clams (*Saxidomus giganteus*) and soft shell clams (*Mytilus californias*)

Established in 1920's that shellfish became toxic during the time of "red tides" during which *Gonyaulax catenella* (algal species) blooms. Under optimal temperature and light conditions blooms of red algae proliferate to generate red tides

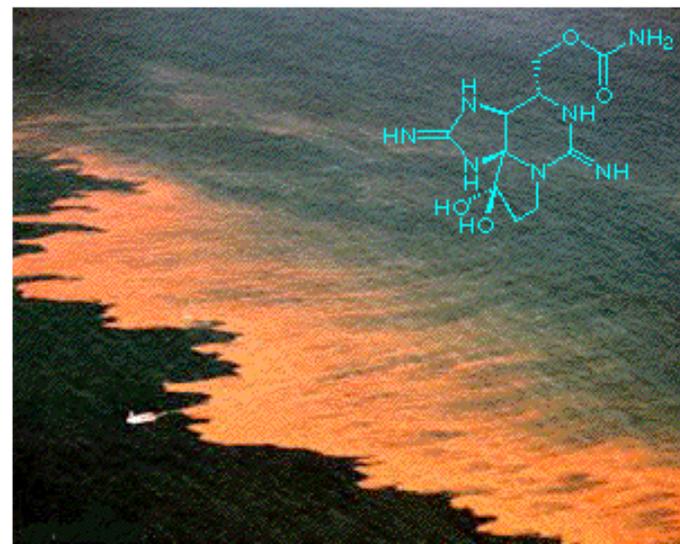
Responsible for paralytic shellfish poisoning.

No known cure, treatment available

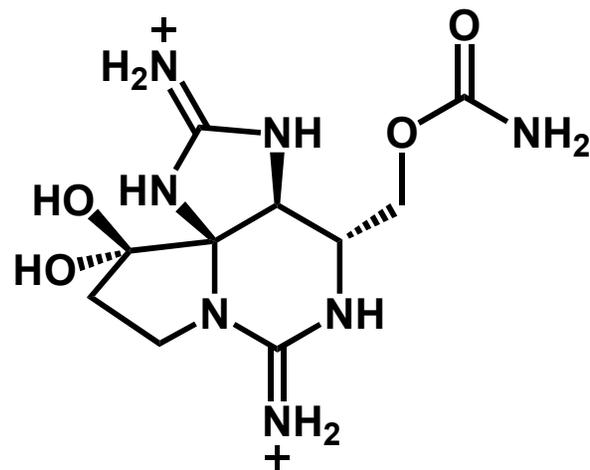
Symptoms include tingling, numbness, weakness, and limp paralysis

Toxicity of 8 μ g/Kg in mice; 0.2-1.0 mg would prove fatal to humans; 100x more poisonous than strychnine, 1000x (sarin gas), 2000x (NaCN)

Biological action: blocks sodium ion channels in neuron membranes (medicinal interest)



Key Synthetic Challenges



Molecule possesses:

All carbon atoms except one carry heteroatoms

3 contiguous stereogenic centers

Tricyclic skeleton possessing two guanidines

Polarity/solubility of the molecule makes manipulation/purification cumbersome

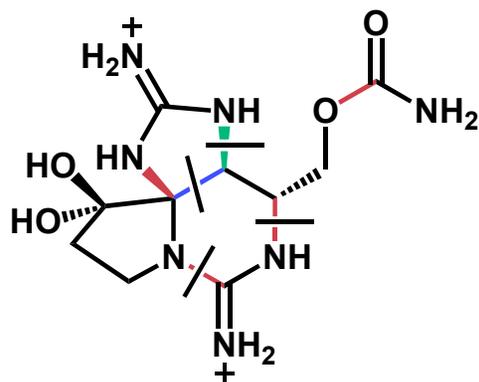
Total Syntheses

First by Kishi, made racemic saxitoxin in 1977, and later reported the synthesis of unnatural (-)-decarbamoylsaxitoxin in 1992.

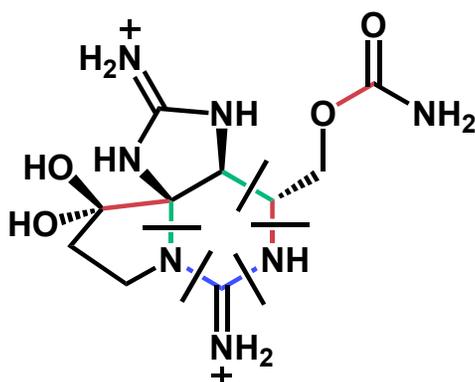
Second synthesis of racemic saxitoxin by Jacobi in 1984

Most recently, synthesis of natural (+)-Saxitoxin by Du Bois (2006)

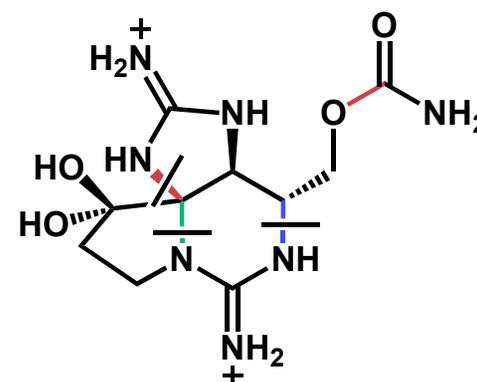
Kishi



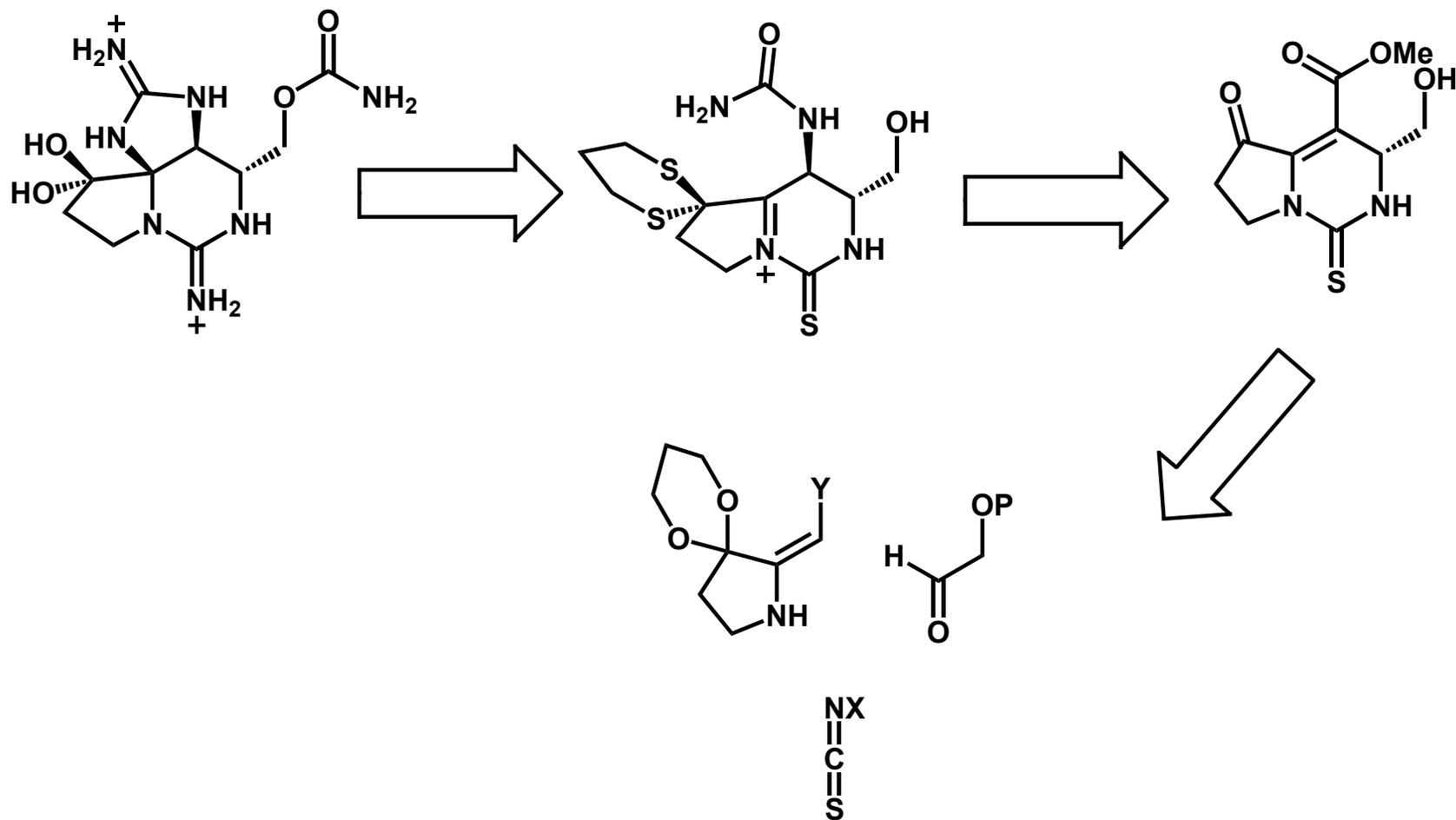
Jacobi



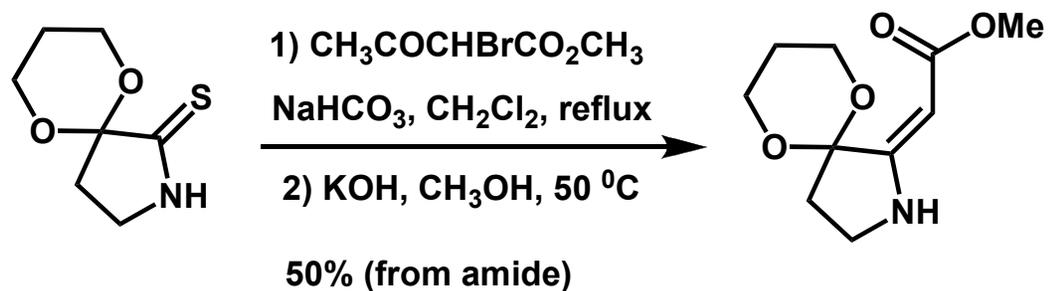
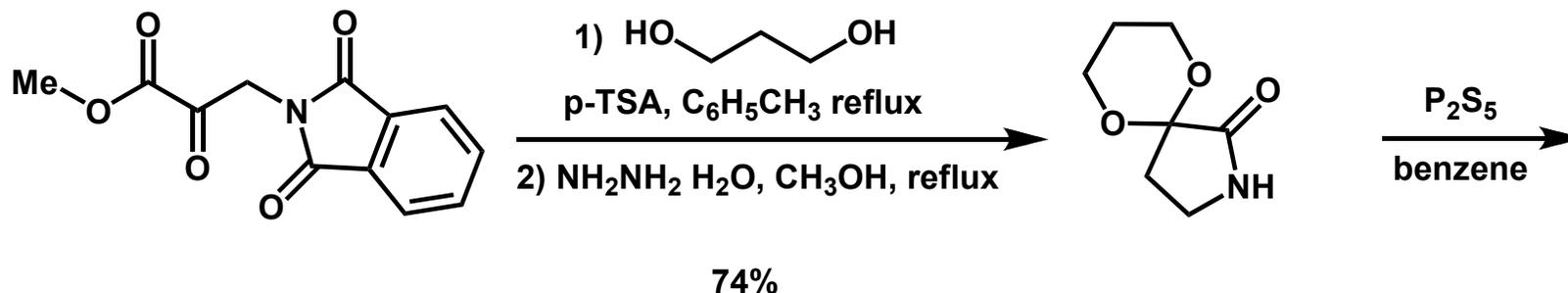
Du Bois



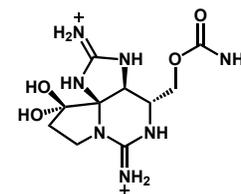
Kishi Retrosynthesis



Preparation of Vinylogous Carbamate



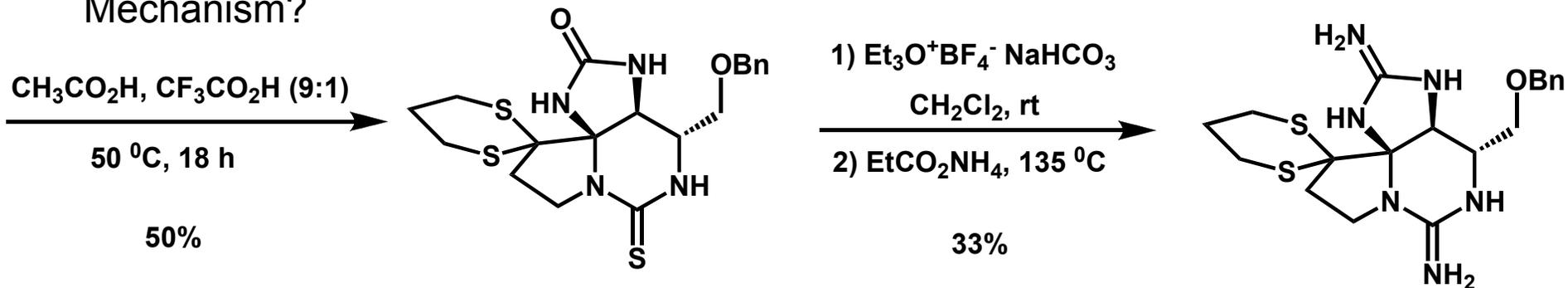
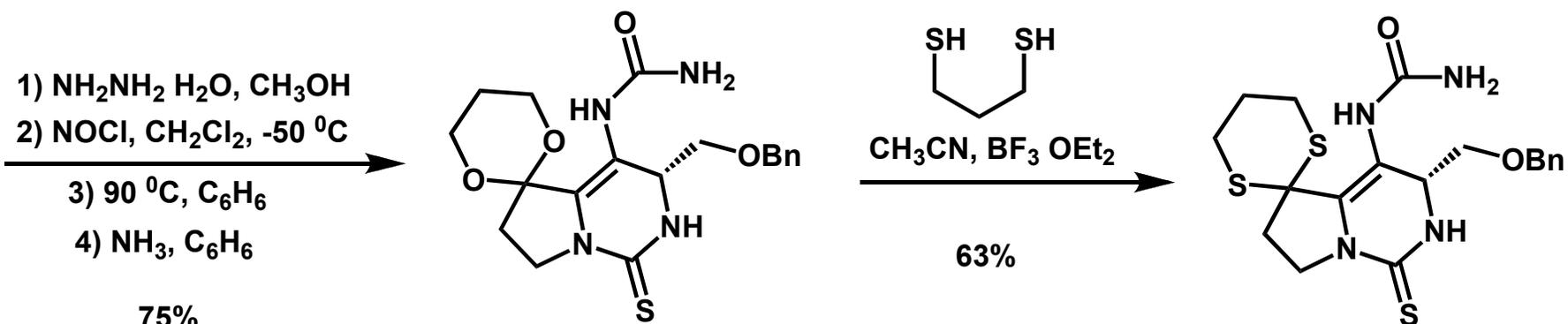
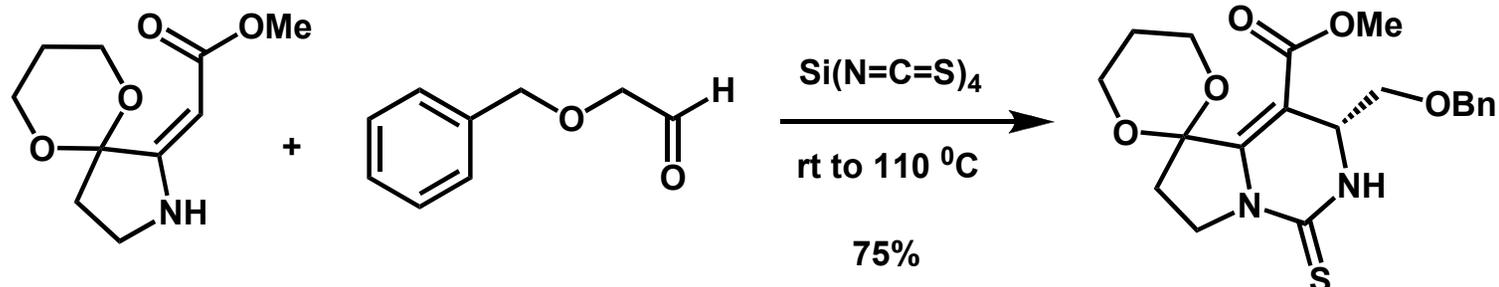
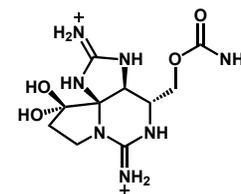
Please propose a mechanism for this reaction



Kishi, Y. et. al. *J. Am. Chem. Soc.* **1977**, *98*, 2818.

Kishi, Y. et. al. *Heterocycles.* **1980**, *14*, 1477.

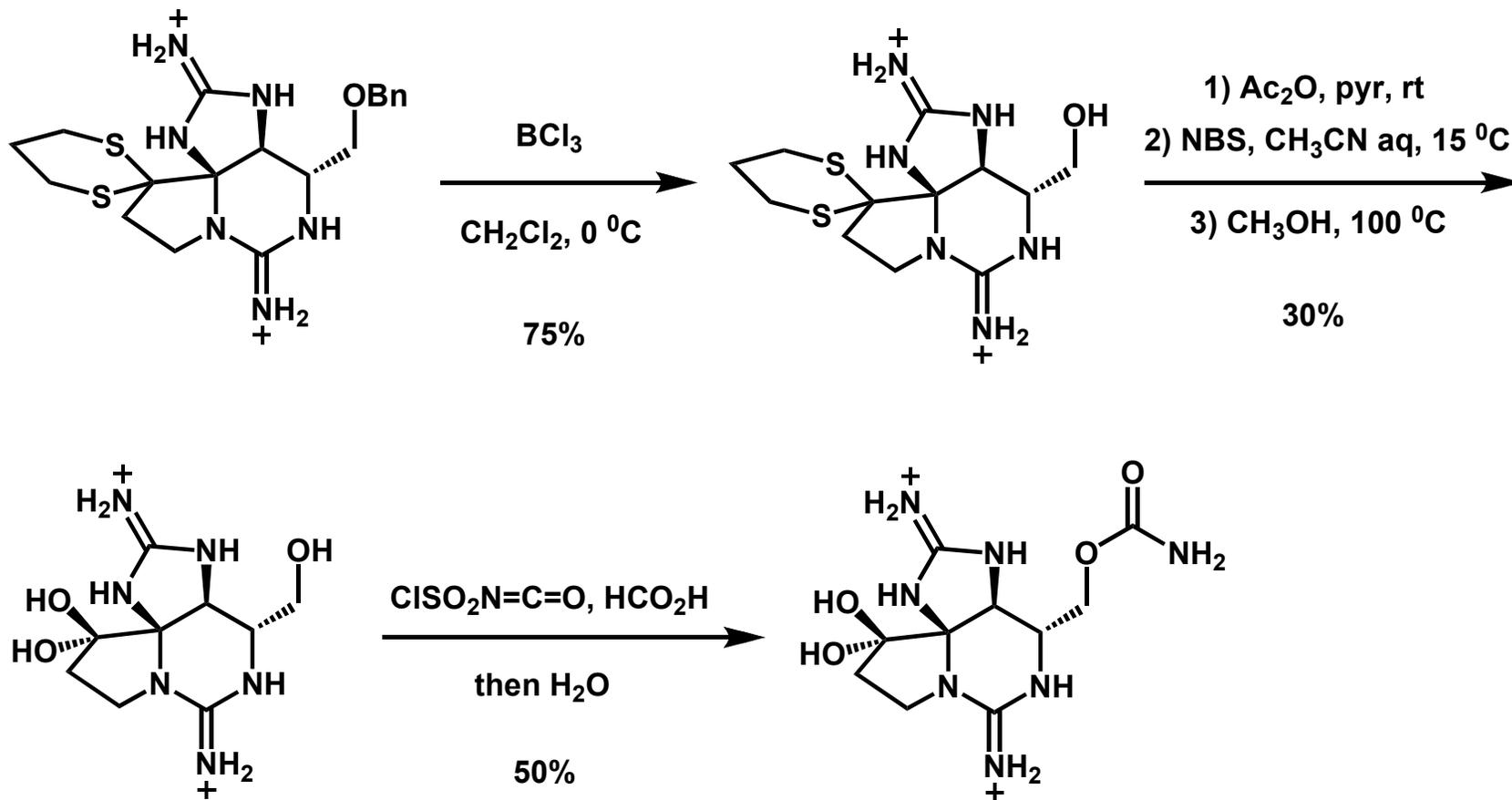
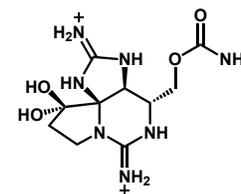
Preparation of Diguandine Core



Kishi, Y. et. al. *J. Am. Chem. Soc.* **1977**, *98*, 2818.

Kishi, Y. et. al. *Heterocycles.* **1980**, *14*, 1477.

Completion of the Synthesis



Kishi, Y. et. al. *J. Am. Chem. Soc.* **1977**, 98, 2818.

Kishi, Y. et. al. *Heterocycles.* **1980**, 14, 1477.

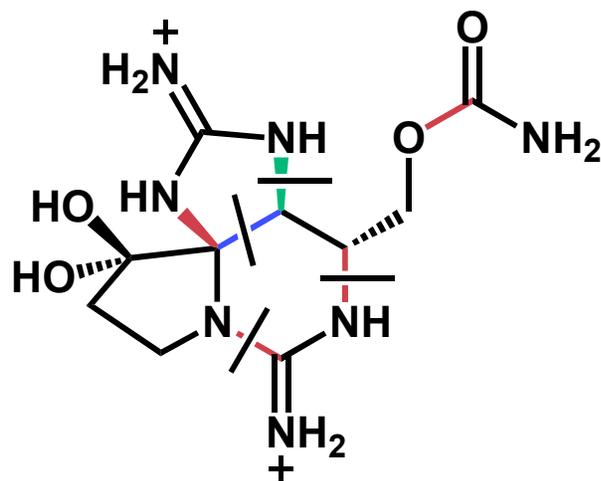
Kishi Synthesis- Summary

First total synthesis (racemic) 1977

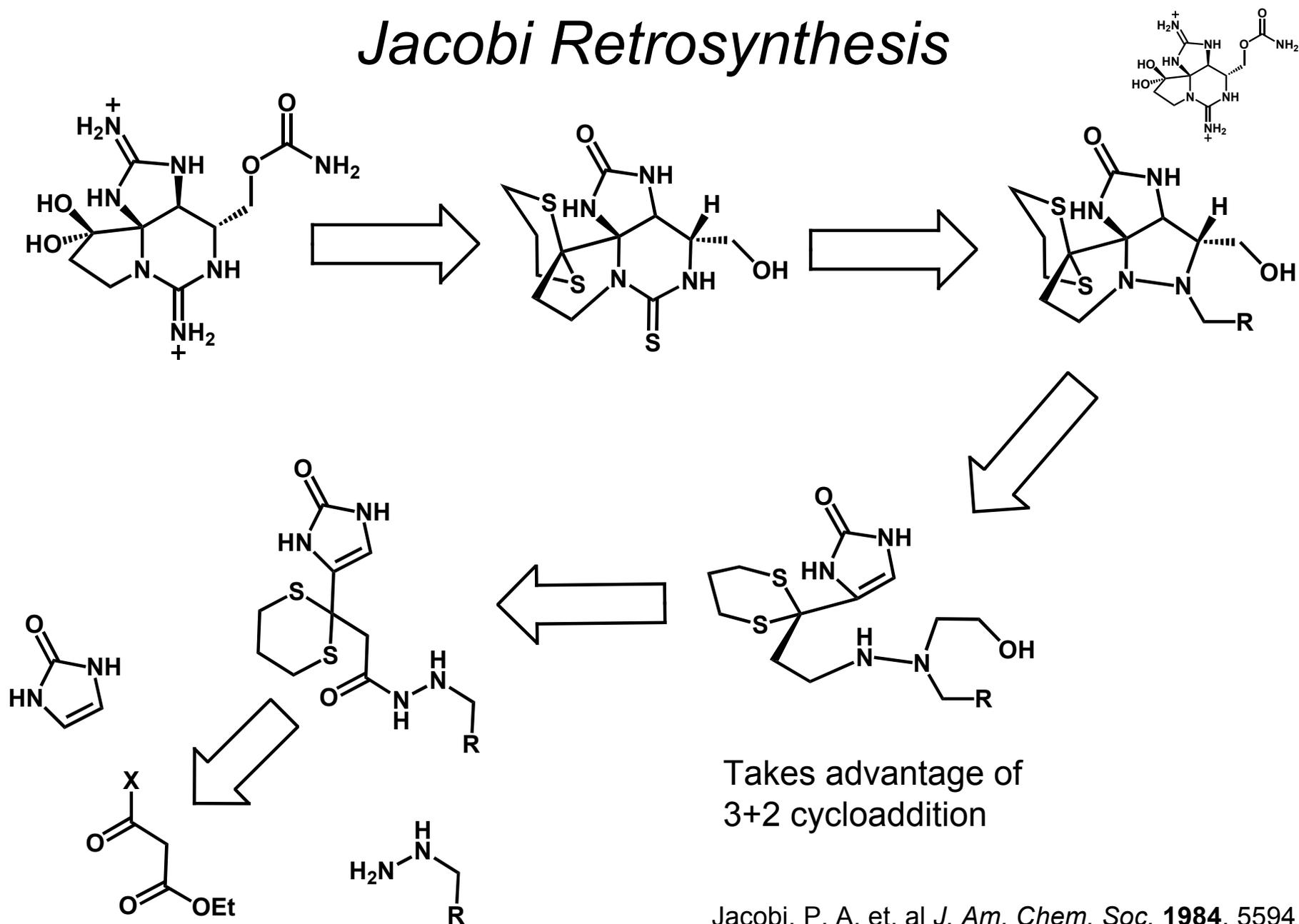
Key Steps:

Eschenmoser sulfide contraction
Curtius Rearrangement
Enamine condensation

17steps, 0.2% overall yield



Jacobi Retrosynthesis

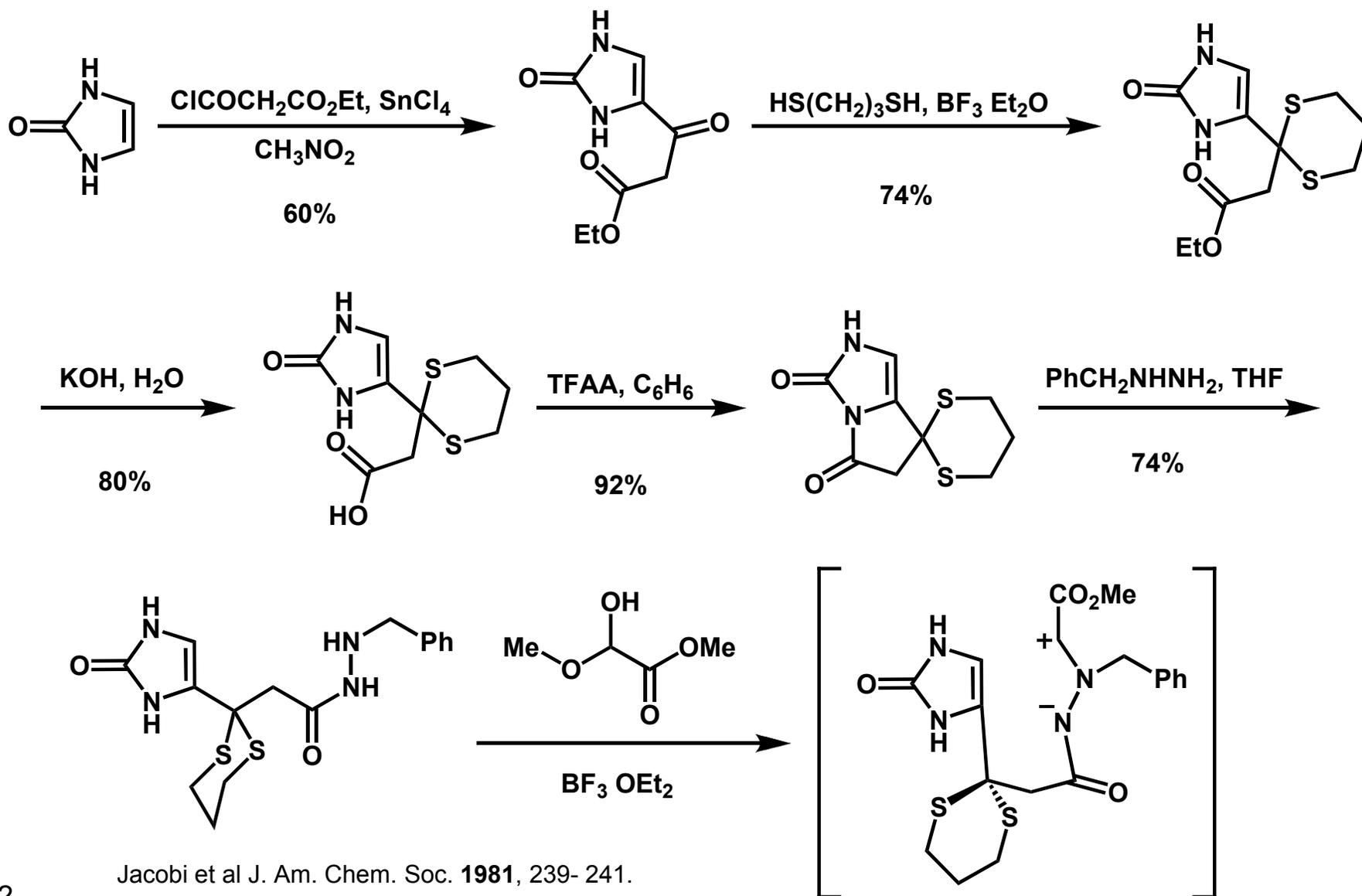
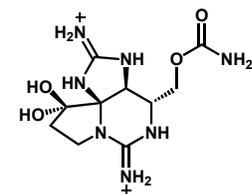


Takes advantage of
3+2 cycloaddition

Jacobi, P. A. et. al *J. Am. Chem. Soc.* **1984**, 5594

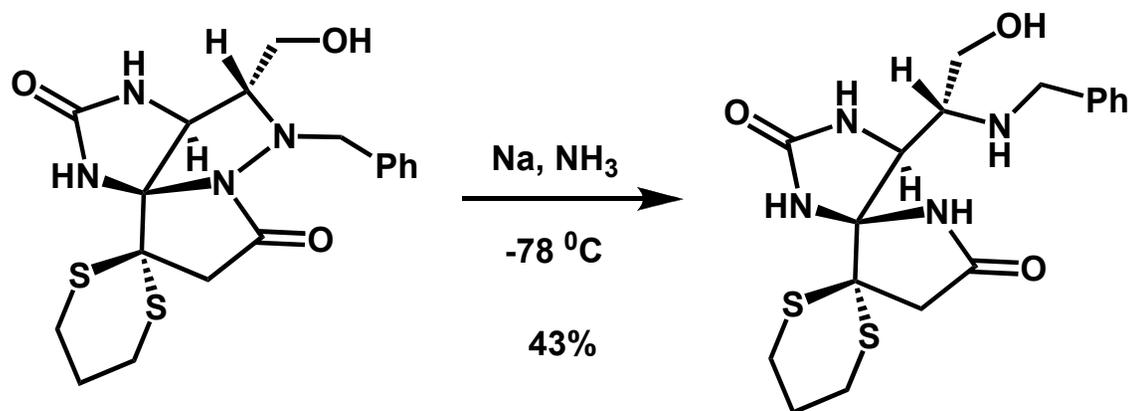
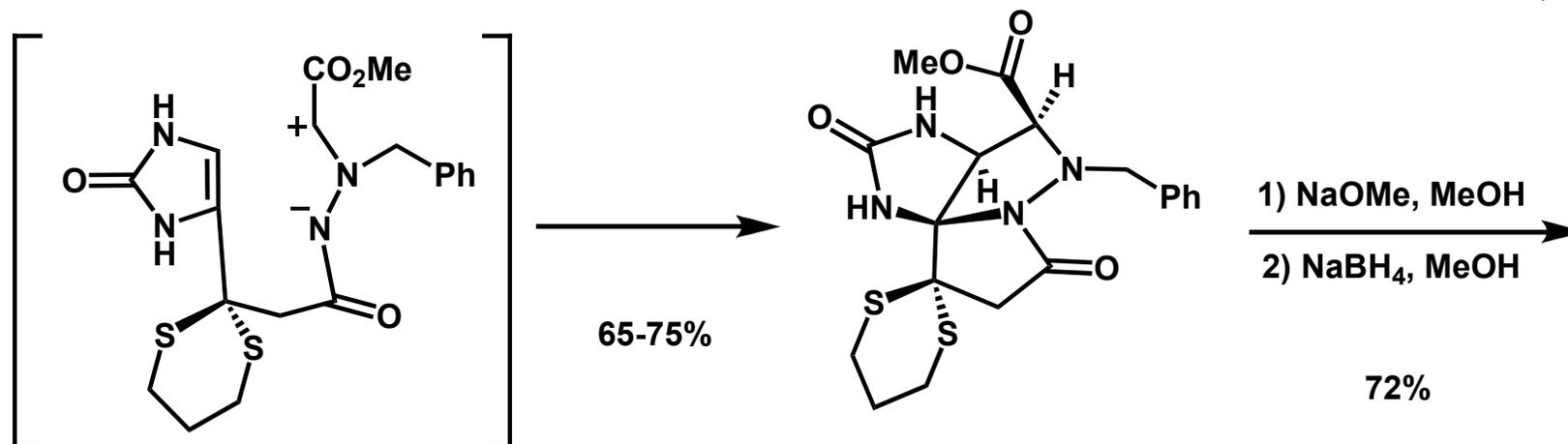
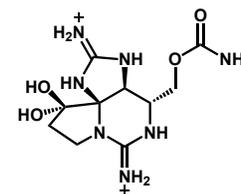
Jacobi, P. A. et. al *Croat. Chem. Acta.* **1986**, 2674

Preparation of Cycloaddition Precursor



Jacobi et al J. Am. Chem. Soc. **1981**, 239- 241.

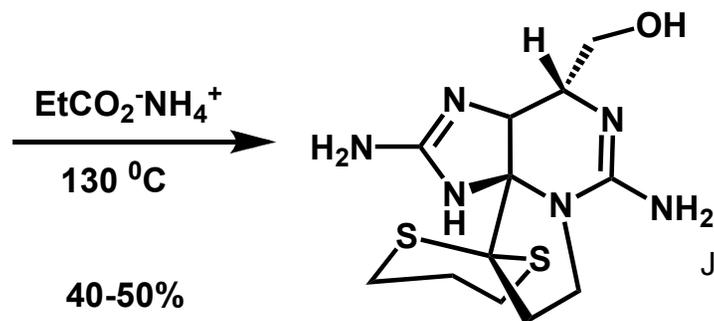
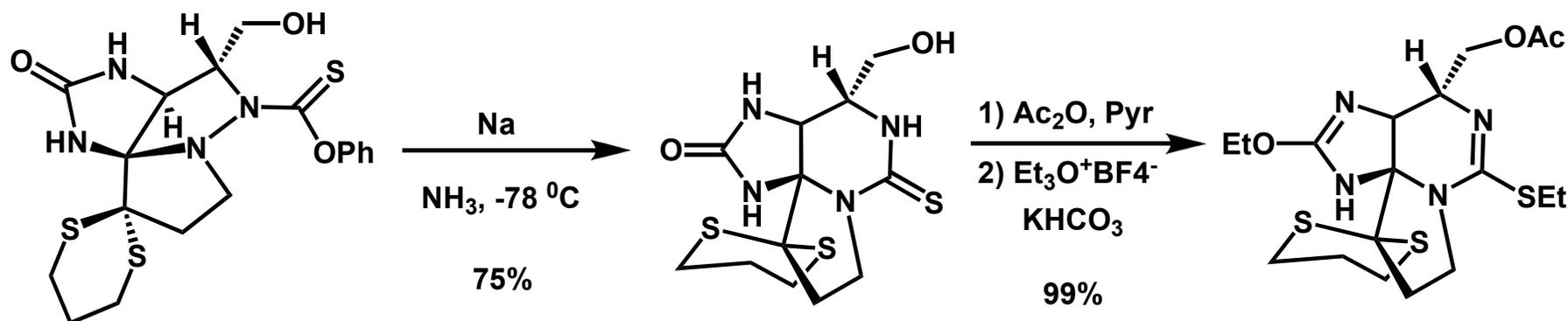
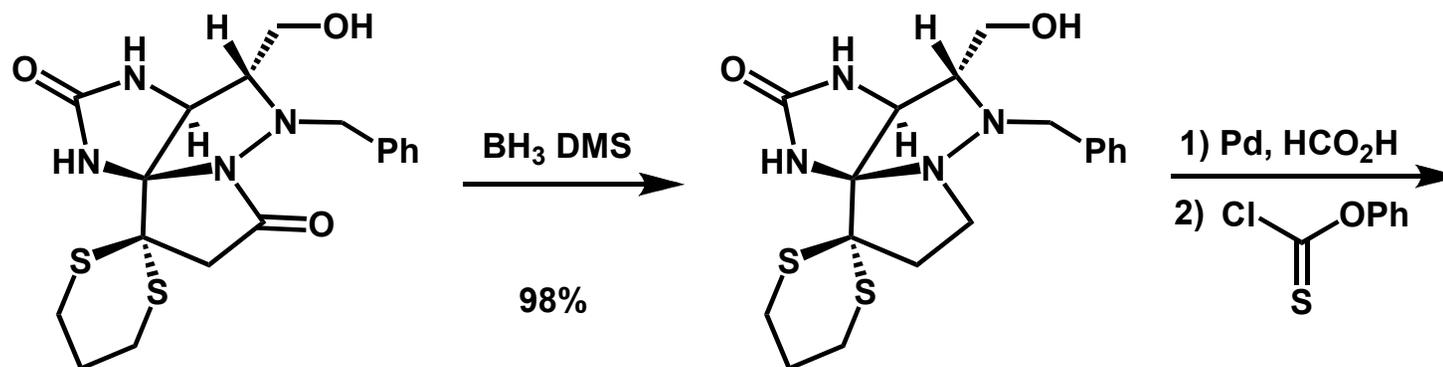
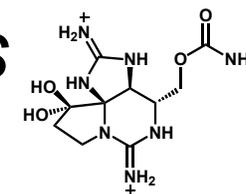
Preparation of Pyrazolidine



Further attempts at reduction
gave decomposition

Jacobi, P. A.; Martinelli, M. J. *J. Am. Chem. Soc.* 1984, 106, 5594-5598

Completion of the Formal Synthesis

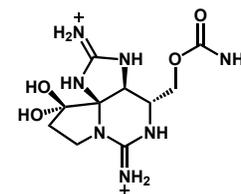


Intersects with
Kishi's synthesis

Jacobi, P. A.; Martinelli, M. J. *J. Am. Chem. Soc.* 1984, 106, 5594-5598

Jacobi, P. A. et al *Croat. Chem. Acta* 1986, 267

Jacobi Synthesis- Summary



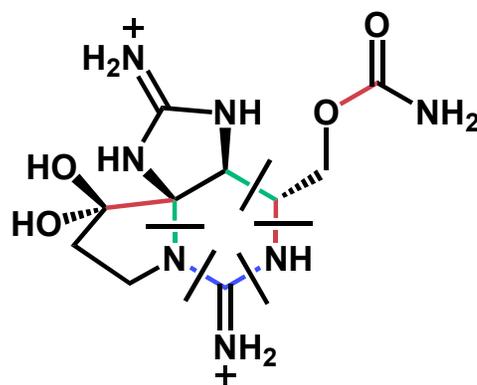
Intersected Kishi intermediate

Utilized [3+2] cycloaddition to access core

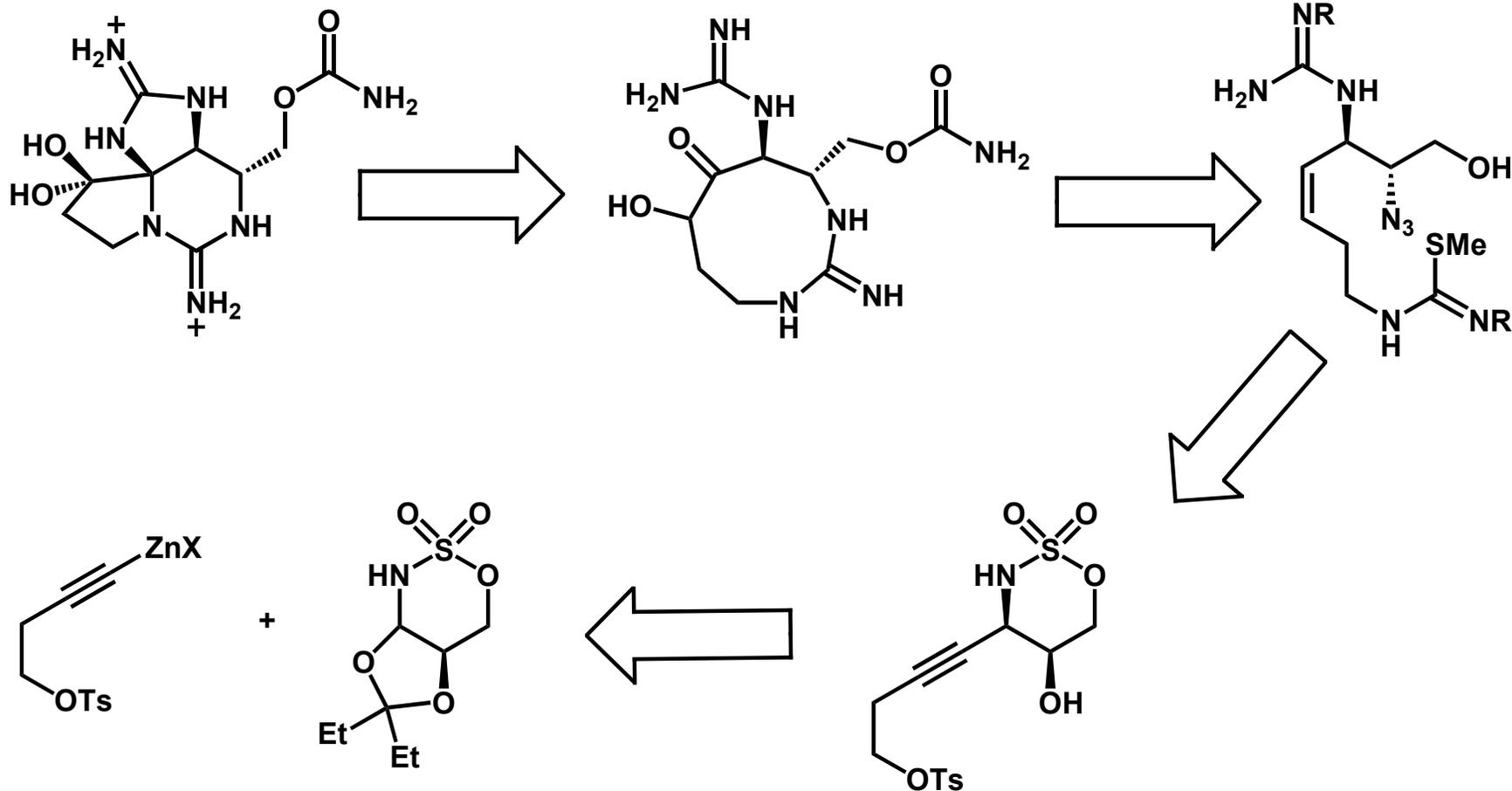
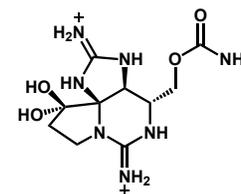
“Scalable and efficient” route has been used to prepare 1.0g of saxitoxin precursor

Second synthesis 1984 (racemic)

15 steps (3.3% yield)



Du Bois Retrosynthesis



Will take advantage of
C-H amination reaction

Novel Iminium Ion Equivalents

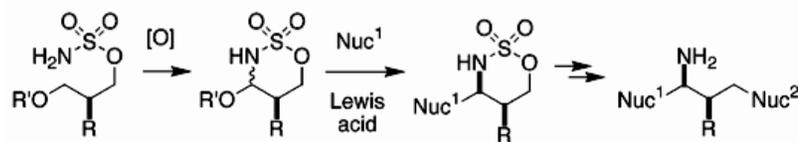
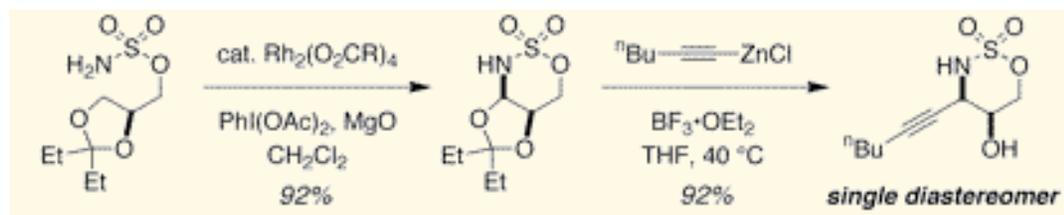


Figure 1. Novel *N,O*-acetal oxathiazinane heterocycles as reactive iminium ion equivalents.



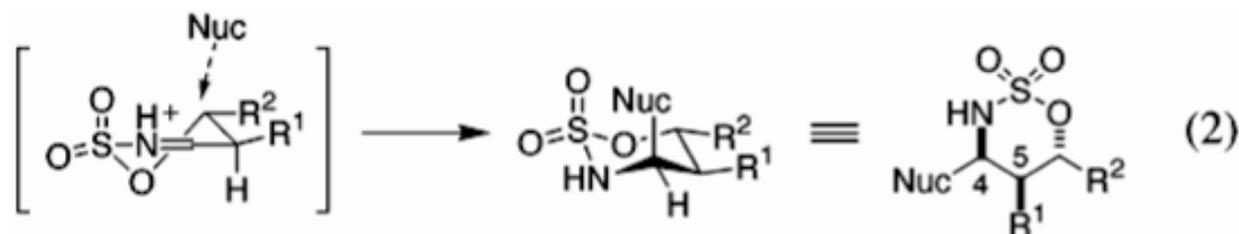
Entry	Substrate	Major isomer ^a	Selectivity ^b	Yield ^c
1			20:1	85
2			20:1	82
3			12:1	71

Novel Iminium Ion Equivalents Cont'd

Table 1. Alkynylzinc Addition Reactions with *N,O*-Acetals

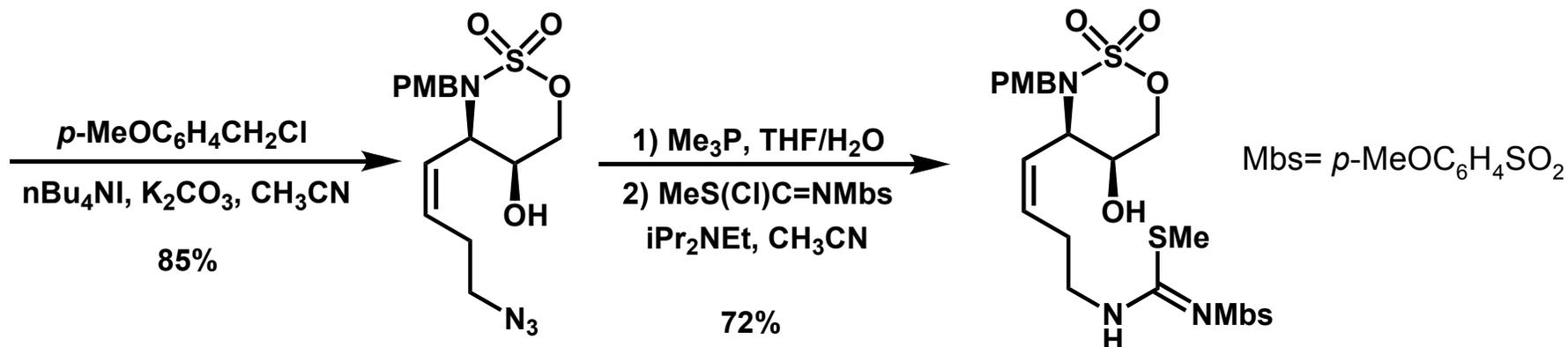
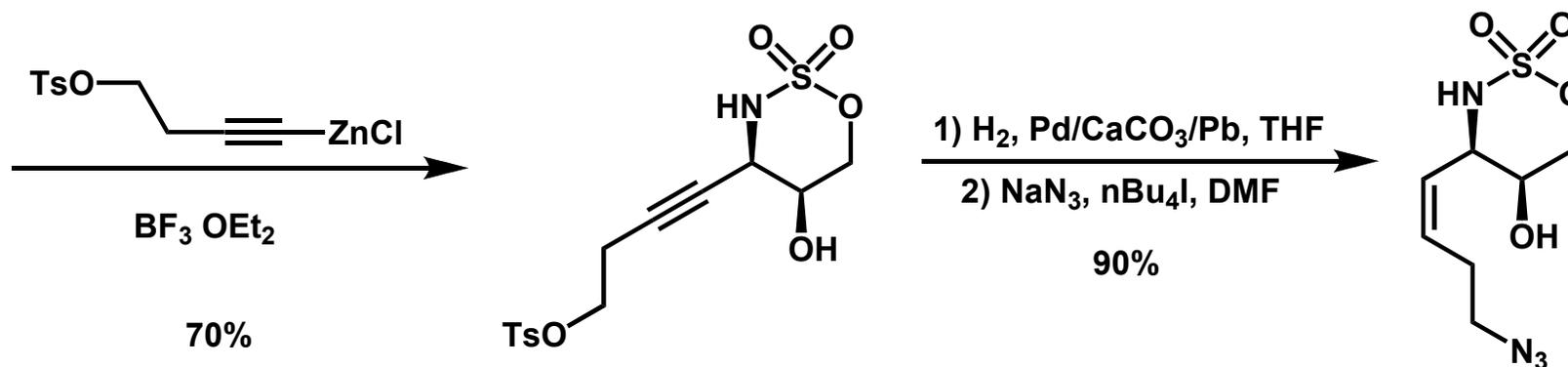
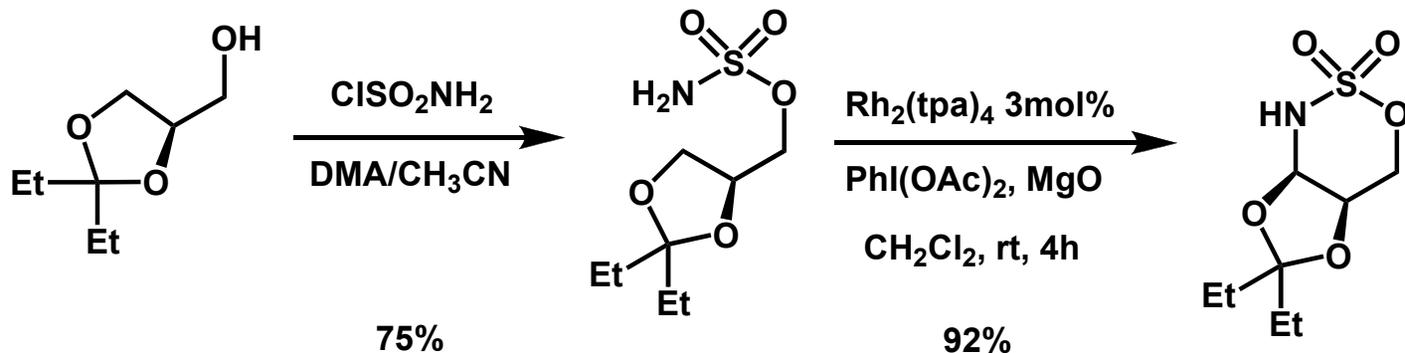
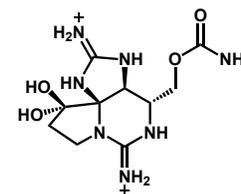
Entry	Substrate	Major isomer ^a	Selectivity ^b	Yield ^c
4			8:1	70
5			20:1	76
6			R = Et 6:1 = OP 6:1	80 63 ^d

Proposed TS model

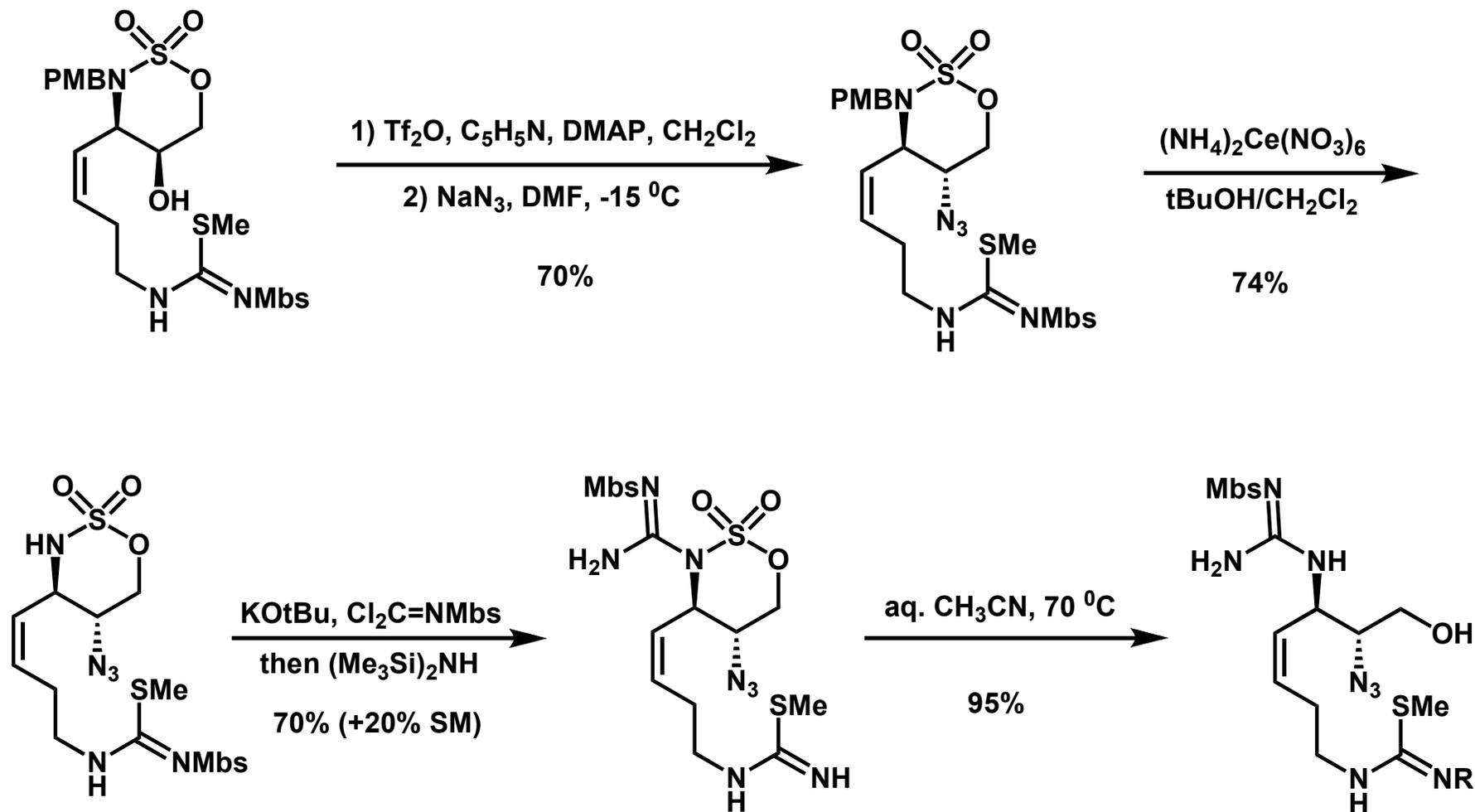
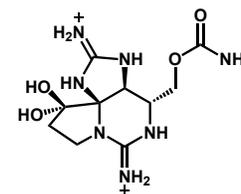


Oxathiazinane heterocycles are easily prepared chiral building blocks

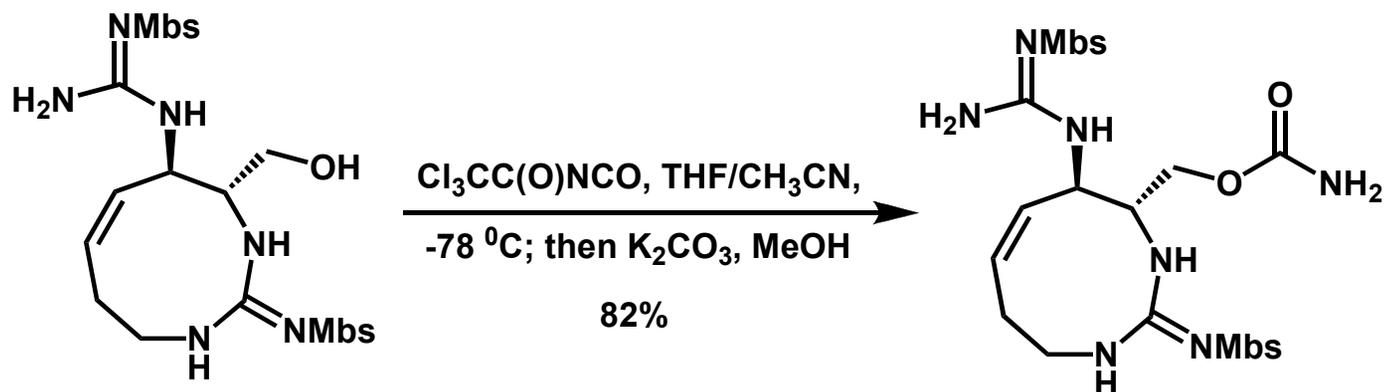
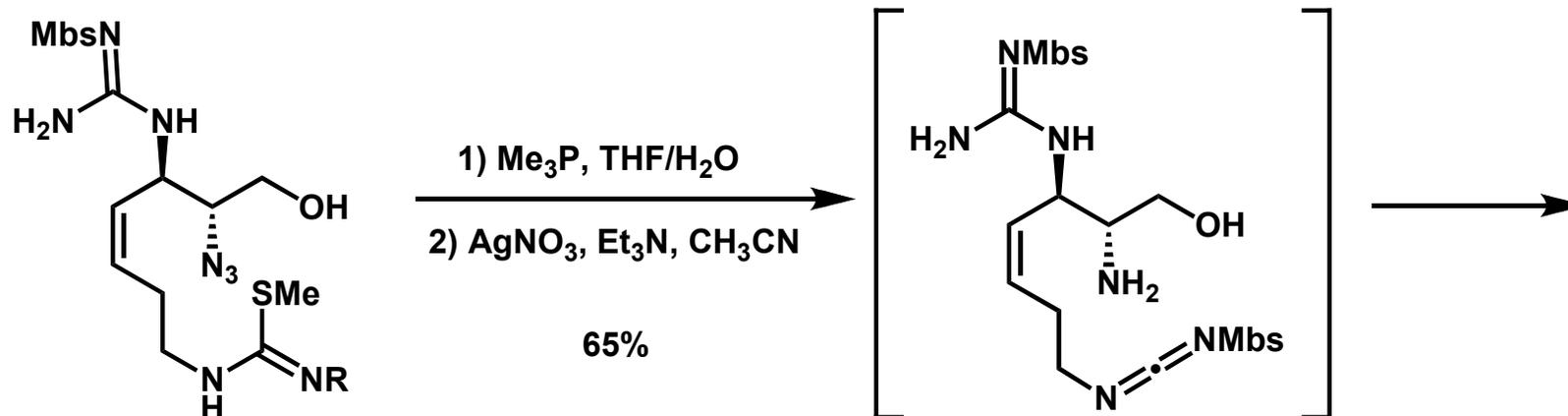
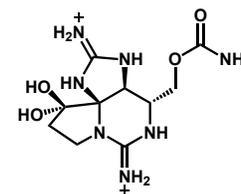
Preparation of Oxathiazinane



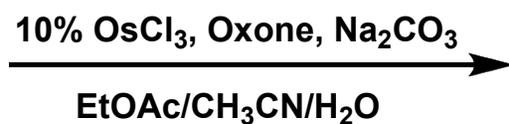
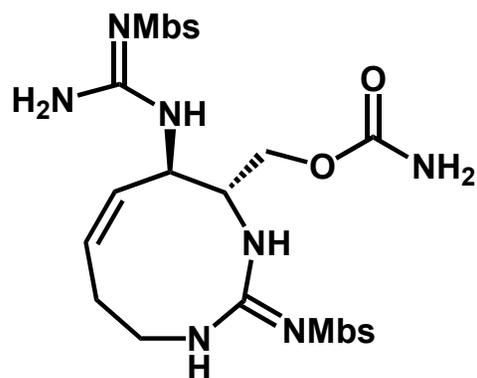
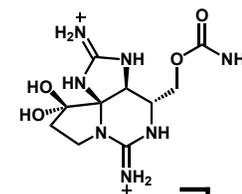
Preparation of Elaborated Azide



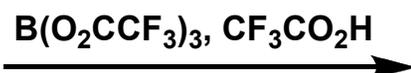
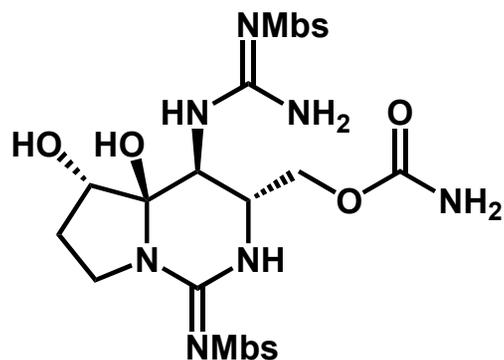
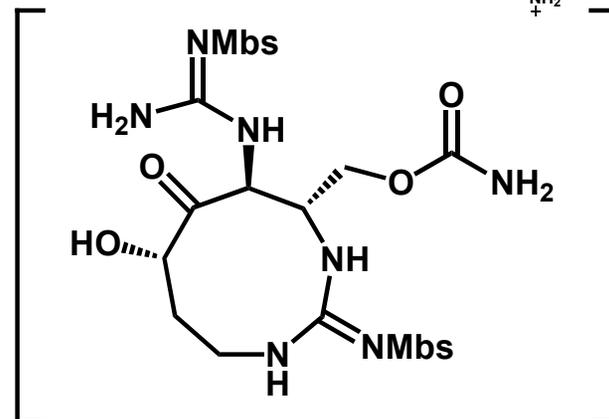
Preparation of 9-membered Ring



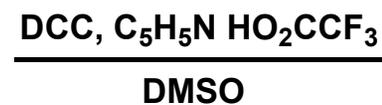
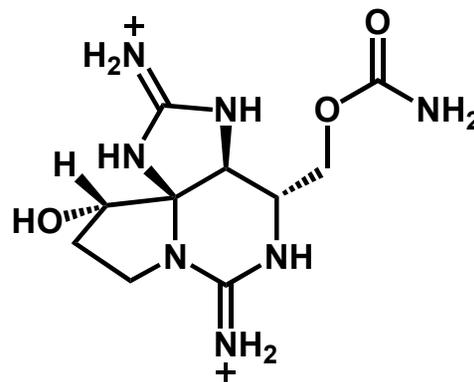
Completion of the Synthesis



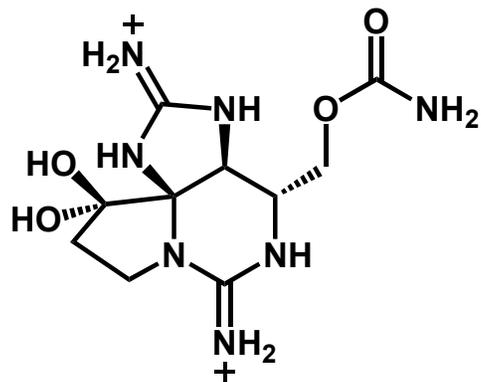
57%



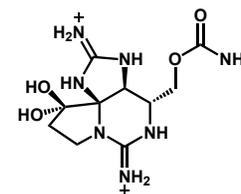
82%



70%



Du Bois Synthesis- Summary



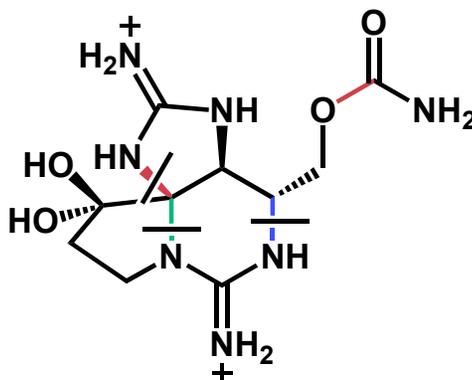
First total synthesis of (+) saxitoxin

Utilized C-H amination and oxathiazinane iminium ion equivalent developed within the Du Bois labs

Key strategies relied upon stereocontrolled formation of the 9-membered ring and condensation to prepare the bicyclic guanidinium core

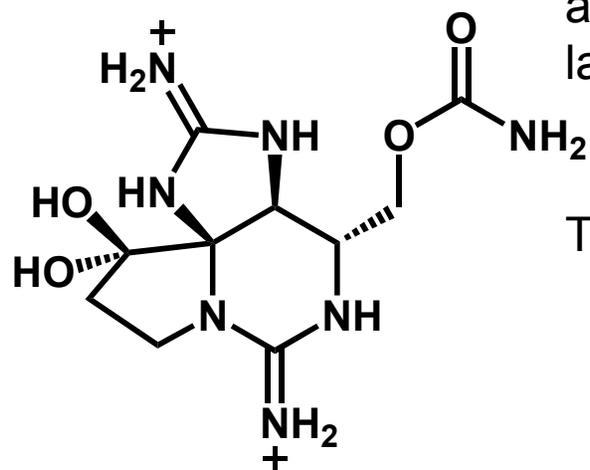
19 steps from commercially available (R) glycerol acetamide

1.6% overall yield



Conclusion

Three syntheses within the past 29 years;
although first non-racemic synthesis published
last month.

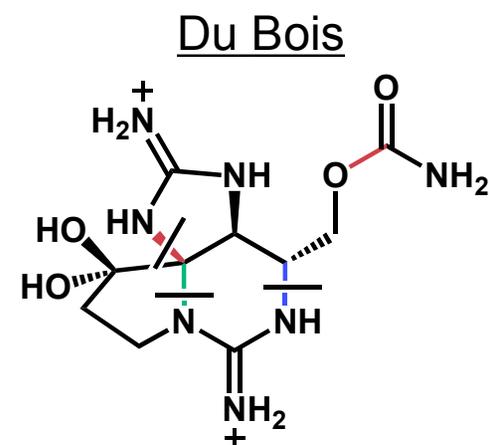
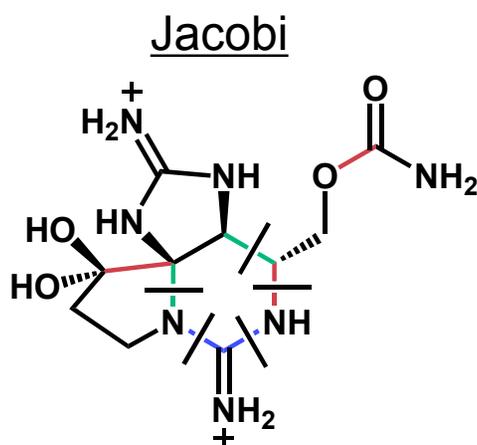
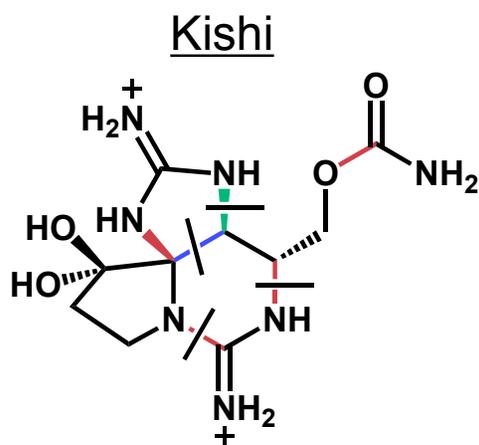


Three very different approaches to the core:

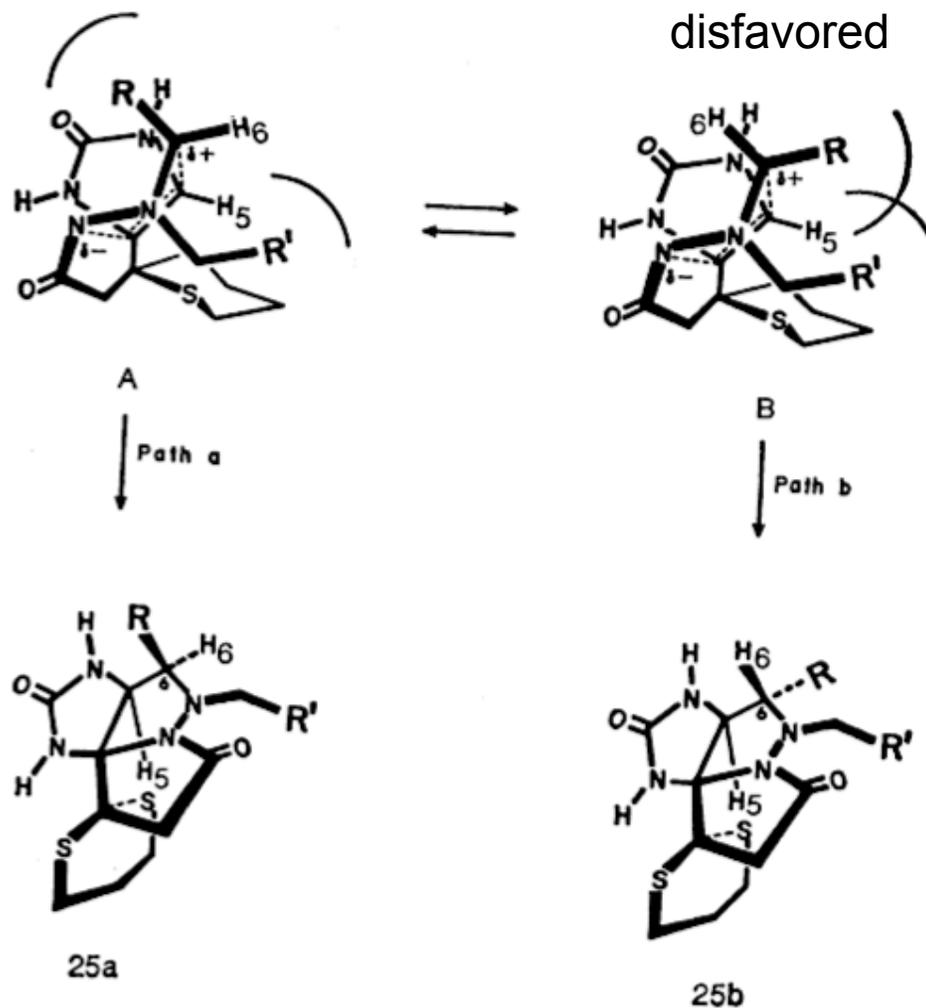
Kishi- build each cyclic guanidine upon pyrrolidine

Jacobi- [3+2] cycloaddition

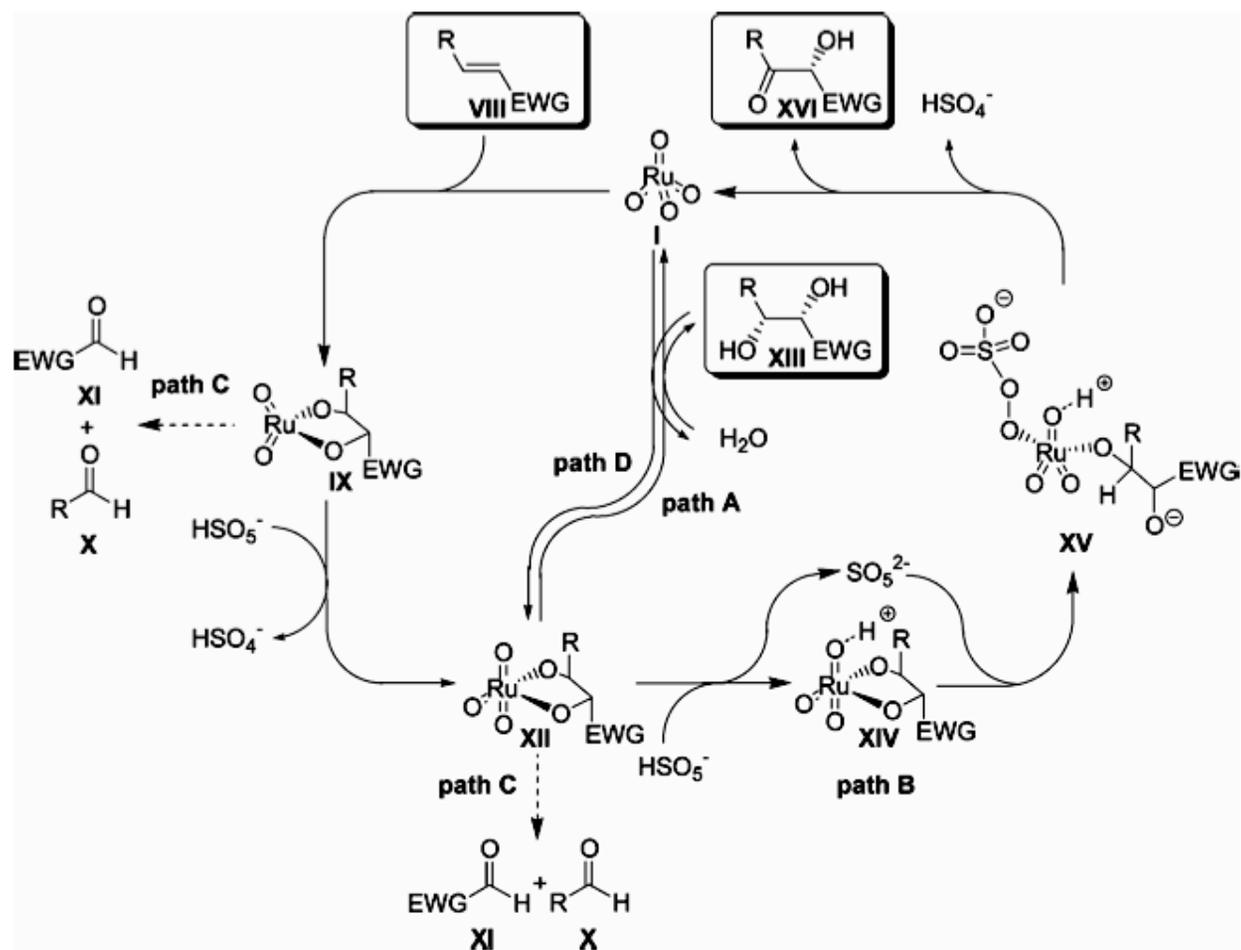
Du Bois- condensation of 9-membered ring



3+2 cycloaddition selectivity



Keto hydroxylation Reaction

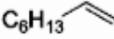
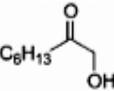
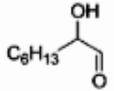
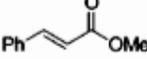
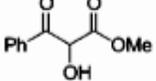
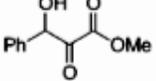
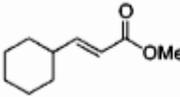
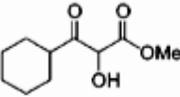
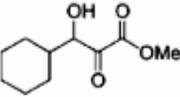
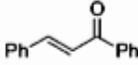
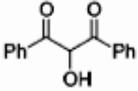
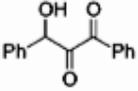
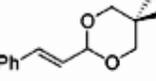
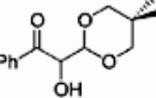
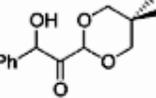
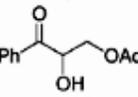
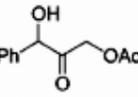


Plietker, JOC, 2004, 8287

Keto hydroxylation cont'd

TABLE 7. Ketohydroxylation of Olefins: Functional Group Tolerance

$$\text{R}^1\text{-CH=CH-R}^2 \xrightarrow[\text{rt}]{\begin{array}{l} \text{RuCl}_3 [1 \text{ mol}\%] \\ \text{Oxone} [5 \text{ equiv.}] \\ \text{NaHCO}_3 [2.5 \text{ equiv.}] \\ \text{EtOAc/CH}_3\text{CN/H}_2\text{O} (6:6:1) \end{array}} \text{R}^1\text{-CH(OH)-C(=O)-R}^2 + \text{R}^1\text{-C(=O)-CH(OH)-R}^2$$

entry ^a	alkene	acyloin ^b	yield[%] ^c	
4	 9	 10a (96%)	 10b (4%)	64
5	 11	 12a (92%)	 12b (8%)	76
6	 13	 14a (96%)	 14b (4%)	82
7	 15	 16a (97%)	 16b (3%)	54
8	 17	 18a (94%)	 18b (6%)	64
9	 19	 20a (87%)	 20b (13%)	68

Proposed Biosynthesis

Scheme 1

